

Summary Report

for the ISO 16140-2 validation of
Assurance® GDS *Listeria monocytogenes* Tq
for the detection of *Listeria monocytogenes* in a broad
range of foods and environmental samples

MicroVal study number	2014LR32
Method/Kit names	Assurance® GDS <i>Listeria monocytogenes</i> Tq
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LIST OF ABBREVIATIONS LISTERIA

Method & protocol

ALOA or O&A	Agar <i>Listeria</i> Ottavani & Agosti
BHI	Brain Heart Infusion Broth
CFU	Colony Forming Units
g	gram
h	hour
HFB	Half Fraser broth
ILS	Interlaboratory Study
MCS	Method Comparison Study
min	Minute
ml	Millilitre
RLOD	Relative Level of Detection
TSA/YE	Tryptone Soya Agar with Yeast
UHT	Ultra Heat Treated

Interpretation

AL	Acceptability Limit
alt	Alternative method
\bar{D}	Average difference
FN	False Negative results
FNR	False Negative Ratio
FP	False Positive
FPR	False Positive Ratio
LOD	Limit of Detection
NA	Negative agreement
NA _{FN (alt)}	Negative Agreement due to false negative alternative-method results
ND	Negative Deviation
ND _{FN (alt)}	Negative Deviation due to false negative alternative-method results
PA	Positive Agreement
PA _{FP (alt)}	Positive Agreement due to false positive alternative-method results
PD	Positive deviation
PD _{FP (alt)}	Positive Deviation due to false positive alternative-method results
ref	Reference method
TNA	Total Negative Agreement
TND	Total Negative Deviation

Raw data

- or neg(-)	Negative/no growth/no reaction/target not detected
(x)	Number of colonies in the plate
H+	Characteristic <i>Listeria</i> colonies with halo
H-	Characteristic <i>Listeria</i> without halo
i	Inhibition
<i>L. mono</i>	<i>L. monocytogenes</i>
<i>L. innoc</i>	<i>L. innocua</i>
<i>L. welsh</i>	<i>L. welshimeri</i>
<i>L. seeli</i>	<i>L. seeligeri</i>
na	Not applicable
ne	New DNA extraction
NC or NS	Non-characteristic colony
/ or nt	Not tested
st	Plate without any colony
Bold typing	Artificially inoculated samples

Foreword

The technical protocol and the result interpretation were carried out according to the ISO 16140-2:2016, ISO 16140-2/A1:2024 and the MicroVal technical rules. This document is prepared in accordance with ISO 16140-2 and MicroVal technical committee interpretation of ISO 16140-2 v.2.6.

Validation protocol	<ul style="list-style-type: none"> ▪ ISO 16140-1:2016 - Microbiology of the food chain - Method validation — <i>Part 1: Vocabulary</i> ▪ ISO 16140-2:2016 & ISO 16140-2/A1:2024 - Microbiology of the food chain - Method validation — <i>Part 2: Protocol for the validation of alternative (proprietary) methods against a reference method</i> ▪ MicroVal technical committee interpretation of ISO 16140-2 v.2.6
Reference method ♦	ISO 11290-1:2017 - Microbiology of the food chain - Horizontal method for the detection and enumeration of <i>Listeria monocytogenes</i> and other <i>Listeria</i> spp. - Part 1: detection method
Method/Kit name	Assurance[®] GDS <i>Listeria monocytogenes</i> Tq
Scope of validation	<ul style="list-style-type: none"> > Broad range of foods > Environmental samples
Certification organization	Lloyd's Register

♦ Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

1 INTRODUCTION

This document reports a MicroVal validation study, based on ISO 16140-2:2016 and ISO 16140-2/A1:2024, of the Assurance® GDS *Listeria monocytogenes* Tq, alternative method for the detection of *Listeria* species in a broad range of foods and environmental samples. The initial evaluation was carried out by ADRIA in 2014 as the MicroVal Expert Laboratory according to ISO 16140:2003.

In 2018, a renewal and an extension for the addition of Fresh produce and vegetable category was carried out according to ISO 16140-2:2016 by QLaboratories. In 2025, the exclusivity was completed with new *Listeria* spp. strain, and the results were reanalysed according to the ISO 16140-2/A1:2024.

This renewal was carried out by ADRIA.

This report combines all data from the initial validation and extension re-evaluated following ISO 16140-2/A1:2024 criteria.

The Assurance® GDS *Listeria monocytogenes* Tq is an automated nucleic acid amplification system for the detection of *Listeria* species in a broad range of foods and environmental samples.

The reference method is: ISO 11290-1:2017 - Microbiology of the food chain - Horizontal method for the detection and enumeration of *Listeria monocytogenes* and other *Listeria* spp. - Part 1: detection method.

Scope of the validation study is: broad range of foods, and environmental samples.

Categories included:

- Meat products
- Milk and dairy products
- Fish and seafood products
- Fresh produce and vegetables
- Ready-to-eat, ready-to-reheat or ready-to-cook products
- Environmental samples

Criteria to be evaluated during the study:

- Method Comparison Study (MCS)
 - Sensitivity study
 - Relative level of detection
 - Inclusivity and exclusivity study
- Interlaboratory Study (ILS).

2 METHOD PROTOCOLS

The Method Comparison Study was carried out using 25 g portions of sample material. For environmental samples, testing was conducted using environmental sponges or 25 g test portions.

2.1 Reference method[♦]

The reference method are the ISO 11290-1:1996/Amd.1:2004 for the initial validation study and the ISO 11290-1 (May 2017): Microbiology of the food chain - Horizontal method for the detection and enumeration of *Listeria monocytogenes* and of *Listeria* spp. - Part 1: detection method for the fresh Produce and Vegetables food types extension study.

See the flow diagram in **Appendix 1**.

Sample preparations used in the reference method and Assurance GDS *Listeria* **species** method was performed according to ISO 6887 – 4, ISO 11290-1:1996/Amd.1:2004 and ISO 11290-1:2017.

25 g test portion were used for the reference method.

2.2 Alternative method

See the flow diagram of the alternative method in **Appendix 2** and the kit insert in **Appendix 3**.

2.2.1 Principle

The alternative method principle is based on PCR.

The Assurance® GDS *Listeria monocytogenes* is an automated nucleic acid amplification system for the detection of *Listeria monocytogenes* in a broad range of foods and environmental samples.

The food samples were prepared for analysis following an IMS concentration procedure as outlined in the Assurance GDS *Listeria monocytogenes* package insert

[♦] Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

and previous MicroVal Validation (MicroVal 2010LR32). Results are available after 26 - 44 h of enrichment.

The alternative method used is:

- Twenty-five (25) gram samples are enriched with 225 mL of Half Fraser Broth (HFB) and environmental sponges are enriched in 90 mL of Half Fraser Broth.
- Test portions are incubated at $30 \pm 1^\circ\text{C}$ for 24 ± 2 hours.
- Following enrichment, the sample is concentrated using the Assurance PickPen[®] device and then transferred into 0.5 mL of Half Fraser Broth.
- The HFB is sealed and incubated at $30 \pm 1^\circ\text{C}$ for 4 to 22 hours for all categories except for the meat products category where only a 22h incubation time is possible.
- Following incubation, the sample is concentrated using the PickPen device, lysed and a 30 μL aliquot of lysate is analyzed using the Assurance GDS Rotor-Gene[®] instrument.
- Confirmations following the ISO 11290-1:2017

It is possible to store the enriched samples for 72 h at $5 \pm 3^\circ\text{C}$ before performing the IMS step and confirmatory tests.

2.2.2 *Protocols applied during the validation study*

> **Enrichment**

The minimum incubation time was applied for the enrichment step: 22 h at $30 \pm 1^\circ\text{C}$.

> **Subculture**

For all categories except meat products category: the IMS and PCR were carried out after the incubation of HFB for 4 and 22 h at $30 \pm 1^\circ\text{C}$.

For meat product categories: the IMS and PCR were carried out after the incubation of HFB for 22 h at $30 \pm 1^\circ\text{C}$.

> **Confirmations**

For all samples, 10µL of HFB was streaked on O&A and palcam plates. Confirmations of ISO 11290-1:2017 were carried out on typical colonies.

> **Broth storage for 72h at 5 ± 3°C**

The enrichment broths from positive and discordant samples were tested after 72 h storage at 5 ± 3°C (PCR and confirmatory tests for unpaired with MICROGEN latex test to confirm the colonies and PCR with direct streaking only for paired samples).

2.3 Study design

For the 25 g portions, the reference and the alternative method share the initial (pre)-enrichment step, therefore the same test portion (item) was used for the two methods. All resulting data were treated as paired data.

3 METHOD COMPARISON STUDY

3.1 Sensitivity study

The sensitivity (SE) is the ability of the method to detect the analyte by either the reference or alternative method.

3.1.3 Categories and sample types

Five food categories and environmental samples category were included in this validation study. A minimum of 60 items for each category were tested by both the reference method and the alternative method in the sensitivity study, with a minimum of 30 positive samples per category.

Each category was made up of 3 types, with at least 20 items representative for that Type.

400 samples were analysed. The distribution of positive and negative samples per tested category and type is given respectively in Table 1.

Table 1 – Distribution per tested category and type

Categories	Types	Sample size	Positive sample	Negative samples	Total
Meat products	a Fresh meat (unprocessed)	25 g	11	11	22
	b Cooked and cured meat products	25 g	11	18	29
	c Fermented meat products	25 g	10	14	24
	<i>Total</i>		32	43	75
Milk and dairy products	a Thermisation/pasteurized products	25 g	12	8	20
	b Fermented/acidified products	25 g	5	13	18
	c Raw milk-based products	25 g	14	12	26
	<i>Total</i>		31	33	64
Fish and seafood products	a Fish and shellfish	25 g	12	6	18
	b Cooked fishery products	25 g	8	11	19
	c Smoked products	25 g	14	13	27
	<i>Total</i>		34	30	64
Ready-to-eat, ready-to-reheat or ready-to-cook products	a Substantial raw ingredients	25 g	11	6	17
	b Processed (cooked) products	25 g	7	9	16
	c Refrigerated products	25 g	13	16	29
	<i>Total</i>		31	31	62
Environmental samples	a Swabs and sponges	25 g	9	13	22
	b Process water	25 g	11	9	20
	c Dusts	25 g	12	15	27
	<i>Total</i>		32	37	69
Fresh produce and vegetables	a Cut-ready-to-eat vegetables	25 g	11	11	22
	b Produce grown in contact with the ground	25 g	11	11	22
	c Cut ready-to-eat fruit	25 g	11	11	22
	<i>Total</i>		33	33	66
ALL CATEGORIES			193	207	400

3.1.4 Test sample preparation

Naturally contaminated samples were preferentially analyzed. If necessary, artificial contaminations were obtained by spiking or seeding protocols.

When spiking the strains were stressed using various injury protocols. The injury efficiency was evaluated by comparing enumeration results onto selective and non-selective agars (respectively O&A and TSYEA plates). The artificial contaminations are presented in **Appendix 4**.

For all studies, 126 samples were artificially contaminated, using 41 different strains with seeding or spiking protocols. 109 gave a positive result.

A total of 84 samples were found naturally contaminated samples

The repartition of the positive samples per inoculation protocol and inoculation level is given in **Erreur ! Source du renvoi introuvable.**

Table 2 - Repartition of the positive samples per inoculation protocol and inoculation level

Study	Naturally contaminated	Artificial contamination						Total
		Seeding protocol			Spiking protocol			
		≤ 3 CFU	3 < x ≤ 7.2 CFU	> 10 CFU	≤ 5 CFU	5 < x ≤ 6.8 CFU	> 10 CFU	
Initial validation study	81	4	28	0	26	21	0	160
Fresh produce and vegetables Extension study	3	0	30	0	0	0	0	33
Total	84	4	58	0	26	21	0	193
%	43.5%	2.1%	30.1%	0.0%	13.5%	10.9%	0.0%	98.0%

43.5 % of the samples were naturally contaminated.

3.1.5 Sensitivity study results

All raw data per category are given in **Appendix 5**. Sample numbers in **bold** indicate artificial inoculation of the sample (see **Appendix 4** for details on artificial inoculation).

Table 3 shows the summary of results of the reference method and the alternative methods for **all Categories**.

Table 4 shows the Interpretation of sample results between the reference and alternative method (based on the confirmed alternative method).

Table 3 – Summary of sensitivity study results – all categories

Response	22 + 4 h		22 + 22 h	
	Reference method positive (R+)	Reference method negative (R-)	Reference method positive (R+)	Reference method negative (R-)
Alternative method positive (A+)	Positive agreement (A+/R+) PA = 158	Positive deviation (R-/A+) PD = 0	Positive agreement (A+/R+) PA = 189	Positive deviation (R-/A+) PD = 2
Alternative method negative (A-)	Total Negative deviation (A-/R+) TND = 2 (2 ND_{FN(alt)})	Total Negative agreement (A-/R-) TNA = 152 (4 NA_{FN(alt)})	Total Negative deviation (A-/R+) TND = 2 (2 ND_{FN(alt)})	Total Negative agreement (A-/R-) TNA = 194 (3 NA_{FN(alt)})

Table 4 - Summary of results obtained with the reference and alternative methods (after confirmation) of all samples for each category

Category	4 h					22 h				
	PA	PD	TND	TNA	Total	PA	PD	TND	TNA	Total
1 Meat products						29	0	2	43	75
2 Milk and dairy products	30	0	0	34	64	30	0	0	33	64
3 Fish and Seafood products	34	0	0	17	64	34	0	0	17	64
4 Ready-to-eat, ready-to-reheat or ready-to-cook products	30	0	1	31	62	31	0	0	31	62
5 Production environmental samples	31	0	1	37	69	32	0	0	37	69
6 Fresh produce and vegetables	33	0	0	33	66	33	0	0	33	66
TOTAL	158	0	2	152	325	189	0	2	194	400

With for a paired evaluation : $TND = ND_{FN(alt)}$ $TNA = NA + PD_{FP(alt)}$

3.1.6 Sensitivity study calculations

The sensitivity study parameters as specified in Table 5 were calculated for all categories and types, and the overview is given in Table 6.

Table 5 – Formula to calculate the sensitivity parameters

		22 + 4 h (except meat products)	22 + 22 h
Sensitivity for the alternative method	$SE_{alt} = \frac{(PA + PD)}{(PA + TND + PD)} \times 100 \%$	98.8 %	99.0 %
Sensitivity for the reference method	$SE_{ref} = \frac{(PA + TND)}{(PA + TND + PD)} \times 100 \%$	100.0 %	99. %
Relative trueness	$RT = \frac{(PA + TNA)}{N} \times 100 \%$	95.4 %	95.8 %
False positive ratio for the alternative method (paired evaluation)	$FPR = \frac{PD_{FP(alt)}}{TNA} \times 100 \%$	0.0 %	0.5 %
False negative ratio for the alternative method (paired evaluation)	$FNR = \frac{ND_{FN(alt)}}{PA+TND+PD} \times 100 \%$	3.8 %	2.6 %



Table 6 – Overview calculated sensitivity parameters per category and type

Category	Incubation time	Type	PA	PA _{FP(alt)}	NA	NA _{FN(alt)}	PD	ND	ND _{FN(alt)}	PD _{FP(alt)}	TND	TNA	SE _{alt} %	SE _{ref} %	RT %	FP %	FNR %
1	22 + 22h	a Fresh meat (unprocessed)	11	0	10	0	0	0	0	1	0	11	100.0	100.0	100.0	0.0	0.0
		b Cooked and cured meat products	11	0	18	0	0	0	0	0	0	18	100.0	100.0	100.0	0.0	0.0
		c Fermented meat products	7	0	14	0	1	0	2	0	2	14	80.0	90.0	87.5	0.0	20.0
		Total	29	0	42	0	1	0	2	1	2	43	93.8	96.9	96.0	2.3	6.3
2	22 + 4h	a Thermisation/pasteurized products	11	0	8	1	0	0	0	0	0	9	100.0	100.0	100.0	0.0	9.1
		b Fermented/acidified products	5	0	13	0	0	0	0	0	0	13	100.0	100.0	100.0	0.0	0.0
		c Raw milk-based products	14	0	12	0	0	0	0	0	0	12	100.0	100.0	100.0	0.0	0.0
		Total	30	0	33	1	0	0	0	0	0	34	100.0	100.0	100.0	0.0	3.3
	22 + 22h	a Thermisation/pasteurized products	11	0	8	0	1	0	0	0	0	8	100.0	91.7	95.0	0.0	0.0
		b Fermented/acidified products	5	0	13	0	0	0	0	0	0	13	100.0	100.0	100.0	0.0	0.0
		c Raw milk-based products	14	0	12	0	0	0	0	0	0	12	100.0	100.0	100.0	0.0	0.0
		Total	30	0	33	0	1	0	0	0	0	33	100.0	96.8	98.4	0.0	0.0
3	22 + 4h	a Fish and shellfish	12	0	6	0	0	0	0	0	0	6	100.0	100.0	100.0	0.0	0.0
		b Cooked fishery products	8	0	11	0	0	0	0	0	0	11	100.0	100.0	100.0	0.0	0.0
		c Smoked products	14	0	13	0	0	0	0	0	0	13	100.0	100.0	100.0	0.0	0.0
		Total	34	0	30	0	0	0	0	0	0	17	100.0	100.0	79.7	0.0	0.0
	22 + 22h	a Fish and shellfish	12	0	6	0	0	0	0	0	0	6	100.0	100.0	100.0	0.0	0.0
		b Cooked fishery products	8	0	11	0	0	0	0	0	0	11	100.0	100.0	100.0	0.0	0.0
		c Smoked products	14	0	13	0	0	0	0	0	0	13	100.0	100.0	100.0	0.0	0.0
		Total	34	0	30	0	0	0	0	0	0	17	100.0	100.0	79.7	0.0	0.0



Category	Incubation time	Type	PA	PA _{FP(alt)}	NA	NA _{FN(alt)}	PD	ND	ND _{FN(alt)}	PD _{FP(alt)}	TND	TNA	SE _{alt} %	SE _{ref} %	RT %	FP %	FNR %
4	22 + 4h	a Substantial raw ingredients	11	0	4	2	0	0	0	0	0	6	100.0	100.0	100.0	0.0	18.2
		b Processed (cooked) products	7	0	8	1	0	0	0	0	0	9	100.0	100.0	100.0	0.0	14.3
		c Refrigerated products	12	0	16	0	0	0	1	0	1	16	92.3	100.0	96.6	0.0	7.7
		Total	30	0	28	3	0	0	1	0	1	31	96.8	100.0	98.4	0.0	12.9
	22 + 22h	a Substantial raw ingredients	11	0	4	2	0	0	0	0	0	6	100.0	100.0	100.0	0.0	18.2
		b Processed (cooked) products	7	0	8	1	0	0	0	0	0	9	100.0	100.0	100.0	0.0	14.3
		c Refrigerated products	13	0	16	0	0	0	0	0	0	16	100.0	100.0	100.0	0.0	0.0
		Total	31	0	28	3	0	0	0	0	0	31	100.0	100.0	100.0	0.0	9.7
5	22 + 4h	a Swabs and sponges	8	0	13	0	0	0	1	0	1	13	88.9	100.0	95.5	0.0	11.1
		b Process water	11	0	9	0	0	0	0	0	0	9	100.0	100.0	100.0	0.0	0.0
		c Dusts	12	0	15	0	0	0	0	0	0	15	100.0	100.0	100.0	0.0	0.0
		Total	31	0	37	0	0	0	1	0	1	37	96.9	100.0	98.6	0.0	3.1
	22 + 22h	a Swabs and sponges	9	0	13	0	0	0	0	0	0	13	100.0	100.0	100.0	0.0	0.0
		b Process water	11	0	9	0	0	0	0	0	0	9	100.0	100.0	100.0	0.0	0.0
		c Dusts	12	0	15	0	0	0	0	0	0	15	100.0	100.0	100.0	0.0	0.0
		Total	32	0	37	0	0	0	0	0	0	37	100.0	100.0	100.0	0.0	0.0
6	22 + 4h	a Cut-ready-to-eat vegetables	11	0	11	0	0	0	0	0	0	11	100.0	100.0	100.0	0.0	0.0
		b Produce grown in contact with the ground	11	0	11	0	0	0	0	0	0	11	100.0	100.0	100.0	0.0	0.0
		c Cut ready-to-eat fruit	11	0	11	0	0	0	0	0	0	11	100.0	100.0	100.0	0.0	0.0
		Total	33	0	33	0	0	0	0	0	0	33	100.0	100.0	100.0	0.0	0.0
	22 + 22h	a Cut-ready-to-eat vegetables	11	0	11	0	0	0	0	0	0	11	100.0	100.0	100.0	0.0	0.0
		b Produce grown in contact with the ground	11	0	11	0	0	0	0	0	0	11	100.0	100.0	100.0	0.0	0.0
		c Cut ready-to-eat fruit	11	0	11	0	0	0	0	0	0	11	100.0	100.0	100.0	0.0	0.0
		Total	33	0	33	0	0	0	0	0	0	33	100.0	100.0	100.0	0.0	0.0
Total short incubation time (4h) - all categories except meat			158	0	161	4	0	0	2	0	2	152	98.8	100.0	95.4	0.0	3.8
Total long incubation time (22h) - all categories			189	0	204	3	2	0	2	0	2	194	99.0	99.0	95.8	0.5	2.6

3.1.7 Discordant results

Negative deviations are listed in Table 7 and positive deviations in Table 8.

