

MicroVal Study 2017LR74: Confirmation Method

Validation study of the MALDI Biotyper® Complete Solution as an Alternative Method for the Confirmation of *Campylobacter* spp.

MCS ILS, Version 2
March 5, 2020

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The report is prepared in accordance with ISO 16140-6:2019 and the most recent version of the MicroVal Technical Committee for interpretation on ISO 16140-6, version 2.0.

Company: Bruker Daltonics

Expert Laboratory: ADRIA Development (FR) and Q Laboroatories (OH,USA)

Method/Kit name: MALDI Biotyper for *Campylobacter* spp. confirmation

Validation standard: ISO 16140-6: 2019 Microbiology of food chain – Method Validation–Part 6: Protocol for the validation of alternative (proprietary) methods for microbiological confirmation and typing procedures.

Reference methods:

ISO 10272:2017 Microbiology of the food chain - Horizontal method for detection and enumeration of *Campylobacter* spp.

Scope of validation: Confirmation of isolates from mCCDA, Campy Cefex Agar, Chromogenics agars (CampyFood Agar, RAPID'*Campylobacter*) and a non-selective agar (Columbia blood agar).

Certification organization: Lloyd's Register

List of abbreviations

-	No typical colonies but presence of background microflora
AL	Acceptability Limit
Alt	Alternative method
BG-	Gram negative rod
CBA	Columbia blood agar
d	Doubtful result
DT	Direct transfer procedure
eDT	Extended direct transfer procedure
EXT	Extraction procedure
EL	Expert Laboratory
g	Gram
h	Hour
MALDI-TOF	Matrix Assisted Laser Desorption/Ionization Time-Of-Flight (MALDI-TOF)
MBT	Maldi Biotyper
MS	Mass Spectrometry
mCCDA	Modified charcoal-cefoperazone-deoxycholate agar
MCS	Method Comparison Study
min	minute
ml	millilitre
MR	(MicroVal) Method Reviewer
MSP	Main Spectrum or Main Spectra
MVTC	MicroVal Technical Committee

NA	Negative agreement
ND	Negative deviation
NOIP	No organism identification possible
PA	Positive agreement
PD	Positive deviation
pos (+)	positive/growth/target detected
RT	Relative Trueness
TSA	Tryptic Soy Agar
w	Weak reaction

Contents

Introduction	6
1 Method protocols	6
1.1 Reference method	6
1.2 Alternative method	7
1.2.1 <i>Instrumentation Workflow and Mechanism</i>	7
1.2.2 <i>Calculating a log (score) value and interpretation</i>	8
1.2.3 <i>Instrumentation Features</i>	10
1.2.4 <i>Software Features</i>	11
1.2.5 <i>Targets and biotargets</i>	11
1.2.6 <i>Associated devices</i>	12
1.2.7 <i>Consumables</i>	12
1.3 Study design	13
2 Method comparison study	13
2.1 Selection of Test Strains	13
2.2 Samples to be tested	13
2.3 Inclusivity and exclusivity	14
2.3.1 Data discrepancy	14
2.3.2 <i>Campylobacter</i> spp. testing at the genus level	14
2.3.3. Summary of the inclusivity and exclusivity testing	15
2.3.3 Expression of results	16
2.3.4 Evaluation	22
5 Interlaboratory Study	23
5.1 <i>Protocol and Materials</i>	23
5.2 <i>Transport</i>	25
5.3 <i>Analysis</i>	25
5.4 <i>Statistics and Results</i>	26

6	Conclusions of Interlaboratory Study	31
	Annex A - Flow diagram of the reference method: ISO 10272 (June 2017) - Microbiology of the food chain - Horizontal method for detection and enumeration of <i>Campylobacter</i> spp.	32
	Annex B - Flow diagram of the alternative method	34
	Annex C - Kit inserts	36
	Annex D – Certificate of equivalence	45
	Annex E – <i>Campylobacter</i> spp. testing at the genus level	46
	Annex F - Raw data inclusivity study	47
	Annex G - Raw data exclusivity study	48
	Annex H - Results inclusivity study:	49
	Annex I - Results exclusivity study	72
	Annex J - Detailed list of participants	126
	Annex K - Raw data generated during the Bruker MADLI Biotyper Inter-Laboratory Study for <i>Campylobacter</i>	127

Introduction

The purpose of this MicroVal study was to validate the use of the Bruker MALDI Biotyper® (MBT) using a presumptive *Campylobacter* spp. (defined by ISO 16140-1:2016) as an alternative method to **ISO 16140-6:2019**. Only the rates of confirmed and non-confirmed results were considered in the final interpretation. Additional details about the strains tested with the MALDI Biotyper are provided in the report for information purposes only.

The MBT utilizes Matrix Assisted Laser Desorption/Ionization Time-Of-Flight (MALDI-TOF) Mass Spectrometry (MS) for the rapid and accurate confirmation and identification of micro-organisms isolated from select media types. The MBT method is designed to rapidly confirm a presumptive result and identify the isolates from a variety of agar plates.

The alternative method was evaluated to confirm *Campylobacter* spp. according to the technical rules described in the ISO 16140 - Part 6 - Microbiology of the food chain - Method validation - Part 6: protocol for the validation of alternative (proprietary) methods for microbiological confirmation and typing procedures. The tested microorganisms were isolated on mCCDA (Charcoal Cefoperazone Deoxycholate Agar), Campy Cefex Agar, Chromogenics agars (CampyFood Agar, RAPID'*Campylobacter*) and one non-selective agar (Columbia blood agar, CBA). This evaluation was performed by ADRIA Development (FR) and Q Laboratories (OH,USA) as the MicroVal Expert Laboratories. Two MBT systems were used for the testing; the MBT with the microflex LT/SH instrument and the MBT smart with the microflex LT/SH smart instrument. Half of the Method Comparison Study (MCS) was carried out at ADRIA and half at Q Laboratories. For the ILS, Q Laboratories organized and included 15 collaborators from different laboratories. This report summarizes the results and interpretations of the MCS and the ILS. A lack of recovery of the strains was observed, and it was decided to organize a second interlaboratory study. This second trial was conducted by Q Laboratories.

Reference method: ISO 10272:2017

Scope of the validation study was the confirmation of *Campylobacter* spp. presumptive colonies isolated on mCCDA, Campy Cefex Agar, Chromogenics agars (CampyFood Agar, RAPID'*Campylobacter*) and a non-selective agar (Columbia blood agar).

1 Method protocols

1.1 Reference method

See the flow diagram in **Annex A**.

The reference methods correspond to the ISO 10272 (June 2017) - Microbiology of the food chain - Horizontal method for detection and enumeration of *Campylobacter* spp. - Part 1: detection method and the ISO 10272 (June 2017) - Microbiology of the

food chain - Horizontal method for detection and enumeration of *Campylobacter* spp.
- Part 2: Colony-count technique.

1.2 Alternative method

See the flow diagram of the alternative method in **Annex B**.

1.2.1 Instrumentation Workflow and Mechanism

An individual colony from a culture plate of the tested strain was transferred to a selected position on a MALDI target plate. The MALDI target plate was air-dried and the matrix was added. The matrix solution was used to extract mostly ribosomal proteins at high concentrations from the microorganisms. When the matrix crystallized and was completely dry, the MALDI target plate was analyzed using the **MBT System microflex LT/SH** or the **MBT System microflex LT/SH smart** with the corresponding MBT software (4.0 and Compass versions).

Strains were analyzed using MALDI-TOF MS. A laser in the MALDI-TOF MS irradiates the dried matrix spot. The rapid evaporation of the matrix and proteins caused by the laser resulted in the release of intact, positively charged proteins and peptides (a "soft" ionization technique). These ions are electrostatically accelerated over a short distance and arrive in the flight tube at a mass-dependent speed. As different proteins/peptides have different masses, ions arrive at the detector at different times (Time Of Flight). The mass spectrometer measures the time (in the microsecond range) between pulsed acceleration and the corresponding detector signal. The speed is converted into a molecular mass. The highly abundant microbial ribosomal proteins result in a mass spectrum with a characteristic mass and intensity distribution pattern. For many microorganisms, this pattern is species-specific and can be used as a "molecular fingerprint" to identify the sample.

The **MBT Explorer Module** allows sophisticated statistical analysis as well as the extension of existing libraries or the creation of new libraries; statistical analysis is supported by PCA (Principal Component Analysis), Clustering (dendrogram creation and examination) and correlation analysis based on a composite correlation index (CCI) matrix approach.

The MALDI Biotyper (MBT) software and library:

- The MBT System microflex LT/SH (MBT) or MBT System microflex LT/SH smart (MBT-smart) instruments are benchtop MALDI-Time-Of-Flight Mass Spectrometers for accurate mass determination of biomolecules.
- The **MBT Compass** or **MBT 4.0 Software** are dedicated software packages for microorganism identification based on MALDI-TOF MS profile spectra. The MBT Compass software controls mass spectrometry data acquisition and matches acquired spectra against the MBT Compass library for final identification results. The software generates a report using score values and scoring symbols.