> **Negative deviations**

Two negative deviations were observed for both HFB subculture incubation time (4 and 22h). Negative PCR result was obtained while confirmation gave positive result after subculture in Fraser (ISO 11290-1 method) or after 72h storage at $5\pm 3^{\circ}\text{C}$ (typical colonies on ALOA and Palcam plates). For only one sample (n°6834), few typical colonies were obtained with the direct streaking on O&A and Palcam plates.

For the other samples, before storage, no typical colony was obtained on plates which may indicate a low level of contamination. The level of detection was probably not reached by the alternative method.

For samples in negative agreement with the reference method, four samples (after 4h subculture in HFB) and three samples (after 22h subculture in HFB) gave positive confirmation while negative PCR result was obtained: $\text{NA}_{\text{FN(alt)}}$ samples. See table 9. It can be noted that positive confirmation result was obtained only after storage for samples n°4626 and 4771.

> **Positive deviations**

Two positive deviations were observed after 22h subculture in HFB. One sample was artificially contaminated and one naturally contaminated. See Table 8.

Table 7 - Negative deviations

N°Sample	Product	ISO 11290-1/A1 method <i>L. monocytogenes</i> result	GDS <i>Listeria monocytogenes</i> method								Category	Type	
			Half Fraser 22h 30°C + 4 or 22h subculture										
			PCR Result 4H 30°C	PCR Result 22H 30°C	Confirmations		Final confirmation	Final result 4 h	Final result 22 h	Agreement Ref/Alt 4 h			Agreement Ref/Alt 22 h
ALOA	Palcam												
3344	Merguez	+		-	H-	+	- (+ ref)		-		ND _{FN(alt)}	1	c
5242	Sliced dehydrated sausage	+		-	-	st	- (+ ref)		-		ND _{FN(alt)}	1	c
4624	Spinach	+	-	+	-	-	- (+ ref and 72h)	-	+	ND _{FN(alt)}	PA	4	c
6834	Wipe (fish industry materials)	+	-	+	2H+	5+	+	-	+	ND _{FN(alt)}	PA	5	a

ref: reference method

Table 8 - Positive deviations

N°Sample	Product	ISO 11290-1/A1 method <i>L. monocytogenes</i> result	GDS <i>Listeria monocytogenes</i> method								Category	Type	
			HFB 22h 30 ± 1°C with 4 or 22h subculture										
			PCR Result 4H 30°C	PCR Result 22H 30°C	Confirmations		Final confirmation	Final result 4 h	Final result 22 h	Agreement Ref/Alt 4 h			Agreement Ref/Alt 22 h
ALOA	Palcam												
4771	Pasteurized fresh cream	-	-	+	st	st	- (+ at 72h)	-	+	NA _{FN(alt)}	PD	2	a
5244	Dehydrated sausage	-	-	+	H-	st	- (+ at 72h)	-	+	NA _{FN(alt)}	PD	1	c



Table 9 – NA_{FN(alt)} samples

N°Sample	Product	ISO 11290-1/A1 method <i>L. mono</i> result	GDS Listeria monocytogenes method												Category	Type
			Before storage						After storage 72h 5 ± 3°C							
			PCR Result 4h 30°C	PCR Result 22h 30°C	Confirmations		Agreement Ref/Alt		PCR Result		Confirmations		Agreement Ref/Alt			
					ALOA	Palcam	4 h	22 h	4 h 30°C	22 h 30°C	ALOA	Palcam	4 h	22 h		
4626	Paella	-	-	-	1H- (+ at 72h)	-	NA _{FN(alt)}	NA _{FN(alt)}	-	-	H- /1H+	+	NA _{FN(alt)}	NA _{FN(alt)}	4	b
4771	Pasteurized fresh cream	-	-	+	st (+ at 72h)	st	NA _{FN(alt)}	PD	-	+	4H+	+	NA _{FN(alt)}	PD	2	a
5179	Dressing (sorrel)	-	-	-	H+	+	NA _{FN(alt)}	NA _{FN(alt)}							4	a
5180	Dressing (green pepper)	-	-	-	H+	+	NA _{FN(alt)}	NA _{FN(alt)}							4	a

The analyses of discordant results according to the ISO 16140-2:2016 and ISO 16140-2/A1:2024 for a paired study is the following (See Table 10):

Table 10 - Interpretation of the sensitivity study results

Category	Type	N+	TND	PD	Paired				
					TND-PD	AL	TND+PD	AL	
1 Meat products 22 + 22h	a Fresh meat (unprocessed)	11	0	0	0	3	0	3	
	b Cooked and cured meat products	11	0	0	0		0		
	c Fermented meat products	10	2	1	1		3		
	Total	32	2	1	1	3	3	3	
2 Milk and dairy products 22 + 4h	a Thermisation/pasteurized products	11	0	0	0	3	0	3	
	b Fermented/acidified products	5	0	0	0		0		
	c Raw milk-based products	14	0	0	0		0		
	Total	30	0	0	0	3	0	3	
	Milk and dairy products 22 + 22h	a Thermisation/pasteurized products	12	0	1	-1	3	1	3
		b Fermented/acidified products	5	0	0	0		0	
		c Raw milk-based products	14	0	0	0		0	
Total		31	0	1	-1	3	1	3	
3 Fish and seafood products 22 + 4h	a Fish and shellfish	12	0	0	0	3	0	3	
	b Cooked fishery products	8	0	0	0		0		
	c Smoked products	14	0	0	0		0		
	Total	34	0	0	0	3	0	3	
	Fish and seafood products 22 + 22h	a Fish and shellfish	12	0	0	0	3	0	3
		b Cooked fishery products	8	0	0	0		0	
		c Smoked products	14	0	0	0		0	
Total		34	0	0	0	3	0	3	
4 Ready-to-eat, ready-to-reheat or ready-to-cook products 22 + 4h	a Substantial raw ingredients	11	0	0	0	3	0	3	
	b Processed (cooked) products	7	0	0	0		0		
	c Refrigerated products	13	1	0	1		1		
	Total	31	1	0	1	3	1	3	
	Ready-to-eat, ready-to-reheat or ready-to-cook products 22 + 22h	a Substantial raw ingredients	11	0	0	0	3	0	3
		b Processed (cooked) products	7	0	0	0		0	
		c Refrigerated products	13	0	0	0		0	
Total		31	0	0	0	3	0	3	
5 Production environmental samples 22 + 4h	a Swabs and sponges	9	1	0	1	3	1	3	
	b Process water	11	0	0	0		0		
	c Dusts	12	0	0	0		0		
	Total	32	1	0	1	3	1	3	
	Production environmental samples 22 + 22h	a Swabs and sponges	9	0	0	0	3	0	3
		b Process water	11	0	0	0		0	
		c Dusts	12	0	0	0		0	
Total		32	0	0	0	3	0	3	

Category		Type	N+	TND	PD	Paired			
						TND-PD	AL	TND+PD	AL
6	Fresh produce and vegetables 22 + 4h	a Cut-ready-to-eat vegetables	11	0	0	0	3	0	3
		b Produce grown in contact with the ground	11	0	0	0		0	
		c Cut ready-to-eat fruit	11	0	0	0		0	
		Total	33	0	0	0		0	
	Fresh produce and vegetables 22 + 22h	a Cut-ready-to-eat vegetables	11	0	0	0	3	0	3
		b Produce grown in contact with the ground	11	0	0	0		0	
		c Cut ready-to-eat fruit	11	0	0	0		0	
		Total	33	0	0	0		0	
Total short incubation time (22 + 4h) – all categories except meat			160	2	0	2	6	2	16
Total long incubation time (22 + 22h) – all categories			193	2	2	0	6	4	16

The observed values for (TND – PD) and (TND + PD) for the individual categories and for all categories meet the acceptability limits (observed values ≤ AL).

3.1.8 Half Fraser enrichment storage at 5 ± 3 °C for 72 h

The storage of the subculture in HFB broth was done at 5°C ± 3°C for 72 h.

All positive and discordant samples were tested again after enrichment broth storage. For this study, 199 enriched samples were tested again after storage of the enriched broth for 72h at 5 ± 3°C. Changes were observed for five samples. The following changes were observed (See Table 11).

Table 11 - Observed changes in results before and after storage of the enrichment broth

Sample n°	Before storage		After storage		Category	Type
	4 h	22 h	4 h	22 h		
4624	ND _{FN(alt)}	PA	PA	PA	4	c
4765	PA	PA	ND _{FN(alt)}	PA	2	a
4766	PA	PA	ND _{FN(alt)}	PA	2	a
6843	NA	NA	NA _{FN(alt)}	PD	5	c
6844	NA	NA	PD	NA _{FN(alt)}	5	c

The analyses of discordant become (See Table 12).

**Table 12 - Interpretation of the sensitivity study results
 after storage of the enrichment broth at 5°C ± 3°C**

Category	Type	N+	TND	PD	TND-PD	Paired			
						AL	TND+PD	AL	
1	Meat products 22 + 22h	a	Fresh meat (unprocessed)	11	0	0		0	
		b	Cooked and cured meat products	11	0	0		0	
		c	Fermented meat products	10	2	1		1	
		Total		32	2	1	1	3	3
2	Milk and dairy products 22 + 4h	a	Thermisation/pasteurized products	11	2	0		2	
		b	Fermented/acidified products	5	0	0		0	
		c	Raw milk-based products	13	0	0		0	
		Total		29	2	0	2	3	2
	Milk and dairy products 22 + 22h	a	Thermisation/pasteurized products	12	0	1		1	
		b	Fermented/acidified products	5	0	0		0	
		c	Raw milk-based products	13	0	0		0	
		Total		30	0	1	-1	3	1
3	Fish and seafood products 22 + 4h	a	Fish and shellfish	12	0	0		0	
		b	Cooked fishery products	8	0	0		0	
		c	Smoked products	14	0	0		0	
		Total		34	0	0	0	3	0
	Fish and seafood products 22 + 22h	a	Fish and shellfish	12	0	0		0	
		b	Cooked fishery products	8	0	0		0	
		c	Smoked products	14	0	0		0	
		Total		34	0	0	0	3	0
4	Ready-to-eat, ready-to-reheat or ready-to-cook products 22 + 4h	a	Substantial raw ingredients	11	0	0		0	
		b	Processed (cooked) products	7	0	0		0	
		c	Refrigerated products	13	0	0		0	
		Total		31	0	0	0	3	0
	Ready-to-eat, ready-to-reheat or ready-to-cook products 22 + 22h	a	Substantial raw ingredients	11	0	0		0	
		b	Processed (cooked) products	7	0	0		0	
		c	Refrigerated products	13	0	0		0	
		Total		31	0	0	0	3	0
5	Production environmental samples 22 + 4h	a	Swabs and sponges	9	1	0		1	
		b	Process water	11	0	0		0	
		c	Dusts	13	0	1		-1	
		Total		33	1	1	0	3	2
	Production environmental samples 22 + 22h	a	Swabs and sponges	9	0	0		0	
		b	Process water	11	0	0		0	
		c	Dusts	13	0	1		-1	
		Total		33	0	1	-1	3	1

Category		Type	N+	TND	PD	Paired				
						TND-PD	AL	TND+PD	AL	
6	Fresh produce and vegetables 22 + 4h	a	Cut-ready-to-eat vegetables	12	0	0	0	0		
		b	Produce grown in contact with the ground	12	0	0	0	0		
		c	Cut ready-to-eat fruit	12	0	0	0	0		
		Total			36	0	0	0	3	0
	Fresh produce and vegetables 22 + 22h	a	Cut-ready-to-eat vegetables	12	0	0	0	0	0	
		b	Produce grown in contact with the ground	12	0	0	0	0	0	
		c	Cut ready-to-eat fruit	12	0	0	0	0	0	
		Total			36	0	0	0	3	0
Total short incubation time (22 + 4h) – All categories except meat			163	3	1	2	6	4	16	
Total long incubation time (22 + 22h) – All categories			196	2	3	-1	6	5	16	

The observed values for TND - PD and TND + PD for the individual categories and for all categories meet the acceptability limits (observed values ≤ AL).

3.1.9 Confirmation

It was impossible to confirm the presence of *Listeria monocytogenes* in the enrichment broth after 22 h HFB subculture for 1 samples (PD_{FP(alt)}): n° 3358 (raw ground beef). Positive PCR results were obtained while no typical colonies were recovered (only *Listeria welshimeri* strain).

3.2 Relative level of detection

The relative level of detection is the level of detection at $P = 0.50$ (LOD_{50}) of the alternative (proprietary) method divided by the level of detection at $P = 0.50$ (LOD_{50}) of the reference method.

The RLOD is defined as the ratio of the alternative and reference methods:

$$RLOD = \frac{LOD_{Alt.}}{LOD_{Ref.}}$$

3.2.1 Categories, sample types and strains

Six (matrix/strain) pairs were analyzed by the reference method and by the alternative method (See Table 13):

Table 13 - List of selected types and strains per category, as tested within the relative level of detection study

Category	Type	Strain	Reference number	Strain origin	Seeding procedure
Meat products	Sausage	<i>L. monocytogenes</i>	Ad669	Meat product	48 h at 5 ± 3°C
Milk and dairy products	Goat cheese	<i>L. monocytogenes</i>	Ad618	Cheese	
Fish and seafood products	Smoked salmon	<i>L. monocytogenes</i>	Ad670	Smoked salmon	
Ready-to-eat, ready-to-reheat and ready-to-cook products	Potatoes	<i>L. monocytogenes</i>	Ad279	Vegetables based, ready-to-eat food	
Environmental samples	Process water	<i>L. monocytogenes</i>	Ad631	Dairy environment	
Fresh produce and vegetables	Pre-cut bagged spinach	<i>L. monocytogenes</i>	CWD 15554	Dairy product	

3.2.2 Test sample preparations

Three levels of artificial contamination were prepared for each type:

- Negative control level: one non-inoculated in order to get 5 test portions,
- Low level: one inoculated between 0.2 and 2.0 CFU/sample in order to get 20 test portions providing fractional recovery,
- Higher level: one inoculated between 2.0 and 5.0 CFU/sample in order to get 5 test portions contaminated at a higher level.

A bulk lot of the matrix was inoculated at each level, homogenized and stored as described in Table 13.

3.2.3 RLOD study results

The tabulated raw data on the RLOD are given in **Appendix 6**.

The RLOD calculations were performed using the Excel spreadsheet available at <http://standards.iso.org/iso/16140> - RLOD version 4 (2024-01-10). The RLOD are given in Table 14.

Table 14 – Presentation of RLOD before and after confirmation of the alternative method results

Category	Matrix / strain	AL	RLOD [95% confidence limit]
		22 + 4 h incubation	22 + 22h incubation
1	Sausage / <i>L. monocytogenes</i> Ad669	/	1.0 [0.4 ; 2.4]
2	Goat cheese / <i>L. monocytogenes</i> Ad618	1.0 [0.5 ; 2.1]	1.0 [0.5 ; 2.1]
3	Smoked salmon / <i>L. monocytogenes</i> Ad670	1.0 [0.4 ; 2.6]	1.0 [0.4 ; 2.6]
4	Potatoes / <i>L. monocytogenes</i> Ad279	1.0 [0.5 ; 2.0]	1.0 [0.5 ; 2.0]
5	Process water / <i>L. monocytogenes</i> Ad631	1.0 [0.4 ; 2.6]	1.0 [0.4 ; 2.6]
6	Pre-cut bagged spinach / <i>L. monocytogenes</i> CWD 15554	1.0 [0.4 ; 2.4]	1.0 [0.4 ; 2.4]
Combined		1.0 [0.7 ; 1.4]	1.0 [0.7 ; 1.4]

The LOD₅₀ calculations were done using the Excel spreadsheet available at <http://standards.iso.org/iso/16140> POD-LOD calculation program - version 12, 2024-03-05. The tests are given in Table 15.

Table 15 - LOD₅₀ results

Category	(Strain / matrix) pair	Level of detection at 50% (CFU / test portion)			Level of detection at 95% (CFU / test portion)		
		Reference method	Alternative method		Reference method	Alternative method	
			22 + 4h	22 + 22h		22 + 4h	22 + 22h
1	Sausage / <i>L. monocytogenes</i> Ad669	0.6 [0.3; 1.1]	/	0.6 [0.3; 1.1]	2.7 [1.5; 4.9]	/	2.7 [1.5; 4.9]
2	Goat cheese / <i>L. monocytogenes</i> Ad618	0.7 [0.4; 1.3]	0.7 [0.4; 1.3]	0.7 [0.4; 1.3]	3.1 [1.7; 5.7]	3.1 [1.7; 5.7]	3.1 [1.7; 5.7]
3	Smoked salmon / <i>L. monocytogenes</i> Ad670	0.6 [0.3; 1.1]	0.6 [0.3; 1.1]	0.6 [0.3; 1.1]	2.5 [1.3; 4.5]	2.5 [1.3; 4.5]	2.5 [1.3; 4.5]
4	Potatoes / <i>L. monocytogenes</i> Ad279	0.8 [0.5; 1.4]	0.8 [0.5; 1.4]	0.8 [0.5; 1.4]	3.6 [2.1; 6.2]	3.6 [2.1; 6.2]	3.6 [2.1; 6.2]
5	Process water / <i>L. monocytogenes</i> Ad631	0.6 [0.3; 1.2]	0.6 [0.3; 1.2]	0.6 [0.3; 1.2]	2.7 [1.4; 5.0]	2.7 [1.4; 5.0]	2.7 [1.4; 5.0]
6	Bagged spinach / <i>L. monocytogenes</i> CWD 15554	0.6 [0.3; 1.3]	0.6 [0.3; 1.3]	0.6 [0.3; 1.3]	2.7 [1.3; 5.4]	2.7 [1.3; 5.4]	2.7 [1.3; 5.4]
Combined results		0.7 [0.5; 0.9]	0.7 [0.5; 0.9]	0.7 [0.5; 0.9]	2.9 [2.3; 3.8]	2.9 [2.3; 3.8]	2.9 [2.3; 3.8]

3.2.4 Conclusion

The RLOD values (using the confirmed alternative method results) meet the acceptability limit, which is 1.5 for paired, for all categories tested.

3.3 Inclusivity / exclusivity

The inclusivity is the ability of the alternative method to detect the target analyte from a wide range of strains. The exclusivity is the lack of interference from a relevant range of non-target strains of the alternative method.

3.3.1 Protocols

> Inclusivity

50 *Listeria monocytogenes* strains were freshly cultured in BHI medium at $37 \pm 1^\circ\text{C}$. Dilutions were made in order to inoculate 10 CFU/ 225 ml HFB. The alternative method protocol was then performed considering 4h of subculture. For 2 strains, UHT milk was added to the enrichment broth (25 ml) in order to mimic the real conditions of analysis.

> Exclusivity

30 strains were freshly cultured in BHI medium at $37 \pm 1^\circ\text{C}$. Dilutions were made in order to inoculate about 10^5 CFU/ ml BHI. The alternative method was then performed.

In 2024, It has been required by the MicroVal technical committee to incorporate new *Listeria* strains into the inclusivity / exclusivity list for *Listeria* spp and *Listeria monocytogenes* validation study. A ratio of 80% of sensu stricto and 20% of sensu lato must be considered.

Therefore, in order to complete the list of strains tested during the initial validation, the strains described in Table 16 were added for this renewal.

Table 16 - List of added strains in exclusivity during the renewal in 2025

Strain	Species	Classification (Carlin et al. 2021)	Classification (Bouznada et al., 2024)	Reference	Origin
<i>Listeria</i>	<i>farberii</i>	Sensu stricto	<i>Listeria</i>	LMG 31917	Soil
<i>Listeria</i>	<i>immobilis</i>	Sensu stricto	<i>Listeria</i>	LMG 31920	Soil
<i>Listeria</i>	<i>marthii</i>	Sensu stricto	<i>Listeria</i>	DSM 23813	Soil
<i>Listeria</i>	<i>cornellensis</i>	Sensu lato	<i>Paenilisteria</i>	DSM26689	water
<i>Listeria</i>	<i>fleischmanii</i> subsp <i>fleischmanii</i>	Sensu lato	<i>Mesolisteria</i>	DSM24998	Hard cheese
<i>Listeria</i>	<i>grandensis</i>	Sensu lato	<i>Paenilisteria</i>	DSM26688	water
<i>Listeria</i>	<i>riparia</i>	Sensu lato	<i>Paenilisteria</i>	DSM26685	Running water
<i>Listeria</i>	<i>rocourtiae</i>	Sensu lato	<i>Listeria / Paenilisteria</i>	DSM 22097	lettuce
<i>Listeria</i>	<i>grayi</i>	Sensu lato	<i>Murraya</i>	Ad3288	Smoked bacon
<i>Listeria</i>	<i>grayi</i>	Sensu lato	<i>Murraya</i>	Ad3205	Chicken nuggets
<i>Listeria</i>	<i>grayi</i>	Sensu lato	<i>Murraya</i>	Ad2415	Rillettes
<i>Listeria</i>	<i>grayi</i>	Sensu lato	<i>Murraya</i>	Ad1295	Spinach

3.3.2 Results

Raw data are given in **Appendix 7**.