- The **MBT Compass Library** includes defined bacteria and yeast strains from industrial, veterinary, clinical isolates provided by collaborating partners, round robin strains and strains from accredited ISO 9001:2008 certified strain collections.
- This validation was run with the MBT Compass Library version 6903 and was, therefore, included as part of the validation scope.

The eDT and EXT sample preparations were used when recommended, based on the calculating log (score) value. The sample stability onto prepared targets was guaranteed up to 24 hours at 20 - 25°C.

1.2.2 Calculating a log (score) value and interpretation

Data acquisition was controlled and analyzed using MBT Compass software. The spectrum of the unknown sample was first translated into a peak list. Using a biostatistical algorithm, this peak list was compared to reference peak lists of organisms in the reference library database and a log (score) between 0.00 and 3.00 was generated. The higher the log (score), the higher the degree of similarity between the pattern for the unknown peak list and the peak list for the database entry in the reference library. Scores higher or equal to 2.00 presented in green were determined to be a high confidence identification. Results presented in yellow have values between 1.70 and 1.99 and were considered low confidence identification. Results presented in red were considered not acceptable for identification. The consistency category of the identification was based on the confidence level of the best and second-best matches as described below.

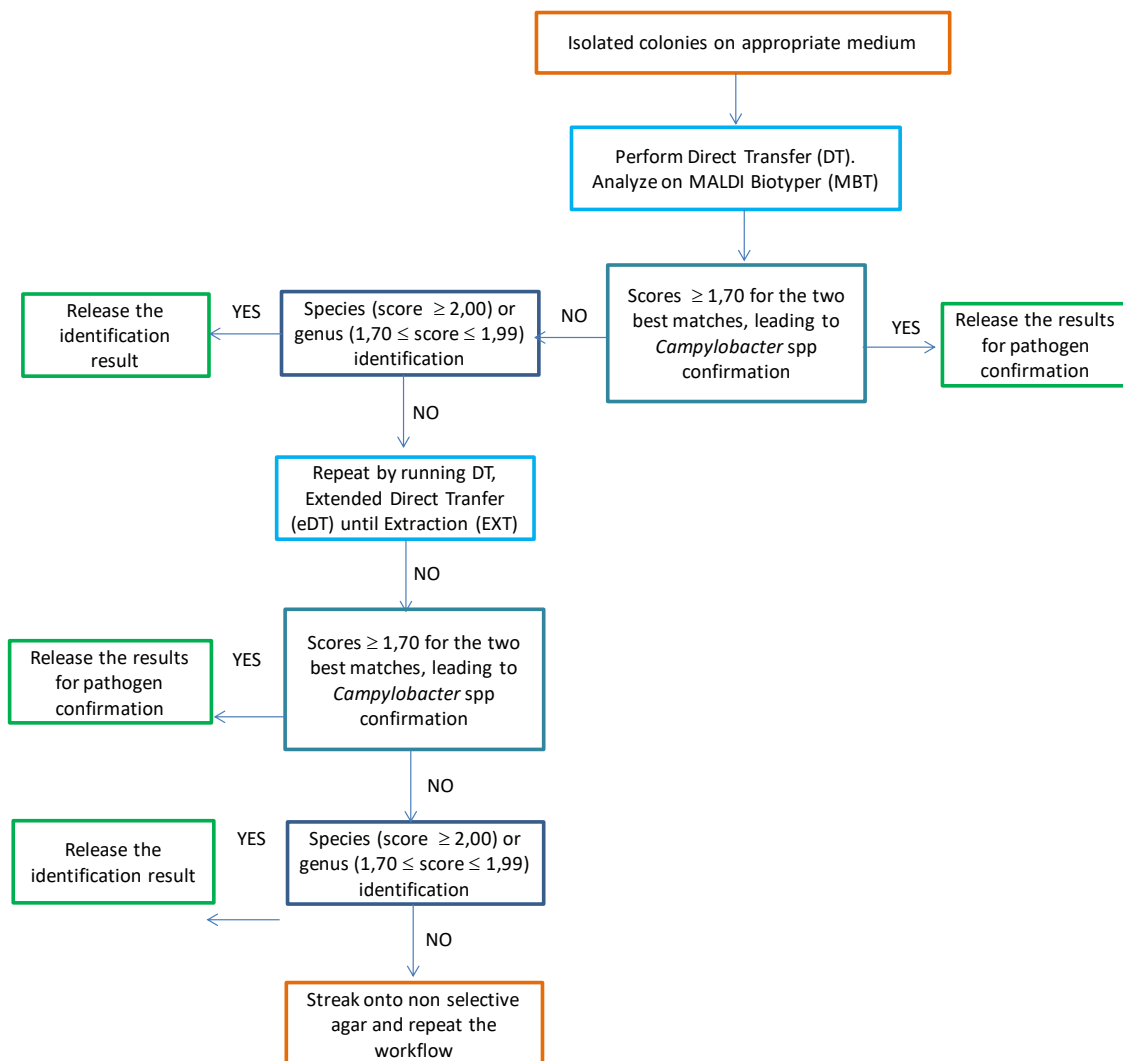
For the confirmation of foodborne pathogens at the genus level, the two highest matches were considered for identification if they both have a log (scores) higher than 1.70 and propose the same identified genus. If the log (score) is < 1.70 after initial analysis, the DT sample preparation was repeated, and if needed the alternative sample preparation procedures, eDT and EXT, was run (See **Annex C**). **Table 1** shows a summary for identification results. If the identification is still not possible, it was advised to streak on a non-selective agar.

Table 1. Shows identification category and description

Identification consistency category	Description
High	The best match is a high-confidence identification. The second-best match was: - a high-confidence identification in which the species is identical to the best match; - a low-confidence identification in which the genus is identical to the best match; - a non-identification.
Low	The requirements for high consistency were not met. The best match was a high or low-confidence identification. The second-best match was: - a high or low-confidence identification in which the genus is identical to the best match; - a non-identification.
None	The requirements for high or low consistency were not met.

The MBT enabled the confirmation of isolated colonies from non-selective and selective culture media. The confirmation was done at the genus level for *Campylobacter* spp. Based on the observed log (score) values, the eDT and/or EXT sample preparations were run as described in the standard operation procedures (SOP) of the MBT Compass Manual. **Figure 1** shows the workflow.

Figure 1



The MBT report for Food Microbiology, activated in the MBT Compass software, provided the confirmation and identification results according to this flowchart.

The Quick Guide for Food Testing was provided in addition to the MBT Compass user manual, in order to describe the scope of the ISO 16140-6 validation, the workflow, the controls, and the interpretation.

1.2.3 Instrumentation Features

As previously stated, two MBT Complete Solutions are currently available depending on the instrument, MBT System microflex LT/SH (MBT) or MBT System microflex LT/SH smart (MBT-smart). The two instruments were tested during the validation: the MBT microflex LT/SH by the expert laboratory. The blind-coded biotargets were sent for a second reading by BRUKER (Europe) and Atlanta Hospital (US) using the **MBT-smart**. Bruker internal data were available to support the sample stability after the matrix addition up to 24 hours at 20-25°C, but in some cases, the biotargets

arrived one day later. The differences result from the laser technology and the vacuum system are presented below (See **Table 2**).

Table 2. A Comparison of the laser technology and the vacuum system

MALDI Biotyper System microflex LT/SH	MALDI Biotyper Smart System microflex LT/SH
<i>Vacuum system</i>	<i>High-performance vacuum system</i>
<i>Gridless microSCOUT™ MALDI ion source with extended pulsed ion extraction PAN™ mass range focusing</i>	<i>Gridless microSCOUT™ MALDI ion source with extended pulsed ion extraction PAN™ mass range focusing</i>
<i>Integrated source cleaning without the need of venting the system</i>	<i>Integrated source cleaning without the need of venting the system</i>
<i>60 Hz nitrogen laser</i>	<i>200 Hz BRUKER smartbeam™ solid-state lifetime laser</i>
<i>TOF analyzer for linear-only measurements with positive acceleration potential</i>	<i>TOF analyzer for linear-only measurements with positive acceleration potential</i>
<i>Data System with digitizer, WIN7 Operating-System, >=24" LCD-Display, Printer and Compass™ for Flex software package for MS control, data acquisition and processing</i>	<i>Data System with digitizer, WIN7 Operating-System, >=24" LCD-Display, Printer and Compass™ for Flex software package for MS control, data acquisition and processing</i>

1.2.4 Software Features

Two versions of the software are currently available: MBT Compass and MBT 4.0. They use the same algorithm and thus provide the same identification result, as presented in the certificate of equivalence (See **Annex D**).

The only differences in the two software versions are the following:

- The MBT Compass enables the use of the Subtyping Module for automated detection of strain specific characteristics, while the MBT 4.0 cannot;
- The MBT 4.0 enables electronic signature and audit trail with the MBT Security Pack, while the MBT Compass still must be implemented for that option.