> Inclusivity

A total of 50 strains were tested for inclusivity. Forty-nine (49) of these strains showed the expected positive result, with *Listeria monocytogenes* Ad253 producing a negative result following the alternative method but a positive result following the reference method.

> Exclusivity

30 strains during the initial validation study and 12 strains during this renewal study were tested for exclusivity. All the strains showed the expected negative result.

3.3.3 Conclusion

The alternative Assurance GDS *Listeria monocytogenes* Tq detection method is selective and specific.

3.4 Conclusion Method Comparison Study

Overall, the conclusions for the Method Comparison Study are:

The observed values for TND-PD and TND+PD for the individual categories and for all categories meet the acceptability limits (observed values \leq AL). The RLOD values (using the confirmed alternative method results) meet the acceptability limit, which is 1.5 for paired studies for all categories tested. The alternative Assurance GDS *Listeria* species Tq detection method is selective and specific.

4 INTER-LABORATORY STUDY

The inter-laboratory study is a study performed by multiple laboratories testing identical samples at the same time, the results of which are used to estimate alternative-method performance parameters.

During the inter-laboratory study, 4 h incubation time was tested.

4.1 Study organisation

> *Collaborators number*

Samples were sent to 7 laboratories gathering 14 collaborators.

> *Matrix and strain used*

Cheese samples were inoculated with *Listeria monocytogenes* 153 strain.

> *Samples*

Samples were inoculated and sent on Monday 13th May 2013, as described below:

- 24 codified samples for *Listeria monocytogenes* research by GDS *Listeria monocytogenes* Tq method and by the reference method (NF EN ISO 11290-1/A1),
- 1 sample for aerobic mesophilic flora enumeration by ISO 4833 method,

- 1 water flask labelled “Temperature Control” with a temperature probe.

The analyses were started on Wednesday 15th May 2013.

> *Inoculation*

The targeted inoculation levels were the following:

- 0 CFU/25 ml,
- 1 – 10 CFU/25 ml,
- 5 – 50 CFU/25 ml.

> *Labelling and shipping*

Blind coded samples were placed in isothermal boxes, which contained cooling blocks, and express-shipped to the different laboratories.

A temperature control flask containing a sensor was added to the package in order to register the temperature profile during the transport, the package delivery and storage until analyses.

Samples were shipped in 24 h to 48 h to the involved laboratories. The temperature conditions had to stay lower or equal to 8°C during transport, and between 0°C – 8°C in the labs.

> *Analyses*

Collaborative study laboratories and the expert laboratory carried out the analyses at Day 2 with the alternative and reference methods.

4.2 Experimental parameters controls

4.2.1 *Detection of Listeria monocytogenes in the matrix before inoculation*

In order to detect the presence of *Listeria monocytogenes*, the reference method was performed on five portions (25 g) before the inoculation. All the results were negative.

4.2.2 Strain stability during transport

Sample stability was checked by inoculating the matrix at 100 CFU/g and 5 CFU/g. Enumerations were performed for the high contamination level and detection analyses were performed for the low contamination level. *Triplicates* were analysed. The mesophilic aerobic flora was also enumerated; the results are given in Table 17.

Table 17 - *Listeria monocytogenes* Sample stability

Day	Reference method (detection)			CFU/g (O&A)			Mesophilic aerobic flora (CFU/g)
	Sample 1	Sample 2	Sample 3	Sample 1	Sample 2	Sample 3	
Day 0	+	+	+	130	130	130	3.3 x 10 ⁶
Day 1	+	+	+	100	150	170	4.0 x 10 ⁶
Day 2	+	+	+	160	150	120	2.9 x 10 ⁶

No evolution was observed during storage at 5°C ± 3°C.

4.2.3 Contamination levels

The contamination levels and the sample codification were the following (see Table 18).

Table 18 - Contamination levels

Level	Samples	Theoretical target level (CFU/test portion)	True level (CFU/test portion)	Low limit (CFU/test portion)	High limit (CFU/test portion)
Level 0	3 – 5 – 9 – 13 – 14 – 18 – 22 – 23	0	/	/	/
Low level	2 – 7 – 8 – 11 – 15 – 17 – 21 – 24	2	1.9	1.5	2.3
High level	1 – 4 – 6 – 10 – 12 – 16 – 19 – 20	25	25.0	22.1	28.4

4.2.4 Logistic conditions

The temperatures measured at reception by the collaborators, the temperatures registered by the thermo-probe, and the receipt dates are given in Table 19.

Table 19 - Sample temperatures at receipt

Collaborators	Temperature measured by the sensor (°C)	Temperature measured at receipt (°C)	Receipt date and time	
A	<i>Lost</i>	3.9	14/05/2013	14h15
B	2.0	4.8	14/05/2013	14h15
C	1.0	2.1	14/05/2013	15h00
D	3.5	4.4	14/05/2013	15h00
E	2.0	3.2	14/05/2013	13h00
F	3.0	3.0 – 4.0	14/05/2013	09h30
G	<i>Lost</i>	3.0 – 4.0	14/05/2013	10h00
H	1.0	4.3	14/05/2013	10h30
I	2.5	5.3	14/05/2013	10h30
J	0.5	2.5	14/05/2013	11h00
K	<i>Lost</i>	3.0	14/05/2013	11h00
L	1.0	2.5	14/05/2013	11h00
M	<i>Out of order</i>	4.0	14/05/2013	11h00
N	2.0	5.0	14/05/2013	11h00

No problem was encountered during the transport or at receipt for the 14 collaborators. All the samples were delivered on time and in appropriate conditions. For four collaborators, probe was not available but temperatures during receipt were all correct.

4.3 Calculation and summary of data

The raw data are given in **Appendix 8**.

4.3.1 MicroVal Expert laboratory results

The results obtained by the expert laboratory are given in Table 20.

Table 20 – Results obtained by the expert Lab.

Level	Reference method	Alternative method
L0	0/8	0/8
L1	7/8	7/8
L2	8/8	8/8

4.3.2 Results observed by the collaborative laboratories

> Mesophilic aerobic flora enumeration

Depending on the Lab results, the enumeration levels varied from $1.7 \cdot 10^6$ to $6.0 \cdot 10^6$ CFU/g.

> Listeria monocytogenes detection

7 Labs participated in the study, involving at least 14 collaborators. The results obtained by the individual collaborators in the inter-laboratory are summarized in Table 21 (reference method) and Table 22 (alternative method).

Table 21 - Positive results by the reference method (ALL the collaborators)

Collaborators	Contamination level		
	L0	L1	L2
A	0	8	8
B	0	7	8
C	0	8	8
D	0	6	8
E	2	8	8
F	0	8	8
G	0	8	8
H	0	7	8
I	0	7	8
J	0	8	8
K	0	8	8
L	0	6	8
M	0	4	8
N	0	7	8
Total	P₀ = 2	P₁ = 100	P₂ = 112

Table 22 - Positive results (before and after confirmation) by the alternative method (ALL the collaborators)

Collaborators	Contamination level								
	L0			L1			L2		
	PCR result	Confirmation result	Final result	PCR result	Confirmation result	Final result	PCR result	Confirmation result	Final result
A	0	0	0	8	8	8	8	8	8
B	0	0	0	7	7	7	8	8	8
C	0	0	0	8	8	8	8	8	8
D	0	0	0	6	6	6	8	8	8
E	0	0	0	7	8	7	8	8	8
F	0	0	0	8	8	8	8	8	8
G	0	0	0	8	8	8	8	8	8
H	0	0	0	7	7	7	7	8	7
I	0	0	0	4	7	4	8	8	8
J	0	0	0	8	8	8	8	8	8
K	1	0	0	8	8	8	8	8	8
L	0	0	0	6	6	6	8	8	8
M	0	0	0	3	4	3	8	8	8
N	0	0	0	7	8	7	8	8	8
Total	P₀ = 1	C₀ = 0	CP₀ = 0	P₁ = 95	C₁ = 101	CP₁ = 95	P₂ = 112	C₂ = 112	CP₂ = 111

One collaborator (E) obtained 2 positive results at Level L0 with the reference method. According to 2023-268 (TC) MVTC interpretation of ISO 16140-2 -6 v2.6, all data from collaborators that found positive results with the blank samples either by the reference or alternative method should not be taken into account. Therefore, the results from the collaborator E were not kept for interpretation.

One collaborator (K) has obtained, at level L1, one positive PCR result with negative confirmation (PD_{FP(alt)} sample). It was decided during the initial validation study to keep this collaborator.

4.3.3 Results of the collaborators retained for interpretation

The results obtained with the 13 collaborators kept for interpretation are presented in Table 23 (reference method) and Table 24 (alternative method).

Table 23 - Positive results by the reference method (Without Lab E)

Collaborators	Contamination level		
	L0	L1	L2
A	0	8	8
B	0	7	8
C	0	8	8
D	0	6	8
F	0	8	8
G	0	8	8
H	0	7	8
I	0	7	8
J	0	8	8
K	0	8	8
L	0	6	8
M	0	4	8
N	0	7	8
Total	P₀ = 0	P₁ = 92	P₂ = 104

**Table 24 - Positive results (before and after confirmation)
 by the alternative methods (Without Lab E)**

Collaborators	Contamination level								
	L0			L1			L2		
	PCR result	Confirmation result	Final result	PCR result	Confirmation result	Final result	PCR result	Confirmation result	Final result
A	0	0	0	8	8	8	8	8	8
B	0	0	0	7	7	7	8	8	8
C	0	0	0	8	8	8	8	8	8
D	0	0	0	6	6	6	8	8	8
F	0	0	0	8	8	8	8	8	8
G	0	0	0	8	8	8	8	8	8
H	0	0	0	7	7	7	7	8	7
I	0	0	0	4	7	4	8	8	8
J	0	0	0	8	8	8	8	8	8
K	1	0	0	8	8	8	8	8	8
L	0	0	0	6	6	6	8	8	8
M	0	0	0	3	4	3	8	8	8
N	0	0	0	7	8	7	8	8	8
Total	P₀ = 1	C₀ = 0	CP₀ = 0	P₁ = 88	C₁ = 93	CP₁ = 88	P₂ = 103	C₂ = 104	CP₂ = 103

4.3.4 Calculation of the specificity percentage (SP)

The percentage specificities (SP) of the reference method and of the alternative method, using the data after confirmation, based on the results of level L0, are the following (See Table 25).

Table 25 - Percentage specificity

Specificity for the reference method	$SP_{ref} = \left(1 - \left(\frac{P_0}{N_-} \right) \right) \times 100 \% =$	100.0 %
Specificity for the alternative method	$SP_{alt} = \left(1 - \left(\frac{CP_0}{N_-} \right) \right) \times 100 \% =$	100.0 %

N: number of all L0 tests

P_0 = total number of false-positive results obtained with the blank samples before confirmation

CP_0 = total number of false-positive results obtained with the blank samples

4.3.5 Calculation of sensitivity (SE_{alt}), sensitivity for the reference method (SE_{ref}), relative trueness (RT), false positive ratio for the alternative method (FPR) and false negative ratio for the alternative method (FNR)

Partial positive results were obtained for the low level (88.5.0% (L1) for alternative method and 84.6% and for the reference method) but above the range described in the ISO 16140-2 for fractional positive results (25-75%). The interpretation of the data was performed for Level 1 only.

Despite this high percentage of positives, the distribution of results across collaborators indicates that the alternative method was still appropriately challenged. Specifically, 7 out of 13 collaborators obtained at least one negative result: 3 collaborators have obtained 7 positives, 2 have obtained 6 positives, 1 have obtained 4 positives and 1 have 3 positives out of 8 contaminated samples. This variability is typical in low-level contamination studies and confirms that the method was tested under limiting conditions.

A summary of the results of the collaborators retained for interpretation and obtained with the reference and the alternative methods for Level 1 is provided in Table 26.

Table 26 - Summary of results for all collaborators obtained with the reference and alternative methods for Level 1

Level	Response	Reference method positive (R+)	Reference method negative (R-)
L1	Alternative method positive (A+)	Positive agreement (A+/R+) PA = 88	Positive deviation (R-/A+) PD = 0
	Alternative method negative (A-)	Total Negative deviation (A-/R+) TND = 4 (4 ND_{FN(alt)})	Total Negative agreement (A-/R-) TNA = 12 (1 NA_{FN(alt)})

Based on the data summarized in Table 26, the values of sensitivity of the alternative and reference methods, as well as the relative trueness and false positive ratio for the alternative method taking account the confirmations, are the following (See Table 27).

Table 27 - Sensitivity, relative trueness, false positive ratio and false negative ratio percentages

		Level L1
Sensitivity for the alternative method	$SE_{alt} = \frac{(PA + PD)}{(PA + TND + PD)} \times 100 \%$	95.7 %
Sensitivity for the reference method	$SE_{ref} = \frac{(PA + TND)}{(PA + TND + PD)} \times 100 \%$	100.0 %
Relative trueness	$RT = \frac{(PA + TNA)}{N} \times 100 \%$	96.2 %
False positive ratio for the alternative method (unpaired evaluation)	$FPR = \frac{PA_{FP(alt)} + PD_{FP(alt)}}{TNA} \times 100 \%$	0.0 %
False positive ratio for the alternative method (paired evaluation)	$FPR = \frac{PA_{FP(alt)}}{TNA} \times 100 \%$	5.4 %

Paired study with $TNA = NA + PD_{FP(alt)}$ $TND = ND_{FN(alt)}$

4.3.6 Interpretation of trueness data

Negative deviations are listed in Table 28 for Level 1. No positive deviation was observed.

All samples with negative deviation have obtained negative PCR result while positive confirmation was obtained (ND_{FN(alt)} samples). Second test was carried out for 3 samples (from collaborator I) and positive PCR result was obtained.

Table 28 - Negative deviations for Level 1

Level	Collaborator	Sample	Reference method result ISO 11290-1	Alternative method: GDS <i>Listeria monocytogenes</i>				Agreement
				PCR	OCLA	Palcam	Final result	
L1	I	2	+	-/+*	+	+	-	ND _{FN(alt)}
		7	+	-/+*	+	+	-	ND _{FN(alt)}
		11	+	-/+*	+	+	-	ND _{FN(alt)}
	M	15	+	-	+	+	-	ND _{FN(alt)}

* second test

For a **paired study design**, the difference between (TND – PD) and the addition (TND + PD) are calculated for the level(s) where fractional recovery is obtained (so L_1 and possibly L_2). The observed value found for (TND – PD) and (TND + PD) shall not be higher than the AL.

For 13 collaborators, the limits are the following:

	Calculated values	AL	Conclusion
	L1		L1
TND - PD	4	4	TND – PD = AL
TND + PD	4	5	TND + PD < AL

The ISO 16140-2:2016 & ISO 16140-2/A1:2024 requirements are fulfilled as (TND - PD) and (TND + PD) meet the AL for Level L1.

4.3.7 Evaluation of the $LOD_{50\%}$ and RLOD between laboratories

The $LOD_{50\%}$, was calculated using the EN ISO 16140-2 Excel spreadsheet available at https://standards.iso.org/iso/16140/-2/ed-1/en/amd/1/PODL0D-interlab_ver2.xlsm

The RLOD is defined as the ratio of the LODs of the alternative method and the reference method: **RLOD = LOD_{alt}/LOD_{ref}** .

The results are given in Table 29.

Table 29 - $LOD_{50\%}$ and RLOD

Method	$LOD_{50\%}$	RLOD
Reference	0.6 [0.5;0.8]	1.2
Alternative	0.7 [0.5;0.9]	

4.4 Conclusion Inter-laboratory study

The observed values for TND-PD and TND+PD are lower than the acceptability limits. The data and interpretations comply with the ISO 16140-2:2016, ISO 16140-2/A1:2024 requirements.

The Assurance GDS *Listeria monocytogenes* Tq is considered equivalent to the ISO standard.

5 GENERAL CONCLUSION

The method comparison study conclusions are:

Overall, the conclusions for the Method Comparison Study are:

The observed values for TND-PD and TND+PD for the individual categories and for all categories meet the acceptability limits (observed values \leq AL).

The RLOD values (using the confirmed alternative method results) meet the acceptability limit, which is 1.5 for paired studies for all categories tested.

The alternative Assurance® GDS *Listeria monocytogenes* Tq detection method is selective and specific.

The inter-laboratory study conclusions are:

The observed values for TND-PD and TND+PD are lower than the acceptability limits.

The data and interpretations comply with the ISO 16140-2:2016, ISO 16140-2/A1:2024 requirements.

The Assurance GDS *Listeria monocytogenes* Tq is considered equivalent to the ISO standard.

Quimper, 25 February 2026

Astrid CARIOU
Manager
Method performance in food microbiology

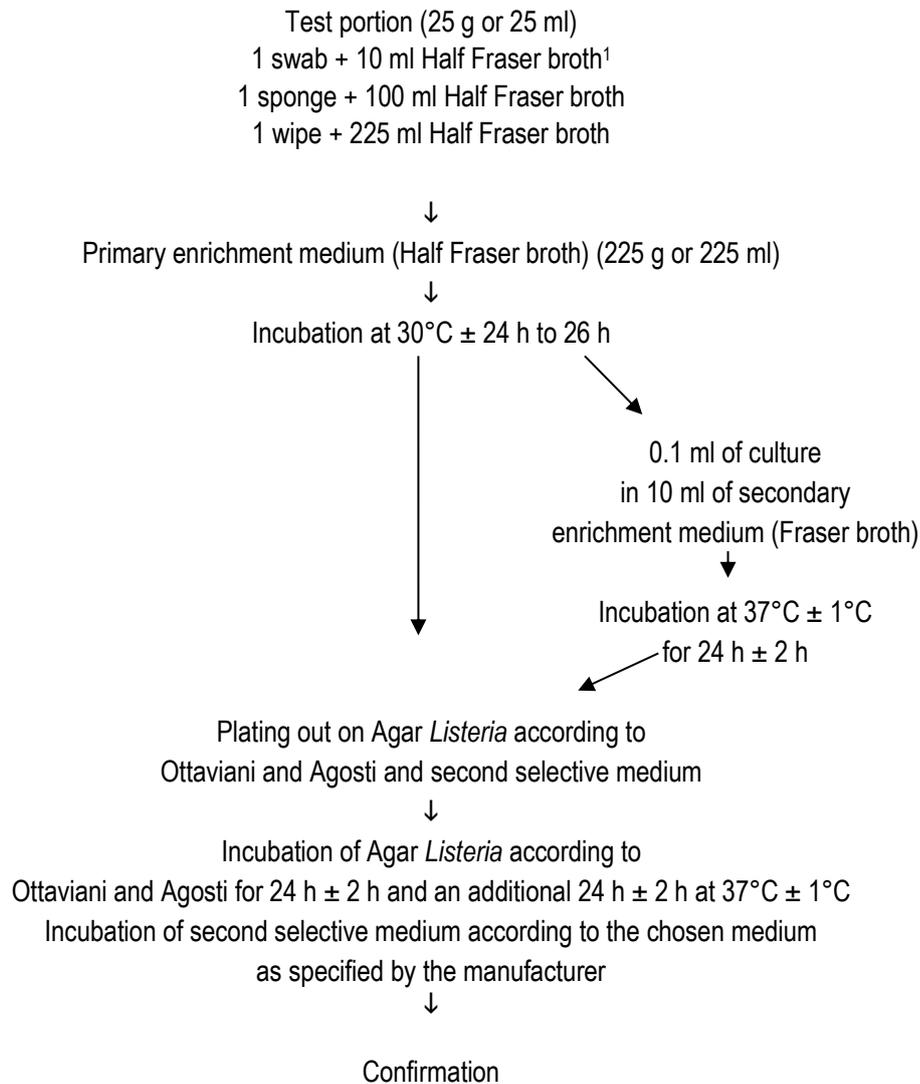


I hereby attest to the validation of the verification of the conformity of the report (opinion and interpretation).