1.2.5 Targets and biotargets

The samples can be plotted on several targets:

- Reusable 48 and 96 polished steel target;
- Disposable Biotarget 96, which can be used until all the sample positions are filled.

These targets were simply materials to plot the samples and data acquired to comply with the ISO 13485 and ISO 9001 certification were available to show the equivalence. The method comparison study was run with disposable biotargets to simplify the workflow and, therefore, no washing step was required.

1.2.6 Associated devices

The MBT was associated to the following devices in order to facilitate the workflow:

- MBT Pilot workstation supported correct sample positioning by guided MALDI target preparation through micro-projection technology; MBT Galaxy automated target preparation system frees laboratory personal from routine work, using exactly the same sample procedure as the manual one; indeed, no additional testing was required, and data acquired to comply with the ISO 13485 and ISO 9001 certification were available to show the equivalence. Note, that the MBT Galaxy is not a colony picker, the MBT Galaxy runs only automatic pipetting of the HCCA (α -cyano-4-hydroxycinnamic acid) matrix of formic acid. Since automation steps were identical to those used in the manual process, it was only necessary to use the manual method within the validation.
- Barcode Scanner can be used for easy reading of barcodes of routine samples.

For additional information, please see: <https://www.bruker.com/products/mass-spectrometry-and-separations/ivd-ce-certified-maldi-biotyper/consumables-accessories-for-mbt-workflows.html>

1.2.7 Consumables

The following consumables were required for the manual procedure:

- HCCA Matrix for the sample preparation;
- Bacterial Test Standard (BTS), i.e. mass spectrometry calibration standard showing a typical *Escherichia coli* DH5 alpha peptide and protein profile plus additional proteins in MALDI-TOF MS. **It is mandatory to get at least one valid BTS control per run.**

Additionally, formic acid was used for the extended direct transfer, as well as ethanol and acetonitrile for the extraction protocol.

1.3 Study design

Microorganisms identified in this MCS using the MBT were isolated by streaking a microbial culture onto an agar plate.

The MBT workflow gathers three sample preparation procedures, which are presented in **Annex C**:

1. Direct transfer sample preparation procedure (DT), which is the usual procedure for most of the bacterial isolates and is used as the general procedure;
2. Extended direct transfer sample preparation procedure (eDT);
3. Extraction sample preparation procedure (EXT).

2 Method comparison study

2.1 Selection of Test Strains

A range of strains were used. The selection of the tested strains was done according to the Annex A of the ISO 16140 – part 6. The strains were selected by considering the measurement principle of the alternative method. Different measurement principles may require the use of different test panels of strains. In addition, the occurrence of suspect and interfering colonies on the media used for the reference method was of importance for selecting strains.

Each strain was characterized biochemically, serologically and/or genetically for it to be correctly identified. Strains were isolated mostly from foods, feed, the food-processing environment, or from primary production in consideration of the scope of the validation. Clinical, environmental and culture collection strains could also be used. The original source of the isolates was traceable and held in a local (e.g. expert laboratory), national or international culture collection enabling them to be used in future testing if required.

The selected strains from the inclusivity and exclusivity lists were sent from collections of the expert laboratories or from culture type collections. They were all identified at the species level using appropriate methods, conventional or molecular identification. This enabled getting a relevant selection of the strains to test a broad range of species for the exclusivity panel and in the inclusivity panel.

2.2 Samples to be tested

Highly characterized pure strains were used for confirmation procedures following the requirements as specified in ISO 7218 and ISO 10272.

The validation study was carried out using strains cultured from stock on a non-selective medium or stored on microbeads at -80°C.

Required previously generated data were already available for the reference confirmation procedure, therefore, it was not necessary to repeat the reference confirmation procedure with the alternative confirmation method for the MCS.

The confirmatory tests described in the ISO 10272 method were applied to the strains for the ILS.

2.3 Inclusivity and exclusivity

The study design is based on the taxonomy level addressed by the alternative method and covered by the scope of the certification. All the strains were blind coded prior to analysis.

Inclusivity

Pure cultures of all strains were required to be tested with the reference confirmation procedure and with the alternative confirmation method. 150 different target strains were tested to the genus level.

Exclusivity

Pure cultures of all strains were required to be tested with the reference confirmation procedure and with the alternative confirmation method. 101 non-target strains were tested to the genus level.

Some of the strains from the exclusivity lists were not able to grow or to show characteristic colonies on the tested selective agars. The strains used in exclusivity testing that exhibited any growth were tested with the MBT procedure to ensure there was no cross-reactivity with the chosen selective agar. The characteristics of the colonies are shown in the raw data.

2.3.1 Data discrepancy

In case of discrepancy (ies) between the alternative and reference methods, the strains were further identified by sequencing the 16S rDNA target. This was the case for some exclusivity strains, but there was no impact on the outcomes of the *Campylobacter* confirmations.

2.3.2 Campylobacter spp. testing at the genus level

150 *Campylobacter* spp. strains and 101 relevant non-*Campylobacter* spp. strains were tested. The list is provided in **Table 3**.

Table 3 - Number of tested strains per species

<i>Campylobacter</i> species tested	Number of strains
<i>Campylobacter coli</i>	64
<i>Campylobacter jejuni</i>	55
<i>Campylobacter lari</i>	25
<i>Campylobacter hyointestinalis</i>	2
<i>Campylobacter upsaliensis</i>	4
Total	150

Each strain was first recovered on a non-selective agar (Columbia blood agar), incubated in microaerobic atmosphere for 24 h at 41.5°C, as well as on four selective agars:

- mCCDA (40 h) and Campy Cefex Agar (24 h) at 41.5°C in microaerobic atmosphere,
- 2 chromogenics, i.e. CampyFood agar, RAPID'*Campylobacter* incubated in microaerobic atmosphere for 24 h at 41.5°C.

Non-target organisms were incubated under optimal growth conditions for the organism. All plates with growth were analyzed by the Bruker Maldi Biotyper System on disposable targets, following the Direct Transfer sample preparation. An isolated colony from each plate was smeared as a thin film directly onto a single sample position on the disposable target. A 1 µL aliquot of BTS was spotted onto a designated position in the disposable target and allowed to dry. After the spotting of the target plate, the HCCA matrix was added within 30 minutes. After the addition of the matrix to the smear, the target plate was analyzed within 24 hours. The protocol is described in **Annex E**.

2.3.3. Summary of the inclusivity and exclusivity testing

Table 4 outlines the tests which were completed. Only strains showing colonies were tested.

Table 4. Tests done during the MCS

Target	Inclusivity		Exclusivity	Total
	Media tested	Tests number		
<i>Campylobacter</i> spp	CBA	150	101	/
	mCCDA	148	12	
	CampyFood Agar	146	11	
	RAPID' <i>Campylobacter</i>	146	4	
	Campy Cefex Agar	149	9	
TOTAL		739	137	876

2.3.3 Expression of results

The raw results are shown in **Annex F** for the inclusivity and **Annex G** for the exclusivity study. The results were tabulated in **Annex H** for the inclusivity and **Annex I** for the exclusivity study.

Table 5 shows the number of strains that showed characteristic colonies, atypical colonies and no growth on the plates, as well as the number of sub-cultures on a non selective agar required to get an identification results.

Table 5. Description of the observed culture data

Study	Media	Number of strains giving characteristic colonies on the plates	Number of strains giving non characteristic colonies on the plates	Number of strains giving no growth on the plates	Number of strains requiring a subculture to get an identification result	
					MBT	MBT smart
Inclusivity	CBA	150	0	0	0	0
	mCCDA	148	0	2	14 (N° 12-14-25-27-28-30-34-36-37-39-43-52-60-72)=NOIP or No peak	18 (N° 4-12-14-15-16-17-25-30-36-37-38-40-43-51-60-62-72-73)=NOIP or No peak
	CampyFood Agar	146	0	4	0	0
	RAPID' <i>Campylobacter</i>	146	0	4	1 (N°72)	0
	Campy Cefex Agar	149	0	1	0	0
Exclusivity	CBA	86	0	15	0	1 (N°40)=NOIP
	mCCDA	9	3	89	0	0
	CampyFood Agar	11	0	90	0	0
	RAPID' <i>Campylobacter</i>	4	0	97	0	0
	Campy Cefex Agar	6	3	92	0	0

NOIP: No organism identification possible

For the inclusivity panel, some strains did not grow on selective agar plates; 2 strains on mCCDA: *Campylobacter upsaliensis* ATCC 49815, *Campylobacter upsaliensis* CIP 103681 and 4 strains on CampyFood Agar and RAPID' *Campylobacter*: *Campylobacter upsaliensis* Ad1139, *Campylobacter upsaliensis* ATCC 49815, *Campylobacter upsaliensis* ATCC 49816 and *Campylobacter upsaliensis* Clp 103681; and 1 strain on Campy Cefex : *Campylobacter upsaliensis* CIP 103681.