6 REFERENCES

- ISO 4833-1:2013; Microbiology of the food chain -- Horizontal method for the enumeration of microorganisms -- Part 1: Colony count at 30 degrees C by the pour plate technique
- ISO 6887 (all parts); Microbiology of the food chain -- Preparation of test samples, initial suspension and decimal dilutions for microbiological examination – All parts.
- ISO 7218:2024; Microbiology of food and animal feeding stuffs -- General requirements and guidance for microbiological examinations.
- ISO 16140-2:2016; Microbiology of the food chain -- Method validation -- Part 2: Protocol for the validation of alternative (proprietary) methods against a reference method.
- 2016-028 (GC) Proposed MicroVal Technical committee interpretation of ISO 16140-2 and ISO
- 2017-063 (TC) The MicroVal Process

Appendix 1 - Flow diagram of the reference method: ISO 11290-1:2017 - Microbiology of the food chain - Horizontal method for the detection and enumeration of *Listeria monocytogenes* and other *Listeria* spp. - Part 1: detection method



Target	Gram	Catalase	Beta hemolysis	CAMP test	Carbohydrates
<i>Listeria monocytogenes</i>	x	Optional	x	Optional	x

¹ For sampling after cleaning process pre-moisten

- 1 swab + 1 ml broth universal neutralizing (+ 9 ml Half-Fraser)
- 1 sponge + 10 ml broth universal neutralizing (+ 90 ml Half-Fraser)
- 1 wipe + BPW + 10 % neutralizing agent (+ 225 ml Half-Fraser)

**Appendix 2 - Flow diagram of the alternative method:
Assurance GDS *Listeria monocytogenes* Tq**



Appendix 3 - Kit insert(s)

See attached

Appendix 4 - Artificial inoculations

N° Sample	Product	Artificial contaminations					Global result
		Strain	Origin	Injury protocol	Injury measurement	Inoculation level/25g	
4777	Fermented milk	<i>L. monocytogenes</i> 18024	Raw milk	pH4 -2 months	0.43	2-5-12-2-3(4.8)	+
4778	Fermented milk	<i>L. monocytogenes</i> 18024	Raw milk	pH4 -2 months	0.43	2-5-12-2-3(4.8)	+
4771	Pasteurized fresh cream	<i>L. monocytogenes</i> 18312	Milk	HT 56°C 10min	0.38	3-5-7-7-6(5.6)	+
4772	Pasteurized half skimmed milk	<i>L. monocytogenes</i> 18312	Milk	HT 56°C 10min	0.38	3-5-7-7-6(5.6)	-
4773	Pasteurized milk	<i>L. monocytogenes</i> 18312	Milk	HT 56°C 10min	0.38	3-5-7-7-6(5.6)	+
265	Wipe (ready to eat production)	<i>L. monocytogenes</i> 1973/2400	Quiche lorraine	48h 4°C	Seeding	2-0-2-1-3 (1.6)	-
266	Wipe (ready to eat production)	<i>L. monocytogenes</i> 877/113	Environment	48h 4°C	Seeding	1-2-2-1-1 (1.4)	-
267	Wipe (ready to eat production)	<i>L. monocytogenes</i> 877/113	Environment	48h 4°C	Seeding	1-2-2-1-1 (1.4)	-
4774	Fermented milk	<i>L. monocytogenes</i> 909	Milk	pH4 -2 months	0.47	3-5-7-4-4(4.6)	+
4760	Ready to eat meal (pork meat with dressing)	<i>L. monocytogenes</i> A00C014	Sausages	HT 56°C 10min	0.62	4-5-9-5-5(5.6)	+
4763	Ready to eat meal (beef balls with ketchup)	<i>L. monocytogenes</i> A00C014	Sausages	HT 56°C 10min	0.62	4-5-9-5-5(5.6)	+
6830	Wipe (fish industry materials)	<i>L. monocytogenes</i> AA0E033	Environment	4°C/2 days	Seeding	6-2-4-2-2 (3.2)	-
6831	Wipe (fish industry materials)	<i>L. monocytogenes</i> AA0E033	Environment	4°C/2 days	Seeding	6-2-4-2-2 (3.2)	+
6832	Wipe (fish industry materials)	<i>L. monocytogenes</i> AA0E033	Environment	4°C/2 days	Seeding	6-2-4-2-2 (3.2)	-
5174	Frozen artichokes	<i>L. monocytogenes</i> Ad 1212	Mushrooms	-20°C-4 days	Seeding	6-3-11-5-6(6.2)	+
5175	Frozen tomatoes	<i>L. monocytogenes</i> Ad 1212	Mushrooms	-20°C-4 days	Seeding	6-3-11-5-6(6.2)	+
5176	Frozen tomatoes	<i>L. monocytogenes</i> Ad 1212	Mushrooms	-20°C-4 days	Seeding	6-3-11-5-6(6.2)	+
5177	Frozen zucchini	<i>L. monocytogenes</i> Ad 1212	Mushrooms	-20°C-4 days	Seeding	6-3-11-5-6(6.2)	+
5178	Frozen peppers	<i>L. monocytogenes</i> Ad 1212	Mushrooms	-20°C-4 days	Seeding	6-3-11-5-6(6.2)	+

N° Sample	Product	Artificial contaminations					Global result
		Strain	Origin	Injury protocol	Injury measurement	Inoculation level/25g	
253	Process water (tomato production)	<i>L. monocytogenes</i> Ad 634	Environment	pH10 2 month	0.14	4-5-3-3-1 (3.2)	+
255	Process water (fish industry)	<i>L. monocytogenes</i> Ad 634	Environment	pH10 2 month	0.14	4-5-3-3-1 (3.2)	+
254	Process water (cheese production)	<i>L. monocytogenes</i> Ad 635	Environment	pH10 2 month	0.14	4-5-3-3-1 (3.2)	+
235	Prawns	<i>L. monocytogenes</i> Ad1192	Fish filet	4°C 2 month	0.21	4-3-9-2-4 (4.4)	+
236	Seafood cocktail	<i>L. monocytogenes</i> Ad1192	Fish filet	4°C 2 month	0.21	4-3-9-2-4 (4.4)	+
237	Fillet of Cod	<i>L. monocytogenes</i> Ad1192	fish filet	4°C 2 month	0.21	4-3-9-2-4 (4.4)	+
238	Fillet of Cod	<i>L. monocytogenes</i> Ad1192	Fish filet	4°C 2 month	0.21	4-3-9-2-4 (4.4)	+
5184	Vanilla profiterolles	<i>L. monocytogenes</i> Ad1195	Omelet	-20°C-4 days	Seeding	5-9-10-2-10(7.2)	+
5185	Pastry (Paris Brest)	<i>L. monocytogenes</i> Ad1195	Omelet	-20°C-4 days	Seeding	5-9-10-2-10(7.2)	+
5186	Cakes (macaron)	<i>L. monocytogenes</i> Ad1195	Omelet	-20°C-4 days	Seeding	5-9-10-2-10(7.2)	+
5187	Dairy based dessert	<i>L. monocytogenes</i> Ad1195	Omelet	-20°C-4 days	Seeding	5-9-10-2-10(7.2)	+
5188	Pastry	<i>L. monocytogenes</i> Ad1195	Omelet	-20°C-4 days	Seeding	5-9-10-2-10(7.2)	+
4757	Ready to eat meal (beef with dressing)	<i>L. monocytogenes</i> Ad1218	Ground beef	HT 56°C 10min	0.5	5-3-2-6-7(4.6)	+
4758	Ready to eat meal (beef tongue with dressing)	<i>L. monocytogenes</i> Ad1218	Ground beef	HT 56°C 10min	0.5	5-3-2-6-7(4.6)	+
4759	Ready to eat meal (beef balls with dressing)	<i>L. monocytogenes</i> Ad1218	Ground beef	HT 56°C 10min	0.5	5-3-2-6-7(4.6)	+
4761	Ready to eat meal (beef Bourguignon)	<i>L. monocytogenes</i> Ad1218	Ground beef	HT 56°C 10min	0.5	5-3-2-6-7(4.6)	+
4762	Ready to eat meal (beef kidneys with dressing)	<i>L. monocytogenes</i> Ad1218	Ground beef	HT 56°C 10min	0.5	5-3-2-6-7(4.6)	+
251	Process water (ham production)	<i>L. monocytogenes</i> Ad1278	Environment	pH10 2 month	0.2	6-6-4-7-6 (5.8)	+
252	Process water (fish industry)	<i>L. monocytogenes</i> Ad1278	Environment	pH10 2 month	0.2	6-6-4-7-6 (5.8)	+
245	Salmon tartar	<i>L. monocytogenes</i> Ad1279	Smoked fish	NaCl 10% 2 month	0.18	5-1-1-6-6 (3.8)	+

N° Sample	Product	Artificial contaminations					Global result
		Strain	Origin	Injury protocol	Injury measurement	Inoculation level/25g	
246	Smoked salmon	<i>L. monocytogenes</i> Ad1279	Smoked fish	NaCl 10% 2 month	0.18	5-1-1-6-6 (3.8)	+
247	Smoked salmon bacon	<i>L. monocytogenes</i> Ad1279	Smoked fish	NaCl 10% 2 month	0.18	5-1-1-6-6 (3.8)	+
243	Sliced salmon	<i>L. monocytogenes</i> Ad1412	Smoked salmon	NaCl 10% 2 month	0.48	9-8-8-1-6 (6.4)	+
244	Marinated salmon	<i>L. monocytogenes</i> Ad1413	Smoked salmon	NaCl 10% 2 month	0.48	9-8-8-1-6 (6.4)	+
231	Prawns	<i>L. monocytogenes</i> Ad1499	Trout eggs	4°C 2 month	0.22	9-14-5-6-5 (6.8)	+
232	Prawns	<i>L. monocytogenes</i> Ad1499	Trout eggs	4°C 2 month	0.22	9-14-5-6-5 (6.8)	+
233	Crayfish	<i>L. monocytogenes</i> Ad1499	Trout eggs	4°C 2 month	0.22	9-14-5-6-5 (6.8)	+
234	Prawns	<i>L. monocytogenes</i> Ad1499	Trout eggs	4°C 2 month	0.22	9-14-5-6-5 (6.8)	+
6849	Process water (vegetables industry)	<i>L. monocytogenes</i> Ad1672	Environment	10min 56°C	0.06	3-4-1-3-1 (2.4)	-
6850	Process water (vegetables industry)	<i>L. monocytogenes</i> Ad1672	Environment	10min 56°C	0.06	3-4-1-3-1 (2.4)	-
263	Wipe (ready to eat production)	<i>L. monocytogenes</i> Ad1678	Ready-to-eat meal (cheese/spinach)	48h 4°C	Seeding	1-0-1-1-0 (0.6)	+
264	Wipe (ready to eat production)	<i>L. monocytogenes</i> Ad1678	Ready-to-eat meal (cheese/spinach)	48h 4°C	Seeding	1-0-1-1-0 (0.6)	+
6837	Wipe (fish industry materials)	<i>L. monocytogenes</i> Ad1679	Environment	4°C/2 days	Seeding	4-5-9-3-5 (5.2)	+
6855	Process water (fish industry)	<i>L. monocytogenes</i> Ad243	Environment	10min 56°C	1.02	0-3-5-3-3 (2.8)	-
6856	Process water (fish industry)	<i>L. monocytogenes</i> Ad243	Environment	10min 56°C	1.02	0-3-5-3-3 (2.8)	-
5241	Dehydrated sausage	<i>L. monocytogenes</i> Ad267	Sausage	pH3 22 days	0.28	3-2-3-2-4(2.8)	-
5242	Sliced dehydrated sausage	<i>L. monocytogenes</i> Ad267	Sausage	pH3 22 days	0.28	3-2-3-2-4(2.8)	+
5243	Salami	<i>L. monocytogenes</i> Ad267	Sausage	pH3 22 days	0.28	3-2-3-2-4(2.8)	+
5244	Dehydrated sausage	<i>L. monocytogenes</i> Ad267	Sausage	pH3 22 days	0.28	3-2-3-2-4(2.8)	+
5245	Dehydrated sausage	<i>L. monocytogenes</i> Ad270	Sausage	pH3 22 days	0.15	4-9-9-8-3(6.6)	+
5246	Dehydrated sausage	<i>L. monocytogenes</i> Ad270	Sausage	pH3 22 days	0.15	4-9-9-8-3(6.6)	-

N° Sample	Product	Artificial contaminations					Global result
		Strain	Origin	Injury protocol	Injury measurement	Inoculation level/25g	
5247	Dehydrated sausage	<i>L. monocytogenes</i> Ad270	Sausage	pH3 22 days	0.15	4-9-9-8-3(6.6)	-
5248	Dehydrated sausage	<i>L. monocytogenes</i> Ad270	Sausage	pH3 22 days	0.15	4-9-9-8-3(6.6)	+
5169	Pizza (cheese)	<i>L. monocytogenes</i> Ad543	Pepper	-20°C-4 days	Seeding	2-8-8-5-2(5.0)	+
5170	Pizza (ham, cheese)	<i>L. monocytogenes</i> Ad543	Pepper	-20°C-4 days	Seeding	2-8-8-5-2(5.0)	+
5171	Pizza (tuna, vegetables)	<i>L. monocytogenes</i> Ad543	Pepper	-20°C-4 days	Seeding	2-8-8-5-2(5.0)	+
5172	Antipasti (artichoke, tomato, olives)	<i>L. monocytogenes</i> Ad543	Pepper	-20°C-4 days	Seeding	2-8-8-5-2(5.0)	-
5173	Frozen carrots	<i>L. monocytogenes</i> Ad543	Pepper	-20°C-4 days	Seeding	2-8-8-5-2(5.0)	+
6833	Wipe (fish industry materials)	<i>L. monocytogenes</i> Ad548	Environment	4°C/2 days	Seeding	3-5-2-5-4 (3.8)	+
6834	Wipe (fish industry materials)	<i>L. monocytogenes</i> Ad548	Environment	4°C/2 days	Seeding	3-5-2-5-4 (3.8)	+
4764	Pasteurized cheese (Gouda)	<i>L. monocytogenes</i> Ad610	Raw milk	HT 56°C 10min	0.45	4-6-7-5-6(5.6)	+
4765	Pasteurized cheese (Gouda)	<i>L. monocytogenes</i> Ad610	Raw milk	HT 56°C 10min	0.45	4-6-7-5-6(5.6)	+
4768	Semolina with milk	<i>L. monocytogenes</i> Ad610	Raw milk	HT 56°C 10min	0.45	4-6-7-5-6(5.6)	+
4770	Pasteurized fresh cream	<i>L. monocytogenes</i> Ad610	Raw milk	HT 56°C 10min	0.45	4-6-7-5-6(5.6)	+
6841	Dusts (fish industry)	<i>L. monocytogenes</i> Ad614	Environment	4°C/2 days	Seeding	4-7-2-5-7 (5.0)	+
6842	Dusts (fish industry)	<i>L. monocytogenes</i> Ad614	Environment	4°C/2 days	Seeding	4-7-2-5-7 (5.0)	+
273	Dusts (dairy industry)	<i>L. monocytogenes</i> Ad614	Dairy environment	48h 4°C	Seeding	3-3-4-1-1 (2.4)	+
274	Dusts (dairy industry)	<i>L. monocytogenes</i> Ad614	Dairy environment	48h 4°C	Seeding	3-3-4-1-1 (2.4)	-
272	Dusts (dairy industry)	<i>L. monocytogenes</i> Ad621	Dairy environment	48h 4°C	Seeding	0-0-1-0-1 (0.4)	-
270	Dusts (dairy industry)	<i>L. monocytogenes</i> Ad627	Dairy environment	48h 4°C	Seeding	1-0-1-2-2 (1.2)	+
271	Dusts (dairy industry)	<i>L. monocytogenes</i> Ad627	Dairy environment	48h 4°C	Seeding	1-0-1-2-2 (1.2)	-
340	Dusts (ready to eat production)	<i>L. monocytogenes</i> Ad631	Dairy environment	48h 4°C	Seeding	9-8-5-5-3(6.0)	+
341	Dusts (ready to eat production)	<i>L. monocytogenes</i> Ad631	Dairy environment	48h 4°C	Seeding	9-8-5-5-3(6.0)	+

N° Sample	Product	Artificial contaminations					Global result
		Strain	Origin	Injury protocol	Injury measurement	Inoculation level/25g	
342	Dusts (ready to eat production)	<i>L. monocytogenes</i> Ad631	Dairy environment	48h 4°C	Seeding	9-8-5-5-3(6.0)	+
337	Dusts (ready to eat production)	<i>L. monocytogenes</i> Ad633	Dairy environment	48h 4°C	Seeding	5-2-5-6-4(4.4)	+
338	Dusts (ready to eat production)	<i>L. monocytogenes</i> Ad633	Dairy environment	48h 4°C	Seeding	5-2-5-6-4(4.4)	+
339	Dusts (ready to eat production)	<i>L. monocytogenes</i> Ad633	Dairy environment	48h 4°C	Seeding	5-2-5-6-4(4.4)	+
6845	Dusts (Fish industry)	<i>L. monocytogenes</i> Ad635	Environment	4°C/2 days	Seeding	4-5-6-3-3 (4.2)	+
6846	Dusts (Fish industry)	<i>L. monocytogenes</i> Ad635	Environment	4°C/2 days	Seeding	4-5-6-3-3 (4.2)	+
4766	Pasteurized cheese (Chaumes)	<i>L. monocytogenes</i> Ad665	Raw milk	HT 56°C 10min	0.49	1-1-7-3-5(3.4)	+
4767	Pasteurized cheese (Brie)	<i>L. monocytogenes</i> Ad665	Raw milk	HT 56°C 10min	0.49	1-1-7-3-5(3.4)	+
4769	Rice with milk	<i>L. monocytogenes</i> Ad665	Raw milk	HT 56°C 10min	0.49	1-1-7-3-5(3.4)	+
4775	Fermented milk	<i>L. monocytogenes</i> Ad665	Raw milk	pH4 -2 months	0.65	4-4-4-3-8(4.6)	+
4776	Fermented milk	<i>L. monocytogenes</i> Ad665	Raw milk	pH4 -2 months	0.65	4-4-4-3-8(4.6)	+
248	Smoked trout	<i>L. monocytogenes</i> Ad996	Smoked trout	NaCl 10% 2 month	0.3	7-8-7-10-2 (6.8)	+
249	Smoked tuna	<i>L. monocytogenes</i> Ad996	Smoked trout	NaCl 10% 2 month	0.3	7-8-7-10-2 (6.8)	+
250	Smoked haddock	<i>L. monocytogenes</i> Ad996	Smoked trout	NaCl 10% 2 month	0.3	7-8-7-10-2 (6.8)	+



Category	Item Type	Sample n°	Strain	Origin	Injury Protocol	Injury Evaluation (Log10)	Inoculation (CFU/Sample)	Alternative Method Result	Reference Method Result
Fresh Produce and Vegetables	Sliced apples	1	<i>L. monocytogenes</i> ATCC 51780	Dairy Product	Seeding 48 h at 5 ± 3°C	/	4.2	Positive	Positive
	Asparagus 8oz	2						Positive	Positive
	Medjool Dates 12 oz	4						Positive	Positive
	Pitted Dates	6						Positive	Positive
	Broccoli 8 oz	7						Positive	Positive
	Broccoli 12 oz	8	<i>L. monocytogenes</i> QL 030911-10	Shellfish	Seeding 48 h at 5 ± 3°C	/	3.3	Positive	Positive
	Squash 8 oz	9						Positive	Positive
	Carrot & Celery	12						Positive	Positive
	Celery Sticks	14						Positive	Positive
	Salad Kit	19						Positive	Positive
	Baby spinach	20	<i>L. monocytogenes</i> CWD 1609	Turkey Frankfurter	Seeding 48 h at 5 ± 3°C	/	4.6	Positive	Positive
	Chopped salad kit	22						Positive	Positive
	Chopped Romaine	23						Positive	Positive
	Classic coleslaw	25						Positive	Positive
	Classic romaine	26						Positive	Positive



Category	Item Type	Sample n°	Strain	Origin	Injury Protocol	Injury Evaluation (Log10)	Inoculation (CFU/Sample)	Alternative Method Result	Reference Method Result
Fresh Produce and Vegetables	Fruit medley 48 oz	30	<i>L. monocytogenes</i> QL 327429	Ground Beef	Seeding 48 h at 5 ± 3°C	/	4.1	Positive	Positive
	Pineapple chunks	32						Positive	Positive
	Fresh Cauliflower	34						Positive	Positive
	Spring mix	41						Positive	Positive
	Garden salad	43						Positive	Positive
	Coleslaw salad	44	<i>L. monocytogenes</i> QL 327430	Environmental Sponges	Seeding 48 h at 5 ± 3°C	/	5.7	Positive	Positive
	Shredded carrots	48						Positive	Positive
	Veggie blend salad	50						Positive	Positive
	Broccoli pearls	51						Positive	Positive
	Brussel sprouts	52						Positive	Positive
	Snap peas	54	<i>L. monocytogenes</i> QL 313058	Deli Sandwich	Seeding 48 h at 5 ± 3°C	/	5.3	Positive	Positive
	Mango chunks	56						Positive	Positive
	Pineapple cored	58						Positive	Positive
	Pineapple chunks	59						Positive	Positive
	Chopped salad	60						Positive	Positive

Appendix 5 - Sensitivity study: raw data

MEAT PRODUCTS																			Category
N° Sample	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq										
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C				Half Fraser 22h 30°C + 72h 4°C / Half Fraser 4h 30°C						
			OAA	Palcam	OAA	Palcam			PCR Result 22h subculture	Confirmations ALOA	Palcam	Final result 22 h	Agreement Ref/Alt 22h	PCR Result 22h subculture	Confirmations ALOA	Palcam	Final result 22 h	Agreement Ref/Alt 22 h	
1533	Poultry meat	+	H+/H-	+	H+	+	<i>L.monocytogenes/ L.welshimeri</i>	+	+	H+/H-	+	+	PA	+	H+/H-	+	+	PA	1
1534	Sliced beef	-	st	-	st	st	/	-	-	st	st	-	NA						1
1535	Gizzards	-	st	-	st	st	/	-	-	st	st	-	NA						1
1536	Smoked pork sausage	+	H-	+	H-	+	<i>L.welshimeri</i>	-	-	H-	+	-	NA						1
1537	Merguez	+	H-	+/-3col (NC on TSYEA)	H-	+	<i>L.innocua</i>	-	-	H-	+	-	NA						1
1538	Pork meat	+	H+/H-	+	H+/H-	+	<i>L.monocytogenes/ L.innocua</i>	+	+	H+/H-	+	+	PA	+	H+/H-	+	+	PA	1
1539	Sweet bread	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
1805	Merguez	-	-	-	-	-	/	-	-	+/- 1col	-	-	NA						1
1806	Raw poultry meat	+	H-	+	H-	+	<i>L.welshimeri</i>	-	-	+/-	+	-	NA						1
1807	Pork meat	-	st	st	-	st	/	-	-	-	-	-	NA						1
1808	Smoked pork sausage	-	-	-	-	-	/	-	-	-	-	-	NA						1
1809	Chitterling	-	st	-	-	-	/	-	-	-	-	-	NA						1
1810	Roasted pork meat	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
1811	Raw poultry meat	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
1949	Ready to cook poultry dish	+	H-	+	H-	+	<i>L.innocua</i>	-	-	H-	-	-	NA						1
2068	Smoked pork sausage	-	-	-	-	-	/	-	-	-	-	-	NA						1
2069	Marinated poultry meat	-	-	-	-	-	/	-	-	st	-	-	NA						1
2070	Ready to eat poultry meat	+	H+	+	H+/H-	+	<i>L.monocytogenes/ L.innocua</i>	+	+	H+/H-	+	+	PA	+	H+	+	+	PA	1
2071	Marinated poultry meat	+	H+/H-	+	H+/H-	+	<i>L.monocytogenes/ L.welshimeri</i>	+	+	H+/H-	+	+	PA	+	H+/H-	+	+	PA	1
2072	Marinated beef meat	-	st	st	-	st	/	-	-	st	st	-	NA						1
3339	Smoked cocktail sausages	-	-	-	-	-	/	-	-	-	-	-	NA						1
3340	Cooked poultry meat	-	st	st	-	-	/	-	-	-	-	-	NA						1
3341	Marinated beef meat	-	st	-	st	st	/	-	-	st	-	-	NA						1
3342	Marinated poultry meat	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
3343	Stuffed tomatoes	-	st	-	-	-	/	-	-	st	-	-	NA						1
3344	Merguez	+	1H+/H-	+	H+/H-	+	<i>L.monocytogenes/ L.welshimeri</i>	+	-	H-	+	-	ND _{FN(alt)}	-	H+/H-	+	-	ND _{FN(alt)}	1
3345	Raw sausages	+	H+/H-	+	H+	+	<i>L.monocytogenes/ L.welshimeri</i>	+	+	H+/1H-	+	+	PA	+	H+/H-	+	+	PA	1
3346	Marinated poultry meat	-	-	-	-	-	/	-	-	-	-	-	NA						1
3347	Chitterlings	-	st	-	st	st	/	-	-	st	-	-	NA						1
3348	Ham	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
3349	Stuff	-	st	-	-	-	/	-	-	st	-	-	NA						1

* Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

MEAT PRODUCTS

N° Sample	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq										Category
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C				Half Fraser 22h 30°C + 72h 4°C / Half Fraser 4h 30°C						
			OAA	Palcam	OAA	Palcam			PCR Result 22h subculture	Confirmations		Final result 22 h	Agreement Ref/Alt 22h	PCR Result 22h subculture	Confirmations		Final result 22 h	Agreement Ref/Alt 22 h	
							ALOA	Palcam		ALOA	Palcam								
3350	Poultry meat	+	H+	+	H+/H-	+	<i>L.monocytogenes/ L.welshimeri</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
3352	Ground beef(15% fat)	-	H-	+	H-	+	<i>L.welshimeri</i>	-	-	H-	+	-	NA						1
3353	Delicatessen	-	st	-	-	st	/	-	-	st	-	-	NA						1
3354	Cooked sausage	+	1H+/H-	+	H+/H-	+	<i>L.monocytogenes/ L.welshimeri</i>	+	+	H+/H-	+	+	PA	+	H+/H-	+	+	PA	1
3355	Beef meat	+	st	-	H+	+	<i>L.monocytogenes</i>	+	+	H+	-	+	PA	+	H+	-	+	PA	1
3356	Gizzards	-	st	-	-	-	/	-	-	st	-	-	NA						1
3357	Delicatessen(terrine)	-	st	st	st	st	/	-	-	st	st	-	NA						1
3358	Ground beef	-	H-	+	H-	+	<i>L.welshimeri</i>	-	+	H-	+	-	PDFP(alt)	-	H-	+	-	NA	1
3361	Ground beef	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
3451	Precooked ground poultry meat	-	-	-	-	-	/	-	-	-	-	-	NA						1
3452	Raw pork meat	-	st	st	st	st	/	-	-	st	st	-	NA						1
3453	Raw turkey meat	-	st	st	st	st	/	-	-	st	st	-	NA						1
3454	Raw poultry meat	-	st	st	st	st	/	-	-	st	st	-	NA						1
3455	Raw beef meat	-	st	st	st	st	/	-	-	st	st	-	NA						1
4110	Dehydrated sausage	-	st	-	st	st	/	-	-	st	-	-	NA						1
4111	Dehydrated sausage	+	H-	+	H+/H-	+	<i>L.welshimeri/ L.monocytogenes</i>	+	+	H+/H-	+	+	PA	+	6H+/H-	+	+	PA	1
4112	Dehydrated sausage	-	-	-	st	-	/	-	-	-	-	-	NA						1
4113	Ready to cook plate (veal)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
4114	Roe terrine	-	st	-	-	-	/	-	-	-	-	-	NA						1
4115	Liver pâté	-	st	st	st	st	/	-	-	st	st	-	NA						1
4116	Cooked poultry meat with spices	-	st	st	-	-	/	-	-	-	st	-	NA						1
4117	Pâté	-	st	st	st	st	/	-	-	st	st	-	NA						1
4620	Ground beef	-	H-	+	H-	+	<i>L.welshimeri</i>	-	-	H-	+	-	NA						1
4755	Delicatessen(rillettes)	+	st	st	H+	+	<i>L.monocytogenes</i>	+	+	8H+	+	+	PA	+	H+	+	+	PA	1
4757	Ready to eat meal (beef with dressing)	+	6H+	st	H+	+	<i>L.monocytogenes</i>	+	+	3H+	2+	+	PA	+	H+	+	+	PA	1
4758	Ready to eat meal (beef tongue with dressing)	+	3H+	st	H+	+	<i>L.monocytogenes</i>	+	+	3H+	2+	+	PA	+	H+	+	+	PA	1
4759	Ready to eat meal (beef balls with dressing)	+	7H+	+	H+	+	<i>L.monocytogenes</i>	+	+	6H+	+	+	PA	+	H+	+	+	PA	1
4760	Ready to eat meal (pork meat with dressing)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
4761	Ready to eat meal (beef Bourguignon)	+	1H+	5+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
4762	Ready to eat meal (beef kidneys with dressing)	+	4H+	2+	H+	+	<i>L.monocytogenes</i>	+	+	1H+	1+	+	PA	+	H+	+	+	PA	1
4763	Ready to eat meal (beef balls with ketchup)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
5241	Dehydrated sausage	-	-	st	st	st	/	-	-	-	-	-	NA						1
5242	Sliced dehydrated sausage	+	-	st	H+	+	<i>L.monocytogenes</i>	+	-	-	st	-	PDFN(alt)	-	-	st	-	PDFN(alt)	1
5243	Salami	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
5244	Dehydrated sausage	+	st	st	H-	+	<i>L.welshimeri</i>	-	+	H-	st	+	PD	+	H+(L.mono)/H-	+	+	PD	1

MEAT PRODUCTS																			Category
N° Sample	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq										
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C				Half Fraser 22h 30°C + 72h 4°C / Half Fraser 4h 30°C						
			OAA	Palcam	OAA	Palcam			PCR Result 22h subculture	Confirmations		Final result 22 h	Agreement Ref/Alt 22h	PCR Result 22h subculture	Confirmations		Final result 22 h	Agreement Ref/Alt 22 h	
								(H+ at 72H)	(+ at 72H)										
5245	Dehydrated sausage	+	1H+	6+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
5246	Dehydrated sausage	-	st	st	st	st	/	-	-	st	st	-	NA						1
5247	Dehydrated sausage	-	st	st	st	st	/	-	-	st	st	-	NA						1
5248	Dehydrated sausage	+	-	st	H+	+	<i>L.monocytogenes</i>	+	+	-(+Ref)	-(+Fraser 1 ref)	+	PA	+	2H-(<i>L.welshimer</i>)	1+	-	ND _{FN(alt)}	1
5711	Ground beef	+	H+	+	H+	+	<i>L.monocytogenes</i> / <i>L.innocua</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
5712	Ground beef	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	H+	+	+	PA	+	H+	+	+	PA	1
5713	Dehydrated sausage	-	-	-	-	-	/	-	-	-	-	-	NA						1
5714	Poultry meat	-	-	-	-	-	/	-	-	-	-	-	NA						1
5715	Turkey meat	-	-	st	st	st	/	-	-	-	-	-	NA						1

MILK AND DAIRY PRODUCTS

N° Sample	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq														Category			
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C				Half Fraser 22h 30°C + 72h 4°C / Half Fraser 4h 30°C													
			OAA	Palcam	OAA	Palcam			PCR Result		Confirmations		Final result		Agreement Ref/Alt		PCR Result		Confirmations		Final result			Agreement Ref/Alt		
									4 h subculture	22 h subculture	ALO	Palcam	4 h	22 h	4 h	22 h	4 h subculture	22 h subculture	ALO	Palcam	4 h	22 h		4 h	22 h	
1602	Raw milk cheese	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
1603	Raw milk cheese (cow)	+	H+	+ 2 col	H+	+	<i>L.monocytogenes</i>	+	+	+	H+ 2col	+ 3col	+	+	PA	PA	+	+	H+ 1col	+	+	+	+	PA	PA	2
1604	Raw milk cheese (ewe)	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	NA	NA	2	
1605	Raw milk cheese (cow)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
1606	Raw milk cheese	-	H?	-	-	-	NC	-	-	-	-	-	-	-	NA	NA	-	i/-	-	-	-	-	NA	NA	2	
1607	Raw milk cheese	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	NA	NA	2	
1608	Raw milk cheese	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
1884	Raw milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
1885	Raw milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
1886	Raw milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
1887	Raw milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
1888	Raw milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
1889	Raw milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
1890	Raw milk	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
1943	Raw milk cheese	+	H+	-	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
1944	Raw milk cheese	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
1945	Raw milk (ewe)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
1946	Raw milk (ewe)	-	st	st	-	-	/	-	-	-	-	st	-	-	NA	NA									2	
1947	Raw milk (ewe)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
1948	Raw milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
2066	Raw milk cheese	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
2067	Raw milk cheese (ewe)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	2	
3433	Fermented milk	-	st	st	-	-	/	-	-	-	st	-	-	-	NA	NA									2	
3434	Fermented milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
3435	Fermented milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
3436	Fermented milk	-	st	-	st	-	/	-	-	-	st	st	-	-	NA	NA									2	
3437	Fermented milk	-	st	-	-	st	/	-	-	-	st	st	-	-	NA	NA									2	
3438	Fermented milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
3439	Fermented milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
3440	White cheese	-	H-	+	H-	H-	<i>L.innocua</i>	-	-	-	H-	+	-	-	NA	NA									2	
3441	Fermented milk	-	st	-	-	-	/	-	-	-	st	-	-	-	NA	NA									2	
3442	Pasteurized whole milk	-	st	st	-	-	/	-	-	-	st	st	-	-	NA	NA									2	
3443	Pasteurized whole milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
3444	Pasteurized half skimmed milk	-	st	st	-	-	/	-	-	-	st	-	-	-	NA	NA									2	
3445	Pasteurized half skimmed milk	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
3446	White cheese	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA									2	
3447	White cheese	-	st	-	st	-	/	-	-	-	st	-	-	-	NA	NA									2	
3448	Whole fresh cream	-	st	st	-	-	/	-	-	-	st	-	-	-	NA	NA									2	
3449	Pasteurized whole fresh cream	-	st	st	-	-	/	-	-	-	st	-	-	-	NA	NA									2	

* Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

FISH AND SEAFOOD PRODUCTS

N° Sample	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq														Category			
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C						Half Fraser 22h 30°C + 72h 4°C / Half Fraser 4h 30°C											
			OAA	Palcam	OAA	Palcam			PCR Result		Confirmations		Final result		Agreement Ref/Alt		PCR Result		Confirmations		Final result			Agreement Ref/Alt		
									4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h	4 h	22 h	4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h		4 h	22 h	
5801	Breaded fish with lemon	-	-	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5802	Piece of Salmon and lemon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
5803	Cooked shrimp	-	-	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5804	Cooked scallops	-	st	st	st	-	/	-	-	-	st	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5805	Cooked ground salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
5806	Ground fish	+	st	1+	H+	+	<i>L.monocytogenes</i>	+	+	+	2H+	4+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
5807	Cooked scallops	+	H+	-	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	-	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
5808	Cooked fillet of fish	-	st	-	-	-	/	-	-	-	st	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5809	Cooked fillet of fish	-	st	-	-	-	/	-	-	-	st	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5810	Cooked fillet of fish	+	H+/H-	+	H-	+	<i>L.monocytogenes/L.welshimeri</i>	+	+	+	2H+/H-	+	+	+	PA	PA	+	+	H-(X5 OAA:H+)	+	+	+	+	PA	PA	3
5811	Salmon puff	-	-	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5812	Cooked fillet of fish	+	H+/H-	+	H+/H-	+	<i>L.monocytogenes/L.innocua</i>	+	+	+	H+/H-	+	+	+	PA	PA	+	+	H+/H-	+	+	+	+	PA	PA	3
5813	Ground salmon	+	st	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	-	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
5814	Cod salad	-	-	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5815	Cooked ground salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
5816	Raw marinated salmon	-	H-	+	H-	+	<i>L.welshimeri</i>	-	-	-	H-	+	-	-	NA	NA	-	-	H-	+	-	-	-	-	3	
5817	Raw salmon	-	st	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5818	Ready to cook salmon	+	H+/H-	+	H+/H-	+	<i>L.monocytogenes/L.innocua</i>	+	+	+	H+/H-	+	+	+	PA	PA	+	+	H+/H-	+	+	+	+	PA	PA	3
5819	Cooked fish	+	H-	+	H+/H-	+	<i>L.monocytogenes/L.welshimeri</i>	+	+	+	H+/H-	+	+	+	PA	PA	+	+	H+/H-	+	+	+	+	PA	PA	3
5820	Smoked salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
5821	Smoked salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
5822	Salmon puff	-	-	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
5823	Raw tuna	+	H+/H-	+	H+/H-	+	<i>L.monocytogenes/L.welshimeri</i>	+	+	+	H+/H-	+	+	+	PA	PA	+	+	H+/H-	+	+	+	+	PA	PA	3
6701	Smoked salmon bacon	-	-	-	st	st	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
6702	Smoked salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3	
6703	Marinated salmon	-	-	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
6704	Smoked tuna	-	-	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
6705	Smoked trout	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA	-	-	-	-	-	-	-	3		
6706	Smoked herring	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA	-	-	-	-	-	-	-	3		
6707	Marinated smoked salmon	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA	-	-	-	-	-	-	-	3		
6708	Smoked salmon	-	st	st	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
6709	Smoked haddock	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA	-	-	-	-	-	-	-	3		
6710	Smoked sliced salmon	-	H-	+	H-	+	<i>L.welshimeri</i>	-	-	-	H-	+	-	-	NA	NA	-	-	-	-	-	-	-	3		
6711	Smoked sliced salmon	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA	-	-	-	-	-	-	-	3		
6712	Salmon smoked tartar	-	-	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
6713	Cooked fillet of fish	-	st	-	-	-	/	-	-	-	-	-	-	NA	NA	-	-	-	-	-	-	-	-	3		
6714	Fillet of fish	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA	-	-	-	-	-	-	-	3		

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FISH AND SEAFOOD PRODUCTS

N° Sample	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq														Category		
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C						Half Fraser 22h 30°C + 72h 4°C / Half Fraser 4h 30°C										
			OAA	Palcam	OAA	Palcam			PCR Result		Confirmations		Final result		Agreement Ref/Alt		PCR Result		Confirmations		Final result			Agreement Ref/Alt	
									4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h	4 h	22 h	4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h		4 h	22 h
6715	Raw breaded fish	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
6716	Salmon tarama	-	st	-	-	-	/	-	-	-	st	st	-	-	NA	NA									3
6717	Salmon tarama	-	st	-	-	-	/	-	-	-	st	st	-	-	NA	NA									3
6718	Smoked salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
194	Smoked salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
231	Prawns	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
232	Prawns	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
233	Crayfish	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
234	Prawns	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
235	Prawns	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
236	Seafood cocktail	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
237	Fillet of Cod	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
238	Fillet of Cod	+	1H+	6+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
243	Sliced salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
244	Marinated salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
245	Salmon tartar	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
246	Smoked salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
247	Smoked salmon bacon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
248	Smoked trout	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
249	Smoked tuna	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
250	Smoked haddock	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
277	Smoked salmon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	3
343	Prawns	-	1H-	-	-	-		-	-	-	-	-	-	-	NA	NA								3	
344	Prawns	-	st	st	-	-	/	-	-	-	-	-	-	-	NA	NA								3	
345	Prawns	-	st	-	-	-	/	-	-	-	-	-	-	-	NA	NA								3	
346	Prawns	-	st	-	-	-	/	-	-	-	-	-	-	-	NA	NA								3	
347	Prawns	-	-	-	st	st	/	-	-	-	-	-	-	-	NA	NA								3	

READY-TO-EAT, READY-TO-REHEAT OR READY-TO-COOK PRODUCTS																									
N° Sample	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq																Category
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C				Half Fraser 22h 30°C + 72h 4°C / Half Fraser 4h 30°C												
			OAA	Palcam	OAA	Palcam			PCR Result		Confirmations		Final result		Agreement Ref/Alt		PCR Result		Confirmations		Final result		Agreement Ref/Alt		
									4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h	4 h	22 h	4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h	4 h	22 h	
3351	Ham and butter sandwich	-	st	-	st	st	/	-	-	-	st	-	-	-	NA	NA								4	
3359	Pancake with mushrooms	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
3360	Croque Kebab	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
4102	Roll of puff pastry	-	st	-	-	-	/	-	-	-	st	-	-	-	NA	NA								4	
4103	Deli salad (vegetables)	-	st	-	-	-	/	-	-	-	st	-	-	-	NA	NA								4	
4104	Deli salad(pasta)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA								4	
4105	Cubes of sweet potatoes	+	H+/H-	+	H+/H-	+	<i>L.innocua/ L.monocytogenes</i>	+	+	+	H+/H-	+	+	+	PA	PA	+	+	H+/H-	+	+	+	PA	PA	4
4106	Roll of puff pastry	-	st	-	-	-	/	-	-	-	st	-	-	-	NA	NA								4	
4107	Sliced vegetables	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	-	+	+	PA	PA	+	+	H+	+1col	+	+	PA	PA	4
4108	Sandwich (ham, goat cheese)	-	st	-	-	-	/	-	-	-	st	-	-	-	NA	NA								4	
4109	Ready to cook plate (poultry)	+	H+/H-	-	H+/H-	+	<i>L.innocua/ L.monocytogenes</i>	+	+	+	H+/H-	-	+	+	PA	PA	+	+	H+/H-	+	+	+	PA	PA	4
4617	Cake with cream	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA								4	
4621	Sliced tomatoes	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA								4	
4622	Salad	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA								4	
4623	Couscous seeds	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
4624	Spinach	+	-	-	H+	+	<i>L.monocytogenes</i>	+	-	+	-(Few H+ colonies at 72H)	-	-	+	ND _{FN(alt)}	PA	+	+	H+	+	+	+	PA	PA	4
4625	Puck leek and potatoes	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA								4	
4626	Paella	-	H-?	-	H-	+	<i>L.innocua</i>	-	-	-	1H-? (1col H+ at 72H)	-	-	-	NA _{FN(alt)}	NA _{FN(alt)}	-	-	H-/1H+	+	-	-	NA _{FN(alt)}	NA _{FN(alt)}	4
4627	Green pepper dressing	-	st	-	-	-	/	-	-	-	st	st	-	-	NA	NA								4	
4628	Frozen cauliflower purée	-	st	-	st	st	/	-	-	-	st	-	-	-	NA	NA								4	
4629	Green beans purée	+	H+	-	H+	-	<i>L.monocytogenes</i>	+	+	+	H+	-	+	+	PA	PA	+	+	H+	-	+	+	PA	PA	4
4630	Stuffed tomatoes	-	-	+2col	H-	+	<i>L.welshimeri</i>	-	-	-	-	-	-	-	NA	NA								4	
4631	Deli salad (fish)	+	-	+2col	H+	+	<i>L.monocytogenes</i>	+	+	+	H+/H-?	+/-	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
4632	Sandwich (poultry, tomatoes)	+	H+/1H-	+	H+	+	<i>L.welshimeri/ L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
4633	Pastry with coffee cream	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
4634	Deli salad (vegetables, egg, ham)	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA								4	
4635	Pastry	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA								4	
4636	Sandwich (ham, egg)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA								4	
4637	Sandwich (ham, butter)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA								4	
4638	Sandwich (ham, cheese)	-	st	-	-	-	/	-	-	-	st	st	-	-	NA	NA								4	
4639	Cake	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA								4	
4748	Toasted croque monsieur	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
4749	Roll of puff pastry	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA								4	