For one inclusivity data set (out of two data sets provided by two expert laboratories), subcultures on a non-selective agar were required for 14 strains from mCCDA when analyzed with the MBT and for 18 strains when analyzed with the MBT-smart. Both expert laboratories were using the same mCCDA provider, excluding a formulation impact. However, the Bruker Quick Guide clearly highlights that "mCCDA contains

charcoal.” This component may result in a lower quality of spectra. Attention shall be paid to not transfer a piece of mCCDA agar when smearing the colonies on the target”. Therefore, a sub-culture on a non-selective agar was required to get a confirmation result as described in the flowchart.

The strains were correctly identified at the species level with the MBT, except three wild strains. The strains Ad 1902 and Ad 1910 were sometimes identified to *C. coli* or *C. jejuni* with the MBT. The 16S rDNA results provided the same identity for both *C. coli* and *C. jejuni* species. The original identification at the species level was done by specific PCR according to Denis et al (1999)¹. The strain *C. subantarcticus* Ad 1891 was misidentified to *C. lari*, as there is no *C. subantarcticus* in the MBT library. This corresponds to what some authors consider *C. subantarcticus* as belonging to *C. lari* group based on Whole Genome Sequencing and molecular phylogeny. The *C. lari* group is thus composed of five species *C. lari*, *Campylobacter insulaenigrae*, *Campylobacter volucris*, *Campylobacter subantarcticus*, and *Campylobacter peloridis*, as well as a group of strains termed the urease-positive thermophilic *Campylobacter* (UPTC) and other *C. lari*-like strains^{2,3}

For the exclusivity panel, 88 strains were not able to recover on mCCDA, 89 on CampyFood Agar, 96 on RAPID[®] *Campylobacter*, and 91 on Campy Cefex Agar.

All the strains of the exclusivity panel were recovered on CBA under microaerobe or optimal conditions. They were reflexed to the MBT for testing and none were confirmed as *Campylobacter* spp. The strain *Psychrobacter psychrophilus* Ad1343 was not identified using the MBT-smart from any of the tested medium. The log(score) of the first match was very low, i.e. 1.72 with no second match, with the MBT, while the measurement was done just after the sample preparation. Only two mass spectra of *Psychrobacter* strains are present in the library, i.e. one strain identified at the genus level only and one *Psychrobacter lutiphocae*. The measured mass spectra was different from the reference mass spectra in the library.

All the *Campylobacter* strains were confirmed at the genus level. No strain of the exclusivity panel was identified to *Campylobacter* spp.

For the inclusivity and exclusivity studies, the eDT (extended direct transfer) and EXT (extraction) procedures were used in some cases. This is summarized in **Table 6**.

¹ Denis et al, 1999. Development of a m-PCR assay for simultaneous identification of *Campylobacter jejuni* and *C. coli*. Applied in Letters Microbiology, 29, 406 - 410

² Miller et al, 2014. Comparative genomics of the *Campylobacter lari* group. Genome Biol Evol, 6, 3252-3266

³ Petrovska et al, 2017. Genome Reduction for Niche Association in *Campylobacter Hepaticus*, A Cause of Spotty Liver Disease in Poultry. Front. Cell. Infect. Microbiol., <https://doi.org/10.3389/fcimb.2017.00354> <https://doi.org/10.3389/fcimb.2017.00354>

Table 6. Use of the eDT and EXT Procedures in the MBT workflow

Target	Media	No growth	MBT			MBT smart		
			DT ⁴	eDT ⁵	EXT ⁶	DT	eDT	EXT
Inclusivity	CBA	0	150	0	0	149	0	1
	mCCDA	2	118	0	30	115	5	28
	CampyFood Agar	4	145	1	0	146	0	0
	RAPID' <i>Campylobacter</i>	4	144	1	1	144	2	0
	Campy Cefex Agar	1	148	0	1	146	3	0
	Total	11	705	2	32	700	10	29
	Percentage	/	95,4%	0,3%	4,3%	94,7%	1,4%	3,9%
Exclusivity	CBA	0	100	1	0	99	1	1
	mCCDA	89	12	0	0	12	0	0
	CampyFood Agar	90	11	0	0	10	1	0
	RAPID' <i>Campylobacter</i>	97	4	0	0	4	0	0
	Campy Cefex Agar	92	9	0	0	9	0	0
	Total	368	136	1	0	134	2	1
	Percentage	/	99,3%	0,7%	0,0%	97,8%	1,5%	0,7%

Inclusivity study

For the MBT, the eDT was run for 0.3% of the identification results and the EXT for 4.3%. For the MBT-smart, the eDT was run for 1.4% of the identification results and the EXT for 3.9 %. In that case, the biotargets were analyzed 24 hours (minimum) after the sample preparations.