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READY-TO-EAT, READY-TO-REHEAT OR READY-TO-COOK PRODUCTS																									
N° Sample	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq														Category		
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C				Half Fraser 22h 30°C + 72h 4°C / Half Fraser 4h 30°C												
			OAA	Palcam	OAA	Palcam			PCR Result		Confirmations		Final result		Agreement Ref/Alt		PCR Result		Confirmations		Final result			Agreement Ref/Alt	
									4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h	4 h	22 h	4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h		4 h	22 h
4750	Grated bread with cheese and bacon	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
4751	Croque Kebab	-	-	-	-	-	/	-	-	-	st	st	-	-	NA	NA									4
4752	Green beans	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA									4
4753	Frozen spinach	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
4754	Frozen vegetables mix	-	-	st	-	-	/	-	-	-	-	st	-	-	NA	NA									4
4756	Frozen vegetables mix	-	st	st	-	-	/	-	-	-	st	st	-	-	NA	NA									4
5077	Sandwich (chicken, vegetables)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5081	Sandwich (smoked chicken, goat cheese, vegetables)	-	H-	3+	H-	+	<i>L.welshimeri</i>	-	-	-	H-	+	-	-	NA	NA									4
5083	Sandwich (ham,egg, tomato)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5086	Sliced cooked eggs	+	1H+	2+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5169	Pizza (cheese)	+	H+	-	H+	-	<i>L.monocytogenes</i>	+	+	+	H+	-	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5170	Pizza (ham, cheese)	+	H++	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5171	Pizza (tuna, vegetables)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5172	Antipasti (artichoke, tomato, olives)	-	st	st	-	-	/	-	-	-	st	-	-	-	NA	NA									4
5173	Frozen carrots	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5174	Frozen artichokes	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5175	Frozen tomatoes	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5176	Frozen tomatoes	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5177	Frozen zucchini	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5178	Frozen peppers	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5179	Dressing (sorrel)	-	st	st	-	-	/	-	-	-	H+	+	-	-	NA _{FN(alt)}	NA _{FN(alt)}									4
5180	Dressing (green pepper)	-	H+	+	H+	+	<i>L.ivanovii</i>	-	-	-	H+	+	-	-	NA _{FN(alt)}	NA _{FN(alt)}									4
5184	Vanilla profiteroles	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5185	Pastry (Paris-Brest)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5186	Cakes (macaron)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5187	Dairy based dessert	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5188	Pastry	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA	4
5256	Deli salad (bacon, potatoes)	-	H-	+	H-	+	<i>L.innocua</i>	-	-	-	H-	+	-	-	NA	NA									4
5257	Deli salad (tuna, pasta)	-	H-	+	H-	+	<i>L.seeligeri</i>	-	-	-	H-	+	-	-	NA	NA									4

ENVIRONMENTAL SAMPLES

N° Sampl e	Product	Global result	ISO 11290-1/A1 method*						Alternative method: Assurance GDS <i>Listeria monocytogenes</i> Tq														Category					
			Half Fraser		Fraser 1		Identification	Result	Half Fraser 22h 30°C				Half Fraser 22h 30°C + 72H 4°C / Half Fraser 4h 30°C															
			OAA	Palcam	OAA	Palca m			PCR Result		Confirmations		Final result		Agreement Ref/Alt		PCR Result		Confirmations		Final result			Agreement Ref/Alt				
									4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h	4 h	22 h	4 h subculture	22 h subculture	ALOA	Palcam	4 h	22 h		4 h	22 h			
6849	Process water (vegetables industry)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA											5	
6850	Process water (vegetables industry)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA												5
6851	Process water (vegetables industry)	-	st	st	st	st	/	-	-	-	-	-	-	-	NA	NA												5
6852	Process water (vegetables industry)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA												5
6853	Process water (fish industry)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
6854	Process water (fish industry)	-	st	st	st	st	/	-	-	-	-	-	-	-	NA	NA												5
6855	Process water (fish industry)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA												5
6856	Process water (fish industry)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA												5
6857	Process water (fish industry)	+	1H+	2+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
251	Process water (ham production)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
252	Process water (fish industry)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
253	Process water (tomato production)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
254	Process water (cheese production)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
255	Process water (fish industry)	+	5H+	1+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
263	Wipe (ready to eat production)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
264	Wipe (ready to eat production)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
265	Wipe (ready to eat production)	-	st	st	st	st	/	-	-	-	st	st	-	-	NA	NA												5
266	Wipe (ready to eat production)	-	st	st	st	st	/	-	-	-	st	-	-	-	NA	NA												5
267	Wipe (ready to eat production)	-	st	st	st	st	/	-	-	-	st	-	-	-	NA	NA												5
268	Dusts (ready to eat production)	-	-	-	st	st	/	-	-	-	-	-	-	-	NA	NA												5
269	Dusts (ready to eat production)	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA												5
270	Dusts (dairy industry)	+	H+	-	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	-	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
271	Dusts (dairy industry)	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA												5
272	Dusts (dairy industry)	-	st	st	-	-	/	-	-	-	st	-	-	-	NA	NA												5
273	Dusts (dairy industry)	+	H+	-	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	1+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
274	Dusts (dairy industry)	-	-	-	-	-	/	-	-	-	-	-	-	-	NA	NA												5
275	Dusts (dairy industry)	-	st	st	-	-	/	-	-	-	st	-	-	-	NA	NA												5
337	Dusts (ready to eat production)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	-	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
338	Dusts (ready to eat production)	+	H+	1+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	2+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
339	Dusts (ready to eat production)	+	H+	5+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
340	Dusts (ready to eat production)	+	H+	1+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	1+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
341	Dusts (ready to eat production)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5
342	Dusts (ready to eat production)	+	H+	+	H+	+	<i>L.monocytogenes</i>	+	+	+	H+	+	+	+	PA	PA	+	+	H+	+	+	+	PA	PA				5



FRESH PRODUCE AND VEGETABLES

Item Type	Sample n°	Reference Method: ISO 11290-1:2017				Alternative Method: Assurance GDS <i>Listeria monocytogenes</i> Tq						
		MOX	ALOA	Confirm	Final Result	PCR		72 Hour Hold PCR		Confirmation	Final Result	Agreement
						4 h	22 h	4 h	22 h			
Sliced apples	1	+	+	+	+	+	+	+	+	+	+	PA
Asparagus 8oz	2	+	+	+	+	+	+	+	+	+	+	PA
Asparagus 8.25 oz	3	-	-	-	-	-	-	/	/	-	-	ND
Medjool Dates 12 oz	4	+	+	+	+	+	+	+	+	+	+	PA
Medjool Dates 16 oz	5	-	-	-	-	-	-	/	/	-	-	ND
Pitted Dates	6	+	+	+	+	+	+	+	+	+	+	PA
Broccoli 8 oz	7	+	+	+	+	+	+	+	+	+	+	PA
Broccoli 12 oz	8	+	+	+	+	+	+	+	+	+	+	PA
Squash 8 oz	9	+	+	+	+	+	+	+	+	+	+	PA
Cantaloupe 8 oz	10	-	-	-	-	-	-	/	/	-	-	ND
Cantaloupe 12 oz	11	-	-	-	-	-	-	/	/	-	-	ND
Carrot & Celery	12	+	+	+	+	+	+	+	+	+	+	PA
Celery Sticks	13	-	-	-	-	-	-	/	/	-	-	ND
Celery Sticks	14	+	+	+	+	+	+	+	+	+	+	PA
Diced Red Onions	15	-	-	-	-	-	-	/	/	-	-	ND
Organic Apple Slices	16	-	-	-	-	-	-	/	/	-	-	ND
Diced Yellow Onions	17	-	-	-	-	-	-	/	/	-	-	ND
Blend Salad	18	-	-	-	-	-	-	/	/	-	-	ND
Salad Kit	19	+	+	+	+	+	+	+	+	+	+	PA
Baby spinach	20	+	+	+	+	+	+	+	+	+	+	PA
Salad mix	21	-	-	-	-	-	-	/	/	-	-	ND
Chopped salad kit	22	+	+	+	+	+	+	+	+	+	+	PA
Chopped Romaine	23	+	+	+	+	+	+	+	+	+	+	PA
Asian Salad kit	24*	+	+	+	+	+	+	+	+	+	+	PA
Classic coleslaw	25	+	+	+	+	+	+	+	+	+	+	PA
Classic romaine	26	+	+	+	+	+	+	+	+	+	+	PA
Summer salad kit	27	-	-	-	-	-	-	/	/	-	-	ND
Fruit medley 10 oz	28	-	-	-	-	-	-	/	/	-	-	ND

1. The confirmation procedures included a Gram stain, Beta-Hemolysis, L-Rhamnose and D-Xylose.

*Indicates a naturally contaminated sample

FRESH PRODUCE AND VEGETABLES

Item Type	Sample n°	Reference Method: ISO 11290-1:2017				Alternative Method: Assurance GDS <i>Listeria monocytogenes</i> Tq						
		MOX	ALOA	Confirm	Final Result	PCR		72 Hour Hold PCR		Confirmation	Final Result	Agreement
						4 h	22 h	4 h	22 h			
Fruit medley 24 oz	29	-	-	-	-	-	-	/	/	-	-	ND
Fruit medley 48 oz	30	+	+	+	+	+	+	+	+	+	+	PA
Mixed berries 24 oz	31	-	-	-	-	-	-	/	/	-	-	ND
Pineapple chunks	32	+	+	+	+	+	+	+	+	+	+	PA
Fresh Squash	33	-	-	-	-	-	-	/	/	-	-	ND
Fresh Cauliflower	34	+	+	+	+	+	+	+	+	+	+	PA
Sweet potato	35	-	-	-	-	-	-	/	/	-	-	ND
Honeydew chunks	36	-	-	-	-	-	-	/	/	-	-	ND
Seedless watermelon	37	-	-	-	-	-	-	/	/	-	-	ND
Jicama sticks	38	-	-	-	-	-	-	/	/	-	-	ND
Baby spinach	39	-	-	-	-	-	-	/	/	-	-	ND
Spring mix	40	-	-	-	-	-	-	/	/	-	-	ND
Spring mix	41	+	+	+	+	+	+	+	+	+	+	PA
Garden salad	42	-	-	-	-	-	-	/	/	-	-	ND
Garden salad	43	+	+	+	+	+	+	+	+	+	+	PA
Coleslaw salad	44	+	+	+	+	+	+	+	+	+	+	PA
Baby carrots	45	-	-	-	-	-	-	/	/	-	-	ND
Romaine salad	46	-	-	-	-	-	-	/	/	-	-	ND
Blended salad	47	-	-	-	-	-	-	/	/	-	-	ND
Shredded carrots	48	+	+	+	+	+	+	+	+	+	+	PA
Vegetable medley	49	-	-	-	-	-	-	/	/	-	-	ND
Veggie blend salad	50	+	+	+	+	+	+	+	+	+	+	PA
Broccoli pearls	51	+	+	+	+	+	+	+	+	+	+	PA
Brussel sprouts	52	+	+	+	+	+	+	+	+	+	+	PA
Cauliflower pearls	53	-	-	-	-	-	-	/	/	-	-	ND
Snap peas	54	+	+	+	+	+	+	+	+	+	+	PA
Turnip greens	55	-	-	-	-	-	-	/	/	-	-	ND
Mango chunks	56	+	+	+	+	+	+	+	+	+	+	PA

1. The confirmation procedures included a Gram stain, Beta-Hemolysis, L-Rhamnose and D-Xylose.

FRESH PRODUCE AND VEGETABLES

Item Type	Sample n°	Reference Method: ISO 11290-1:2017				Alternative Method: Assurance GDS <i>Listeria monocytogenes</i> Tq						
		MOX	ALOA	Confirm	Final Result	PCR		72 Hour Hold PCR		Confirmation	Final Result	Agreement
						4 Hour	22 Hour	4 Hour	22 Hour			
Sweet peppers	57	-	-	-	-	-	-	/	/	-	-	ND
Pineapple cored	58	+	+	+	+	+	+	+	+	+	+	PA
Pineapple chunks	59	+	+	+	+	+	+	+	+	+	+	PA
Chopped salad	60	+	+	+	+	+	+	+	+	+	+	PA
Tomatoes	61	-	-	-	-	-	-	/	/	-	-	ND
Watermelon chunks	62	-	-	-	-	-	-	/	/	-	-	ND
Mixed berries	63*	+	+	+	+	+	+	+	+	+	+	PA
Squash	64	-	-	-	-	-	-	/	/	-	-	ND
Zucchini	65	-	-	-	-	-	-	/	/	-	-	ND
Mixed melon	66*	+	+	+	+	+	+	+	+	+	+	PA

1. The confirmation procedures included a Gram stain, Beta-Hemolysis, L-Rhamnose and D-Xylose.

*Indicates a naturally contaminated sample

Appendix 6 - RLOD: raw data

Sausage

Listeria monocytogenes Ad669

Total viable count: 2.0 x 10²/g

Sample N°	Level	Inoculation level (cfu/25g)	ISO 11290-1 method*					Assurance GDS <i>Listeria monocytogenes</i> Tq					
			Half Fraser		Fraser		<i>Listeria monocytogenes</i> result	Positive/Total	PCR result-22 h	Confirmation		Final result	Positive/Total
			OAA	Palcam	OAA	Palcam				OAA	Palcam		
3524	0	/	st	st	st	st	-	0/6	-	st	st	-	0/6
3525			st	-	-	-	-		-	st	st	-	
3526			-	-	-	-	-		-	-	-	-	
3527			st	st	st	st	-		-	st	st	-	
3528			st	st	st	st	-		-	st	st	-	
3529			st	-	st	st	-		-	st	st	-	
3664	1	0.3	st	st	st	st	-	0/6	-	st	st	-	0/6
3665			st	st	st	st	-		-	st	st	-	
3666			st	st	st	st	-		-	st	st	-	
3667			st	st	st	st	-		-	st	st	-	
3668			st	st	st	st	-		-	-	st	-	
3669			st	st	st	st	-		-	st	st	-	
3670	2	0.5	H+	+	/	/	+	2/6	+	H+	+	+	2/6
3671			-	-	-	-	-		-	-	-	-	
3672			st	st	st	st	-		-	st	st	-	
3673			st	st	st	st	-		-	st	st	-	
3674			st	st	st	st	-		-	st	st	-	
3675			H+	+	/	/	+		+	H+	+	+	
3676	3	1.1	H+	+	/	/	+	5/6	+	H+	+	+	5/6
3677			H+	+	/	/	+		+	H+	+	+	
3678			H+	+	/	/	+		+	H+	+	+	
3679			H+	+	/	/	+		+	H+	+	+	
3680			H+	+	/	/	+		+	H+	+	+	
3681			st	st	st	st	-		-	st	st	-	
3682	4	2.2	H+	+	/	/	+	6/6	+	H+	+	+	6/6
3683			H+	+	/	/	+		+	H+	+	+	
3684			H+	+	/	/	+		+	H+	+	+	
3685			H+	+	/	/	+		+	H+	+	+	
3686			H+	+	/	/	+		+	H+	+	+	
3687			H+	+	/	/	+		+	H+	+	+	

* Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

Fresh goat cheese

Total viable count: 1.8 x 10⁷/g

Listeria monocytogenes Ad618

Sample N°	Level	Inoculation level (cfu/25g)	ISO 11290-1 method [♦]					Assurance GDS <i>Listeria monocytogenes</i>							
			Half Fraser		Fraser		<i>Listeria monocytogenes</i> result	Positive/Total	PCR result-4 h	PCR result-22 h	Confirmation		Final result	Positive/Total	
			OAA	Palcam	OAA	Palcam					OAA	Palcam			
3518	0	/	st	-	-	-	-	0/6	-	-	-	-	-	0/6	
3519			-	-	-	-	-		-	-	-	-			
3520			-	-	-	-	-		-	-	-	-			
3521			-	-	-	-	-		-	-	-	-			
3522			-	-	-	-	-		-	-	-	-			
3523			-	-	-	-	-		-	-	-	-			
3576	1	0.2	H+	+	/	/	+	1/6	+	+	H+	+	+	1/6	
3577			-	-	-	-	-		-	-	-	-	-		
3578			-	-	-	-	-		-	-	-	St	-		-
3579			-	-	-	-	-		-	-	-	-	-		-
3580			-	-	-	-	-		-	-	-	-	-		-
3581			-	St	-	-	-		-	-	-	-	-		-
3582	2	0.4	-	St	-	-	-	2/6	-	-	-	-	-	2/6	
3583			H+	+	/	/	+		+	+	H+	+	+		
3584			-	-	-	-	-		-	-	-	-	-		-
3585			-	St	-	-	-		-	-	-	-	-		-
3586			-	-	-	-	-		-	-	-	-	-		-
3587			H+	+	/	/	+		+	+	H+	+	+		
3588	3	0.8	-	-	-	-	-	4/6	-	-	-	-	-	4/6	
3589			H+	+	/	/	+		+	+	H+	+	+		
3590			H+	+	/	/	+		+	+	H+	+	+		
3591			-	-	-	-	-		-	-	-	St	-		-
3592			H+	+	/	/	+		+	+	H+	+	+		
3593			H+	+	/	/	+		+	+	H+	+	+		
3594	4	1.6	H+	+	/	/	+	4/6	+	+	H+	+	+	4/6	
3595			H+	+	/	/	+		+	+	H+	+	+		
3596			H+	+	/	/	+		+	+	H+	+	+		
3597			-	-	-	-	-		-	-	-	-	-		-
3598			-	St	-	-	-		-	-	-	-	-		-
3599			H+	+	/	/	+		+	+	H+	+	+		
3913	5	5	H+	+	/	/	+	6/6	+	+	H+	+	+	6/6	
3914			H+	+	/	/	+		+	+	H+	+	+		
3915			H+	+	/	/	+		+	+	H+	+	+		
3916			H+	+	/	/	+		+	+	H+	+	+		
3917			H+	+	/	/	+		+	+	H+	+	+		
3918			H+	+	/	/	+		+	+	H+	+	+		

♦ Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

Smoked salmon

Total viable count: 6,0.10²/g

Listeria monocytogenes

Ad670

Sample N°	Level	Inoculation level (cfu/25g)	ISO 11290-1 method*					GDS <i>Listeria monocytogenes</i>						
			Half Fraser		Fraser		<i>Listeria monocytogenes</i> result	Positive/ Total	PCR result-4H	PCR result-22H	Confirmation		Final result	Positive/Total
			OAA	Palcam	OAA	Palcam					OAA	Palcam		
5745	0	0	-	-	-	-	-	0/6	-	-	-	-	-	0/6
5746			-	-	-	-	-		-	-	-	-	-	
5747			-	-	-	-	-		-	-	-	-	-	
5748			-	-	st	st	-		-	-	-	-	-	
5749			-	-	st	st	-		-	-	-	-	-	
5750			-	-	-	-	-		-	-	-	-	-	
6151	1	0.2	H-	+(L.welshimeri)	H-	+(L.welshimeri)	-	0/6	-	-	H-	+(camp-)	-	0/6
6152			st	st	H-	+	-		-	-	st	st	-	
6153			st	st	st	st	-		-	-	-	-	-	
6154			st	st	st	st	-		-	-	st	-	-	
6155			st	st	st	st	-		-	-	-	st	-	
6156			st	st	-	-	-		-	-	-	-	-	
6157	2	0.4	st	st	st	st	-	1/6	-	-	st	st	-	1/6
6158			st	st	st	st	-		-	-	-	-	-	
6159			-	-	-	-	-		-	-	-	1col +/- (gram-)	-	
6160			-	st	-	-	-		-	-	-	-	-	
6161			st	1col (L.welshimeri)	H-	+(L.welshimeri)	-		-	-	3H-	+(camp-)	-	
6162			+	+	/	/	+		+	+	+	+	+	
6163	3	0.7	+	+	/	/	+	4/6	+	+	+	+	+	4/6
6164			+	+	/	/	+		+	+	+	+		
6165			H-	+(L.welshimeri)	H-	+(L.welshimeri)	-		-	-	H-	+(camp-)	-	
6166			st	st	st	st	-		-	-	st	st	-	
6167			+	+	/	/	+		+	+	+	+	+	
6168			+	+	/	/	+		+	+	+	+	+	
6169	4	1.4	+	+	/	/	+	6/6	+	+	+	+	+	6/6
6170			+	+	/	/	+		+	+	+	+		
6171			+	+	/	/	+		+	+	+	+		
6172			+	+	/	/	+		+	+	+	+		
6173			+	+	/	/	+		+	+	+	+		
6174			+	+	/	/	+		+	+	+	+		

* Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

Potatoes purée
 Total viable count: 2.0 x 10²/g
 Listeria monocytogenes Ad279

Sample N°	Level	Inoculation level (cfu/25g)	ISO 11290-1 method*					GDS Listeria monocytogenes							
			Half Fraser		Fraser		Listeria monocytogenes result	Positive/ Total	PCR result-4H	PCR result-22H	Confirmation		Final result	Positive/Total	
			OAA	Palcam	OAA	Palcam					OAA	Palcam			
4225	0	/	st	-	-	-	-	0/6	-	-	st	-	-	0/6	
4226			st	-	-	-	-		-	-	st	-	-		
4227			st	-	-	-	-		-	-	st	-	-		
4228			st	-	-	-	-		-	-	st	-	-		
4229			st	-	-	-	-		-	-	st	-	-		
4230			st	-	-	-	-		-	-	st	-	-		
4231	1	0.4	st	-	-	-	-	1/6	-	-	st	-	-	1/6	
4232			st	-	-	-	-		-	-	st	-	-		
4233			st	-	-	-	-		-	-	st	-	-		
4234			H+	+	/	/	+		+	+	H+	+	+		
4235			st	-	-	-	-		-	-	-	st	-		-
4236			-	-	-	-	-		-	-	-	st	-		-
4237	2	0.8	H+	+	/	/	+	5/6	+	+	H+	+	+	5/6	
4238			H+	+	/	/	+		+	+	H+	+	+		
4239			st	-	-	-	-		-	-	-	st	-		-
4240			H+	+	/	/	+		+	+	H+	+	+		
4241			H+	+	/	/	+		+	+	H+	+	+		
4242			H+	+	/	/	+		+	+	H+	+	+		
4243	3	1.6	H+	+	/	/	+	4/6	+	+	H+	+	+	4/6	
4244			st	-	-	-	-		-	-	st	-	-		
4245			H+	+	/	/	+		+	+	H+	+	+		
4246			H+	+	/	/	+		+	+	H+	+	+		
4247			H+	+	/	/	+		+	+	H+	+	+		
4248			st	-	-	-	-		-	-	-	st	-		-
4249	4	3.2	H+	+	/	/	+	5/6	+	+	H+	+	+	5/6	
4250			H+	+	/	/	+		+	+	H+	+	+		
4251			H+	+	/	/	+		+	+	H+	+	+		
4252			H+	+	/	/	+		+	+	H+	+	+		
4253			st	-	-	-	-		-	-	-	st	-		-
4254			H+	+	/	/	+		+	+	H+	+	+		
4396	5	4.9	H+	+	H+	+	+	6/6	+	+	H+	+	+	6/6	
4397			H+	+	H+	+	+		+	+	H+	+	+		
4398			H+	+	H+	+	+		+	+	H+	+	+		
4399			H+	+	H+	+	+		+	+	H+	+	+		
4400			H+	+	H+	+	+		+	+	+	H+	+		+
4401			H+	+	H+	+	+		+	+	+	H+	+		+

* Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

Process water

Total viable count: 460 CFU/g
Listeria monocytogenes Ad631

Sample N°	Level	Inoculation level (cfu/25g)	ISO 11290-1 method*					Assurance GDS <i>Listeria monocytogenes</i>						
			Half-Fraser		Fraser		<i>Listeria monocytogenes</i> result	Positive/ Total	PCR result-4 h	PCR result-22 h	Confirmation		Final result	Positive/Total
			OAA	Palcam	OAA	Palcam					OAA	Palcam		
5751	0	0	st	st	st	st	-	0/6	-	-	st	st	-	0/6
5752			st	st	st	st	-		-	-	st	st	-	
5753			st	st	st	st	-		-	-	st	st	-	
5754			st	st	st	st	-		-	-	st	st	-	
5755			st	st	st	st	-		-	-	st	st	-	
5756			st	st	st	st	-		-	-	st	st	-	
6595	1	0.2	st	st	st	st	-	1/6	-	-	st	st	-	1/6
6596			st	st	st	st	-		-	-	st	st	-	
6597			st	st	st	st	-		-	-	st	st	-	
6598			st	st	st	st	-		-	-	st	st	-	
6599			H+	+	/	/	+		+	+	H+	+	+	
6600	st	st	st	st	-	-	-	st	st	-				
6601	2	0.4	st	st	st	st	-	3/6	-	-	st	st	-	3/6
6602			H+	+	/	/	+		+	+	H+	+	+	
6603			H+	+	/	/	+		+	+	H+	+	+	
6604			H+	+	/	/	+		+	+	H+	+	+	
6605			st	st	st	st	-		-	-	st	st	-	
6606			st	st	st	st	-		-	-	st	st	-	
6607	3	0.7	st	st	st	st	-	1/6	-	-	st	st	-	1/6
6608			st	st	-	-	-		-	-	st	st	-	
6609			st	st	st	st	-		-	-	st	st	-	
6610			H+	+	/	/	+		+	+	H+	+	+	
6611			st	st	st	st	-		-	-	st	st	-	
6612			st	st	st	st	-		-	-	st	st	-	
6613	4	1.4	H+	+	/	/	+	6/6	+	+	H+	+	+	6/6
6614			H+	+	/	/	+		+	+	H+	+	+	
6615			H+	+	/	/	+		+	+	H+	+	+	
6616			H+	+	/	/	+		+	+	H+	+	+	
6617			H+	+	/	/	+		+	+	H+	+	+	
6618			H+	+	/	/	+		+	+	H+	+	+	

* Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)

Pre-Cut Bagged Spinach				
<i>Listeria monocytogenes</i> CW1554				
Low Level: 0.37 CFU/25 g				
Sample #	Assurance GDS [®] <i>Listeria monocytogenes</i> Tq			ISO 11290-1:2017 Result ¹
	PCR Presumptive Result		Confirmed	
	4 Hour	22 Hour	ISO 11290-1:2017 Result	
1	+	+	+	+
2	-	-	-	-
3	-	-	-	-
4	+	+	+	+
5	-	-	-	-
6	-	-	-	-
7	+	+	+	+
8	-	-	-	-
9	+	+	+	+
10	-	-	-	-
11	-	-	-	-
12	-	-	-	-
13	+	+	+	+
14	-	-	-	-
15	-	-	-	-
16	-	-	-	-
17	+	+	+	+
18	-	-	-	-
19	-	-	-	-
20	+	+	+	+
Total	7/20	7/20	7/20	7/20
High Level: 4.1 CFU/25 g				
1	+	+	+	+
2	+	+	+	+
3	+	+	+	+
4	+	+	+	+
5	+	+	+	+
Total	5/5	5/5	5/5	5/5
Uninoculated				
1	-	-	-	-
2	-	-	-	-
3	-	-	-	-
4	-	-	-	-
5	-	-	-	-
Total	0/5	0/5	0/5	0/5

1. A paired analysis was conducted.
2. APC=1.2 x 10⁵ CFU/g

Appendix 7 - Inclusivity and exclusivity: raw data

INCLUSIVITY									
Strain	Species	Reference	Origin	Molecular serotypes	Enrichment broth	Inoculation level (cfu:225ml)	PCR	Streaking on OAA or Palcam	
<i>Listeria</i>	<i>monocytogenes</i>	1011/1410	Frozen broccoli	II a	Half Fraser	1	-	-	
						20	-	-	
						54	-	-	
					Half Fraser +milk	40	-	+	
						Half Fraser (2013)	13	-	-
						Half Fraser +milk (2013)	13	+	+
BHI (2013)	13	+	/						
<i>Listeria</i>	<i>monocytogenes</i>	153	Soft cheese (Munster)	VI b	Half Fraser	4	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	1972/2399	Pie with mushrooms	VI b	Half Fraser	13	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	1973/2400	Egg and ham pastry (Quiche Lorraine)	VI b	Half Fraser	3	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	2760/3145	Raw pork meat	II a	Half Fraser	9	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	32.183	Croque Monsieur	II b	Half Fraser	14	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	38/181	Toulouse sausages	II a	Half Fraser	17	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	5721/6179	Smoked bacon	IV b	Half Fraser	16	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	7111/7516	Pâté (Rillettes)	IV b	Half Fraser	8	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	850/109	Nordic salad	II a	Half Fraser	5	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	877/113	Food industry environment	II a	Half Fraser	10	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	913/1 048	Black pudding	IV b	Half Fraser	2	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C022	Merguez	II a	Half Fraser	18	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C036	Poultry (guinea)	II a	Half Fraser	7	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C039	Sausages (Diots de Savoie)	II a	Half Fraser	8	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C040	Pâté	IV b	Half Fraser	11	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C041	Sausage	La	Half Fraser	11	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C042	Toulouse sausage	IV b	Half Fraser	9	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C043	Smoked bacon	II a	Half Fraser	10	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C044	Poultry (Duck)	II b	Half Fraser	11	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C052	Poultry	II b	Half Fraser	13	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00C053	Poultry	II a	Half Fraser	9	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00E082	Environment (smoked salmon)	II a	Half Fraser	23	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00L097	Milk	II a	Half Fraser	15	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00M009	Smoked salmon	II a	Half Fraser	14	+	/	
<i>Listeria</i>	<i>monocytogenes</i>	A00M032	Smoked salmon	IV b	Half Fraser	7	+	/	

INCLUSIVITY								
Strain	Species	Reference	Origin	Molecular serotypes	Enrichment broth	Inoculation level (cfu:225ml)	PCR	Streaking on OAA or Palcam
<i>Listeria</i>	<i>monocytogenes</i>	Ad235	Poultry	II b	Half Fraser	6	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad249	Environment (Meat product)	II b	Half Fraser	11	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad253	Semi-hard cheese	II b	Half Fraser	7	-	-
						35	-	-
						25	-	+1 colony
					Half Fraser +milk	40	-	+
					Half Fraser (2013)	14	-	-
					Half Fraser +milk (2013)	14	-	+
					BHI (2013)	14	-	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad260	Semi-hard cheese	II a	Half Fraser	3	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad265	Pork	II b	Half Fraser	14	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad266	Poultry	II a	Half Fraser	6	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad267	Fermented sausage	II b	Half Fraser	5	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad268	Cured ham	IV b	Half Fraser	4	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad270	Fermented sausage	IV b	Half Fraser	4	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad273	Cured delicatessen	II b	Half Fraser	6	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad274	Ready-to-eat food (Asiatic meal)	II a	Half Fraser	4	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad285	Ready-to-eat food	La	Half Fraser	15	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad494	Ready-to-eat food (Piemontaise salad)	II a	Half Fraser	15	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad534	Fruits	II b	Half Fraser	10	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad544	Cooked vegetables	II a	Half Fraser	15	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad546	Flour	II a	Half Fraser	12	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad548	Environment (Seafood)	II a	Half Fraser	14	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad551	Environment (Pastry environment)	II a	Half Fraser	9	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad618	Soft cheese (Munster)	IV b	Half Fraser	12	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad623	Bread crumbs	II b	Half Fraser	12	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad625	Environment (Dairy industry)	IV b	Half Fraser	9	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad626	Gorgonzola	II a	Half Fraser	4	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad630	Semi-hard cheese (Cantal)	II a	Half Fraser	9	+	/
<i>Listeria</i>	<i>monocytogenes</i>	Ad665	Raw milk	II a	Half Fraser	3	+	/

EXCLUSIVITY					
Strain	Species	Reference	Origin	Inoculation level (cfu/ml)	PCR result
<i>Listeria</i>	<i>grayi</i>	ATCC19120	/	1.00 10 ⁶	-
<i>Listeria</i>	<i>grayi</i>	CIP76124	/	4.00 10 ⁶	-
<i>Listeria</i>	<i>innocua</i>	1	Smoked salmon	1.20 10 ⁵	-
<i>Listeria</i>	<i>innocua</i>	Ad 658	Gorgonzola	9.20 10 ⁵	-
<i>Listeria</i>	<i>ivanovii</i>	Ad466	Raw veal meat	1.10 10 ⁵	-
<i>Listeria</i>	<i>ivanovii</i>	Ad662	Environment (dairy industry)	1.50 10 ⁵	-
<i>Listeria</i>	<i>seeligeri</i>	Ad649	Cheese	1.20 10 ⁵	-
<i>Listeria</i>	<i>seeligeri</i>	BR1	Trout	1.40 10 ⁵	-
<i>Listeria</i>	<i>welshimeri</i>	Ad1276	Environment (Slaughterhouse)	1.60 10 ⁵	-
<i>Listeria</i>	<i>welshimeri</i>	Ad1175	Ready-to-eat-food	7.20 10 ⁴	-
<i>Bacillus</i>	<i>cereus</i>	Ad 465	Salmon Terrine	4.0 10 ⁵	-
<i>Bacillus</i>	<i>circulans</i>	Ad 769	Vegetables	6.0 10 ⁵	-
<i>Bacillus</i>	<i>coagulans</i>	Ad 731	Dairy product	2.0 10 ⁵	-
<i>Bacillus</i>	<i>licheniformis</i>	Ad 978	Dairy product	1.0 10 ⁵	-
<i>Bacillus</i>	<i>pumilus</i>	Ad 284	Ready-to-eat	8.0 10 ⁴	-
<i>Brochotrix</i>	<i>compressis</i>	CIP 1029205	Environment	3.0 10 ⁵	-
<i>Carnobacterium</i>	<i>piscicola</i>	Ad 369	Raw milk	4.6 10 ⁵	-
<i>Enterococcus</i>	<i>durans</i>	Ad 149	Ham	6.0 10 ³	-
<i>Enterococcus</i>	<i>faecalis</i>	89L326	Soft cheese (Vacherin)	4.6 10 ⁴	-
<i>Lactobacillus</i>	<i>brevis</i>	86L126	Ham	2.0 10 ⁵	-
<i>Lactobacillus</i>	<i>curvatus</i>	Ad 380	Delicatessen	5.0 10 ⁴	-
<i>Lactobacillus</i>	<i>sakei</i>	Ad 473	Ham	2.0 10 ⁵	-
<i>Leuconostoc</i>	<i>camosum</i>	Ad 411	Ham	7.4 10 ⁴	-
<i>Leuconostoc</i>	<i>citreum</i>	Ad 396	Ham	4.8 10 ⁴	-
<i>Micrococcus</i>	<i>luteus</i>	Ad 432	Cocktail	2.0 10 ⁵	-
<i>Staphylococcus</i>	<i>aureus</i>	Ad 165	Smoked delicatessen	2.5 10 ⁵	-
<i>Staphylococcus</i>	<i>epidermis</i>	Ad 931	Fruits	7.5 10 ⁴	-
<i>Staphylococcus</i>	<i>haemoliticus</i>	Ad 989	Dairy product	2.0 10 ⁵	-
<i>Streptococcus</i>	<i>bovis</i>	91L518	Dairy product	1.2 10 ⁵	-
<i>Streptococcus</i>	<i>salivarius</i>	Ad 441	Dairy product	4.0 10 ⁴	-

Exclusivity - Renewal study											
No.	Genus	Species	Sensu	Reference	Origin	Level before incubation (CFU/mL)	Assurance® GDS for <i>Listeria monocytogenes</i> Tq BPW - 24 h at 37°C				Comment
							PCR Assurance GDS <i>Listeria monocytogenes</i> Tq		Direct streaking		
							Target Cq	Result	O&A	Palcam	
1	<i>Listeria</i>	<i>grayi</i>	Lato	Ad2415	Rillette	6.60x10 ⁴	N/A	-	-	-	Troubled enrichment after incubation
2	<i>Listeria</i>	<i>grayi</i>	Lato	Ad1295	Spinach	6.00x10 ⁴	N/A	-	-	st	Troubled enrichment after incubation
3	<i>Listeria</i>	<i>rocourtiae</i>	Lato	DSM22097	Lettuce	1.40x10 ⁴	N/A	-	-	+	/
4	<i>Listeria</i>	<i>grayi</i>	Lato	Ad3288	Smoked bacon	8.40x10 ⁴	N/A	-	H- (light)	-	/
5	<i>Listeria</i>	<i>grayi</i>	Lato	Ad3205	Chicken nuggets	9.00x10 ⁴	N/A	-	H- (light)	+	/
6	<i>Listeria</i>	<i>fleischmanii</i>	Lato	DSM24998	Hard cheese	6.60x10 ⁴	N/A	-	H-	+ (light)	/
7	<i>Listeria</i>	<i>riparia</i>	Lato	DSM26685	Running water	2.00x10 ⁴	N/A	-	H- (white center)	+	/
8	<i>Listeria</i>	<i>farberi</i>	Stricto	LMG31917	Soil	7.60x10 ⁴	N/A	-	H-	+	/
9	<i>Listeria</i>	<i>immobilis</i>	Stricto	LMG31920	Soil	3.00x10 ⁴	N/A	-	-	+	/
10	<i>Listeria</i>	<i>marthii</i>	Stricto	DSM23813	Soil	5.20x10 ⁴	N/A	-	H-	+	/
11	<i>Listeria</i>	<i>cornellensis</i>	Lato	DSM26689	Water	4.60x10 ⁵	N/A	-	st	st	Troubled enrichment after incubation
12	<i>Listeria</i>	<i>grandensis</i>	Lato	DSM26688	Water	6.20x10 ⁵	N/A	-	st	st	Troubled enrichment after incubation

Appendix 8 - Inter-laboratory study: raw data

Collaborator A
 Aerobic mesophilic flora: 4,4 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
A3	-	-	-	-	-	-	-	-	-	NA
A5	-	-	-	-	-	-	-	-	-	NA
A9	-	-	-	-	-	-	-	-	-	NA
A13	-	-	-	-	-	-	-	-	-	NA
A14	-	-	-	-	-	-	-	-	-	NA
A18	-	-	-	-	-	-	-	-	-	NA
A22	-	-	-	-	-	-	-	-	-	NA
A23	-	-	-	-	-	-	-	-	-	NA
A2	+	+	+	+	+	+	+	+	+	PA
A7	+	+	+	+	+	+	+	+	+	PA
A8	+	+	+	+	+	+	+	+	+	PA
A11	+	+	+	+	+	+	+	+	+	PA
A15	+	+	+	+	+	+	+	+	+	PA
A17	+	+	+	+	+	+	+	+	+	PA
A21	+	+	+	+	+	+	+	+	+	PA
A24	+	+	-	-	+	+	+	+	+	PA
A1	+	+	+	+	+	+	+	+	+	PA
A4	+	+	+	+	+	+	+	+	+	PA
A6	+	+	+	+	+	+	+	+	+	PA
A10	+	+	+	+	+	+	+	+	+	PA
A12	+	+	+	+	+	+	+	+	+	PA
A16	+	+	+	+	+	+	+	+	+	PA
A19	+	+	+	+	+	+	+	+	+	PA
A20	+	+	+	+	+	+	+	+	+	PA

Collaborator B

Aerobic mesophilic flora: 5,0 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
B3	-	-	-	-	-	-	-	-	-	NA
B5	-	-	-	-	-	-	-	-	-	NA
B9	-	-	-	-	-	-	-	-	-	NA
B13	-	-	-	-	-	-	-	-	-	NA
B14	-	-	-	-	-	-	-	-	-	NA
B18	-	-	-	-	-	-	-	-	-	NA
B22	-	-	-	-	-	-	-	-	-	NA
B23	-	-	-	-	-	-	-	-	-	NA
B2	+	+	+	+	+	+	+	+	+	PA
B7	+	+	+	+	+	+	+	+	+	PA
B8	-	-	-	-	-	-	-	-	-	NA
B11	+	+	+	+	+	+	+	+	+	PA
B15	+	+	+	+	+	+	+	+	+	PA
B17	+	+	+	+	+	+	+	+	+	PA
B21	+	+	+	+	+	+	+	+	+	PA
B24	+	+	+	+	+	+	+	+	+	PA
B1	+	+	+	+	+	+	+	+	+	PA
B4	+	+	+	+	+	+	+	+	+	PA
B6	+	+	+	+	+	+	+	+	+	PA
B10	+	+	+	+	+	+	+	+	+	PA
B12	+	+	+	+	+	+	+	+	+	PA
B16	+	+	+	+	+	+	+	+	+	PA
B19	+	+	+	+	+	+	+	+	+	PA
B20	+	+	+	+	+	+	+	+	+	PA

Collaborator C

Aerobic mesophilic flora: 1,7 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
C3	-	-	-	-	-	-	/	/	-	NA
C5	-	-	-	-	-	-	/	/	-	NA
C9	-	-	-	-	-	-	/	/	-	NA
C13	-	-	-	-	-	-	/	/	-	NA
C14	-	-	-	-	-	-	/	/	-	NA
C18	-	-	-	-	-	-	/	/	-	NA
C22	-	-	-	-	-	-	/	/	-	NA
C23	-	-	-	-	-	-	/	/	-	NA
C2	+	+	+	+	+	+	+	+	+	PA
C7	+	+	+	+	+	+	+	+	+	PA
C8	+	+	+	+	+	+	+	+	+	PA
C11	+	+	+	+	+	+	+	+	+	PA
C15	+	+	+	+	+	+	+	+	+	PA
C17	+	+	+	+	+	+	+	+	+	PA
C21	+	+	+	+	+	+	+	+	+	PA
C24	+	+	+	+	+	+	+	+	+	PA
C1	+	+	+	+	+	+	+	+	+	PA
C4	+	+	+	+	+	+	+	+	+	PA
C6	+	+	+	+	+	+	+	+	+	PA
C10	+	+	+	+	+	+	+	+	+	PA
C12	+	+	+	+	+	+	+	+	+	PA
C16	+	+	+	+	+	+	+	+	+	PA
C19	+	+	+	+	+	+	+	+	+	PA
C20	+	+	+	+	+	+	+	+	+	PA

Collaborator D

Aerobic mesophilic flora: 5,6 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
D3	-	-	-	-	-	-	/	/	-	NA
D5	-	-	-	-	-	-	/	/	-	NA
D9	-	-	-	-	-	-	/	/	-	NA
D13	-	-	-	-	-	-	/	/	-	NA
D14	-	-	-	-	-	-	/	/	-	NA
D18	-	-	-	-	-	-	/	/	-	NA
D22	-	-	-	-	-	-	/	/	-	NA
D23	-	-	-	-	-	-	/	/	-	NA
D2	-	-	-	-	-	-	/	/	-	NA
D7	-	-	-	-	-	-	/	/	-	NA
D8	+	+	+	+	+	+	+	+	+	PA
D11	+	+	+	+	+	+	+	+	+	PA
D15	+	+	+	+	+	+	+	+	+	PA
D17	+	+	+	+	+	+	+	+	+	PA
D21	+	+	+	+	+	+	+	+	+	PA
D24	+	+	+	+	+	+	+	+	+	PA
D1	+	+	+	+	+	+	+	+	+	PA
D4	+	+	+	+	+	+	+	+	+	PA
D6	+	+	+	+	+	+	+	+	+	PA
D10	+	+	+	+	+	+	+	+	+	PA
D12	+	+	+	+	+	+	+	+	+	PA
D16	+	+	+	+	+	+	+	+	+	PA
D19	+	+	+	+	+	+	+	+	+	PA
D20	+	+	+	+	+	+	+	+	+	PA

Collaborator E
 Aerobic mesophilic flora: 3,4 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
E3	-	-	-	-	-	-	/	/	-	NA
E5	-	-	-	-	-	-	/	/	-	NA
E9	-	-	-	-	-	-	/	/	-	NA
E13	+	+	+	+	-	-	/	/	-	NA
E14	+	-	-	-	-	-	/	/	-	NA
E18	-	-	-	-	-	-	/	/	-	NA
E22	+	-	+	+	+	-	/	/	-	ND
E23	-	-		+	+	-	/	/	-	ND
E2	+	+	+	+	+	+	+	+	+	PA
E7	+	+	+	+	+	+	+	+	+	PA
E8	+	+	+	+	+	+	+	+	+	PA
E11	+	+	+	+	+	+	+	+	+	PA
E15	+	+	+	+	+	+	+	+	+	PA
E17	+	+	+	+	+	-/+*	+	+	-	ND _{FN(alt)}
E21	+	+	+	+	+	+	+	+	+	PA
E24	+	+	+	+	+	+	+	+	+	PA
E1	+	+	+	+	+	+	+	+	+	PA
E4	+	+	+	+	+	+	+	+	+	PA
E6	+	+	+	+	+	+	+	+	+	PA
E10	+	+	+	+	+	+	+	+	+	PA
E12	+	+	+	+	+	+	+	+	+	PA
E16	+	+	+	+	+	+	+	+	+	PA
E19	+	+	+	+	+	+	+	+	+	PA
E20	+	+	+	+	+	+	+	+	+	PA

Collaborator F

Aerobic mesophilic flora: 4,2 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
F3	-	-	-	-	-	-	/	/	-	NA
F5	-	-	-	-	-	-	/	/	-	NA
F9	-	-	-	-	-	-	/	/	-	NA
F13	-	-	-	-	-	-	/	/	-	NA
F14	-	-	-	-	-	-	/	/	-	NA
F18	-	-	-	-	-	-	/	/	-	NA
F22	-	-	-	-	-	-	/	/	-	NA
F23	-	-	-	-	-	-	/	/	-	NA
F2	+	+	+	+	+	+	+	+	+	PA
F7	+	+	+	+	+	+	+	+	+	PA
F8	+	+	+	+	+	+	+	+	+	PA
F11	+	+	+	+	+	+	+	+	+	PA
F15	+	+	+	+	+	+	+	+	+	PA
F17	+	+	+	+	+	+	+	+	+	PA
F21	+	+	+	+	+	+	+	+	+	PA
F24	+	+	+	+	+	+	+	+	+	PA
F1	+	+	+	+	+	+	+	+	+	PA
F4	+	+	+	+	+	+	+	+	+	PA
F6	+	+	+	+	+	+	+	+	+	PA
F10	+	+	+	+	+	+	+	+	+	PA
F12	+	+	+	+	+	+	+	+	+	PA
F16	+	+	+	+	+	+	+	+	+	PA
F19	+	+	+	+	+	+	+	+	+	PA
F20	+	+	+	+	+	+	+	+	+	PA

Collaborator G

Aerobic mesophilic flora: 3.7 x 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
G3	-	-	-	-	-	-	-	-	-	NA
G5	-	-	-	-	-	-	/	/	-	NA
G9	-	-	-	-	-	-	/	/	-	NA
G13	-	-	-	-	-	-	/	/	-	NA
G14	-	-	-	-	-	-	/	/	-	NA
G18	-	-	-	-	-	-	/	/	-	NA
G22	-	-	-	-	-	-	/	/	-	NA
G23	-	-	-	-	-	-	/	/	-	NA
G2	+	+	+	+	+	+	+	+	+	PA
G7	+	+	+	+	+	+	+	+	+	PA
G8	+	+	+	+	+	+	+	+	+	PA
G11	+	+	+	+	+	+	+	+	+	PA
G15	+	+	+	+	+	+	+	+	+	PA
G17	+	+	+	+	+	+	+	+	+	PA
G21	+	+	+	+	+	+	+	+	+	PA
G24	+	+	+	+	+	+	+	+	+	PA
G1	+	+	+	+	+	+	+	+	+	PA
G4	+	+	+	+	+	+	+	+	+	PA
G6	+	+	+	+	+	+	+	+	+	PA
G10	+	+	+	+	+	+	+	+	+	PA
G12	+	+	+	+	+	+	+	+	+	PA
G16	+	+	+	+	+	+	+	+	+	PA
G19	+	+	+	+	+	+	+	+	+	PA
G20	+	+	+	+	+	+	+	+	+	PA

Collaborator H
 Aerobic mesophilic flora: 3.3 x 10⁶
 cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
H3	-	-	-	-	-	-	/	/	-	NA
H5	-	-	-	-	-	-	/	/	-	NA
H9	-	-	-	-	-	-	/	/	-	NA
H13	-	-	-	-	-	-	/	/	-	NA
H14	-	-	-	-	-	-	/	/	-	NA
H18	-	-	-	-	-	-	/	/	-	NA
H22	-	-	-	-	-	-	/	/	-	NA
H23	-	-	-	-	-	-	/	/	-	NA
H2	+	+	+	+	+	+	+	+	+	PA
H7	+	+	+	+	+	+	+	+	+	PA
H8	+	+	+	+	+	+	+	+	+	PA
H11	+	+	+	+	+	+	+	+	+	PA
H15	+	+	+	+	+	+	+	+	+	PA
H17	+	+	+	+	+	+	+	+	+	PA
H21	-	-	-	-	-	-	/	/	-	NA
H24	+	+	+	+	+	+	+	+	+	PA
H1	+	+	+	+	+	-/+*	+	+	-	ND _{FN(alt)}
H4	+	+	+	+	+	+	+	+	+	PA
H6	+	+	+	+	+	+	+	+	+	PA
H10	+	+	+	+	+	+	+	+	+	PA
H12	+	+	+	+	+	+	+	+	+	PA
H16	+	+	+	+	+	+	+	+	+	PA
H19	+	+	+	+	+	+	+	+	+	PA
H20	+	+	+	+	+	+	+	+	+	PA

Collaborator |

Aerobic mesophilic flora: 2,6 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.spp	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
I3	-	-	-	-	-	-	/	/	-	NA
I5	-	-	-	-	-	-	/	/	-	NA
I9	-	-	-	-	-	-	/	/	-	NA
I13	-	-	-	-	-	-	/	/	-	NA
I14	-	-	-	-	-	-	/	/	-	NA
I18	-	-	-	-	-	-	/	/	-	NA
I22	-	-	-	-	-	-	/	/	-	NA
I23	-	-	-	-	-	-	/	/	-	NA
I2	+	+	+	+	+	-/+*	+	+	-	ND _{FN(alt)}
I7	-	-	+	+	+	-/+*	+	+	-	ND _{FN(alt)}
I8	+	+	+	+	+	+	+	+	+	PA
I11	-	-	+	+	+	-/*	+	+	-	ND _{FN(alt)}
I15	-	-	-	-	-	-	/	/	-	NA
I17	+	+	+	+	+	+	+	+	+	PA
I21	+	+	+	+	+	+	+	+	+	PA
I24	+	+	+	+	+	+	+	+	+	PA
I1	+	+	+	+	+	+	+	+	+	PA
I4	+	+	+	+	+	+	+	+	+	PA
I6	+	+	+	+	+	+	+	+	+	PA
I10	+	+	+	+	+	+	+	+	+	PA
I12	+	+	+	+	+	+	+	+	+	PA
I16	+	+	+	+	+	+	+	+	+	PA
I19	+	+	+	+	+	+	+	+	+	PA
I20	+	+	+	+	+	+	+	+	+	PA

Collaborator J

Aerobic mesophilic flora: 5.9×10^6 cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
J3	-	-	-	-	-	-	-	-	-	NA
J5	-	-	-	-	-	-	-	-	-	NA
J9	-	-	-	-	-	-	-	-	-	NA
J13	-	-	-	-	-	-	-	-	-	NA
J14	-	-	-	-	-	-	-	-	-	NA
J18	-	-	-	-	-	-	-	-	-	NA
J22	-	-	-	-	-	-	-	-	-	NA
J23	-	-	-	-	-	-	-	-	-	NA
J2	+	+	+	+	+	+	+	+	+	PA
J7	+	+	+	+	+	+	+	+	+	PA
J8	+	+	+	+	+	+	+	+	+	PA
J11	+	+	+	+	+	+	+	+	+	PA
J15	+	+	+	+	+	+	+	+	+	PA
J17	+	+	+	+	+	+	+	+	+	PA
J21	+	+	+	+	+	+	+	+	+	PA
J24	+	+	+	+	+	+	+	+	+	PA
J1	+	+	+	+	+	+	+	+	+	PA
J4	+	+	+	+	+	+	+	+	+	PA
J6	+	+	+	+	+	+	+	+	+	PA
J10	+	+	+	+	+	+	+	+	+	PA
J12	+	+	+	+	+	+	+	+	+	PA
J16	+	+	+	+	+	+	+	+	+	PA
J19	+	+	+	+	+	+	+	+	+	PA
J20	+	+	+	+	+	+	+	+	+	PA

Collaborator K

Aerobic mesophilic flora: 6,0 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
K3	-	-	-	-	-	+	-	-	-	PD _{FP(alt)}
K5	-	-	-	-	-	-	-	-	-	NA
K9	-	-	-	-	-	-	-	-	-	NA
K13	-	-	-	-	-	-	-	-	-	NA
K14	-	-	-	-	-	-	-	-	-	NA
K18	-	-	-	-	-	-	-	-	-	NA
K22	-	-	-	-	-	-	-	-	-	NA
K23	-	-	-	-	-	-	-	-	-	NA
K2	+	+	+	+	+	+	+	+	+	PA
K7	+	+	+	+	+	+	+	+	+	PA
K8	+	+	+	+	+	+	+	+	+	PA
K11	+	+	+	+	+	+	+	+	+	PA
K15	+	+	+	+	+	+	+	+	+	PA
K17	+	+	+	+	+	+	+	+	+	PA
K21	+	+	+	+	+	+	+	+	+	PA
K24	+	+	+	+	+	+	+	+	+	PA
K1	+	+	+	+	+	+	+	+	+	PA
K4	+	+	+	+	+	+	+	+	+	PA
K6	+	+	+	+	+	+	+	+	+	PA
K10	+	+	+	+	+	+	+	+	+	PA
K12	+	+	+	+	+	+	+	+	+	PA
K16	+	+	+	+	+	+	+	+	+	PA
K19	+	+	+	+	+	+	+	+	+	PA
K20	+	+	+	+	+	+	+	+	+	PA

Collaborator L
 Aerobic mesophilic flora: 5.6 x 10⁶
 cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
L3	-	-	-	-	-	-	/	/	-	NA
L5	-	-	-	-	-	-	/	/	-	NA
L9	-	-	-	-	-	-	/	/	-	NA
L13	-	-	-	-	-	-	/	/	-	NA
L14	-	-	-	-	-	-	/	/	-	NA
L18	-	-	-	-	-	-	/	/	-	NA
L22	-	-	-	-	-	-	/	/	-	NA
L23	-	-	-	-	-	-	/	/	-	NA
L2	-	-	-	-	-	-	/	/	-	NA
L7	+	+	+	+	+	+	+	+	+	PA
L8	+	+	+	+	+	+	+	+	+	PA
L11	+	+	+	+	+	+	+	+	+	PA
L15	+	+	+	+	+	+	+	+	+	PA
L17	+	+	+	+	+	+	+	+	+	PA
L21	-	-	-	-	-	-	/	/	-	NA
L24	+	+	+	+	+	+	+	+	+	PA
L1	+	+	+	+	+	+	+	+	+	PA
L4	+	+	+	+	+	+	+	+	+	PA
L6	+	+	+	+	+	+	+	+	+	PA
L10	+	+	+	+	+	+	+	+	+	PA
L12	+	+	+	+	+	+	+	+	+	PA
L16	+	+	+	+	+	+	+	+	+	PA
L19	+	+	+	+	+	+	+	+	+	PA
L20	+	+	+	+	+	+	+	+	+	PA

Collaborator M

Aerobic mesophilic flora: 4.0 x 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
M3	-	-	-	-	-	-	/	/	-	NA
M5	-	-	-	-	-	-	/	/	-	NA
M9	-	-	-	-	-	-	/	/	-	NA
M13	-	-	-	-	-	-	/	/	-	NA
M14	-	-	-	-	-	-	/	/	-	NA
M18	-	-	-	-	-	-	/	/	-	NA
M22	-	-	-	-	-	-	/	/	-	NA
M23	-	-	-	-	-	-	/	/	-	NA
M2	-	-	-	-	-	-	/	/	-	NA
M7	-	-	-	-	-	-	/	/	-	NA
M8	+	+	+	+	+	+	+	+	+	PA
M11	-	-	-	-	-	-	/	/	-	NA
M15	-	-	+	+	+	-	+	+	-	ND _{FN(alt)}
M17	+	+	+	+	+	+	+	+	+	PA
M21	-	-	-	-	-	-	/	-	-	NA
M24	+	+	+	+	+	+	+	+	+	PA
M1	+	+	+	+	+	+	+	+	+	PA
M4	+	+	+	+	+	+	+	+	+	PA
M6	+	+	+	+	+	+	+	+	+	PA
M10	-	-	+	+	+	+	+	+	+	PA
M12	+	+	+	+	+	+	+	+	+	PA
M16	+	+	+	+	+	+	+	+	+	PA
M19	+	+	+	+	+	+	+	+	+	PA
M20	+	+	+	+	+	+	+	+	+	PA

Collaborator N
 Aerobic mesophilic flora: 3.6 x 10⁶
 cfu/g

Sample N°	Reference method ISO 11290-1					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
N3	-	-	-	-	-	-	/	/	-	NA
N5	-	-	-	-	-	-	/	/	-	NA
N9	-	-	-	-	-	-	/	/	-	NA
N13	-	-	-	-	-	-	/	/	-	NA
N14	-	-	-	-	-	-	/	/	-	NA
N18	-	-	-	-	-	-	/	/	-	NA
N22	-	-	-	-	-	-	/	/	-	NA
N23	-	-	-	-	-	-	/	/	-	NA
N2	+	+	+	+	+	+	+	+	+	PA
N7	+	+	+	+	+	+	+	+	+	PA
N8	-	-	-	-	-	-	+	+	-	NA _{FN(alt)}
N11	+	+	+	+	+	+	+	+	+	PA
N15	+	+	+	+	+	+	+	+	+	PA
N17	+	d	+	+	+	+	+	+	+	PA
N21	+	+	+	+	+	+	+	+	+	PA
N24	+	+	+	+	+	+	+	+	+	PA
N1	+	+	+	+	+	+	+	+	+	PA
N4	+	+	+	+	+	+	+	+	+	PA
N6	+	+	+	+	+	+	+	+	+	PA
N10	+	+	+	+	+	+	+	+	+	PA
N12	+	+	+	+	+	+	+	+	+	PA
N16	+	+	+	+	+	+	+	+	+	PA
N19	+	+	+	+	+	+	+	+	+	PA
N20	+	+	+	+	+	+	+	+	+	PA

d: doubtful colonies

Collaborator O (ADRIA)
 Aerobic mesophilic flora: 2,9 10⁶ cfu/g

Sample N°	Reference method ISO 11290-1 [♦]					Alternative method: Assurance GDS Listeria monocytogenes				Agreement
	Fraser 1/2		Fraser 1		Final result	PCR GDS L.monocytogenes	OCLA	Palcam	Final result	
	OAA	Palcam	OAA	Palcam						
O3	-	-	-	-	-	-	-	-	-	NA
O5	-	-	-	-	-	-	-	-	-	NA
O9	-	-	-	-	-	-	-	-	-	NA
O13	-	-	-	-	-	-	-	-	-	NA
O14	-	-	-	-	-	-	-	-	-	NA
O18	-	-	-	-	-	-	-	-	-	NA
O22	-	-	-	-	-	-	-	-	-	NA
O23	-	-	-	-	-	-	-	-	-	NA
O2	+	+	+	+	+	+	+	+	+	PA
O7	+	+	+	+	+	+	+	+	+	PA
O8	+	+	+	+	+	+	+	+	+	PA
O11	+	+	+	+	+	+	+	+	+	PA
O15	+	+	+	+	+	+	+	+	+	PA
O17	+	+	+	+	+	+	+	+	+	PA
O21	-	-	-	-	-	-	-	-	-	NA
O24	+	+	+	+	+	+	+	+	+	PA
O1	+	+	+	+	+	+	+	+	+	PA
O4	+	+	+	+	+	+	+	+	+	PA
O6	+	+	+	+	+	+	+	+	+	PA
O10	+	+	+	+	+	+	+	+	+	PA
O12	+	+	+	+	+	+	+	+	+	PA
O16	+	+	+	+	+	+	+	+	+	PA
O19	+	+	+	+	+	+	+	+	+	PA
O20	+	+	+	+	+	+	+	+	+	PA

[♦] Analyses performed according to the COFRAC accreditation (Accreditation Testing n°1-0144, scope available on www.cofrac.fr)