Exclusivity study

For the MBT, the eDT was run for 0.7% of the identification results and the EXT for 0%. For the MBT-smart, the eDT was run for 1.5% of the identification results and the EXT for 0.7%.

For the MBT-smart, the biotargets were analyzed 24 hours (minimum) after the sample preparations while the sample stability can be guaranteed up to 24 hours at 20 - 25°C.

In the exclusivity panel, some discrepant were observed between the original identification of the strains obtained with the conventional identification procedures and the MBT. These strains were identified using 16S rDNA sequencing results. **Table 7** gathers all the discrepant observed. The original identification of the strains was confirmed with 16S rDNA sequencing results for 2 strains (*Enterococcus durans* Ad148, *Escherichia fergusonii* ATCC 35470). The original identification of the strain was confirmed with the MALDI biotyper at the genus level for 2 strains (*Enterobacter*

⁴ DT: Direct Transfer procedure

⁵ eDT: extended Direct Transfer

⁶ EXT: extraction procedure

intermedius 88a, *Escherichia vulneris* ATCC 29943). All the methods gave a different result making it a challenge to give a final conclusion (*Shewanella putrefaciens* EN/1534, *Enterobacter cancerogenus* QL11010-1).

This table gives additional information, not within the scope of the validation. The interpretation of the results is summarised in **Table 8**; the first step of the confirmation procedure is the isolation on the tested culture media.

Table 7. Disagreements observed in the exclusivity panel

	Agreement between two methods	Disagreement with the two other methods	Not possible to conclude	
	Original identification of the strains	Identification with the MALDI Biotyper and MALDI Biotyper smart	Identification with 16S rDNA sequencing	Conclusion
-	<i>Enterobacter intermedius</i> 88a	<i>Enterobacter cloacae</i> complex or <i>Lelliottia amnigena</i>	<i>Raoultella planticola</i> / <i>ornithinolytica</i>	The identification to <i>Enterobacter</i> genus is confirmed with the MALDI Biotyper.
-	<i>Enterococcus durans</i> Ad148	<i>Enterococcus faecium</i>	<i>Enterococcus durans</i>	The genus is the same with all the methods; conventional procedure and 16S rDNA sequencing provide both identifications to <i>Enterococcus durans</i> .
-	<i>Shewanella putrefasciens</i> EN15/34	<i>Citrobacter</i> spp	<i>Shigella flexneri</i> / <i>sonnei</i> ; <i>Escherichia fergusonii</i>	Difficult to conclude. This is probably an atypical strain, and the API20E, MBT or 16S rDNA libraries can all have a lack of representatives.
-	<i>Enterobacter cancerogenus</i> QL11010-1	<i>Escherichia coli</i>	<i>Shigella</i> , <i>Escherichia</i> , <i>Enterobacter</i>	Difficult to conclude. This is probably an atypical strain, and the API20E, MBT or 16S rDNA libraries can all have a lack of representatives. Moreover, it is recognized that the sequencing of housekeeping genes such as <i>rpoB</i> is often required in the identification of <i>Enterobacteria</i> .
-	<i>Escherichia fergusonii</i> ATCC35470	<i>Escherichia coli</i> or <i>Escherichia fergusonii</i>	<i>Escherichia fergusonii</i>	The genus is the same with all the methods; conventional procedure and 16S rDNA sequencing provide both identifications to <i>Escherichia fergusonii</i> .
-	<i>Escherichia vulneris</i> ATCC29943	<i>Escherichia hermanii</i>	<i>Enterobacter</i> spp	It is probably an <i>Escherichia</i> spp. strain but it is difficult to conclude about the species.

*The MALDI Biotyper report mentions that the species of these genera have very similar mass spectrum patterns. Therefore, distinguishing the species is very difficult

Table 8: Summary of the results in the method comparison study

MALDI System	Media tested	Study	N	IA	ID	EA*	ED
LT/SH-MALDI-MS System	CBA	Inclusivity	150	150	0	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	101	0
	mCCDA	Inclusivity	150 ^a	134	0	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	12	0
	CampyFood Agar	Inclusivity	150	146	0	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	11	0
	RAPID' <i>Campylobacter</i>	Inclusivity	150 ^b	145	0	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	4	0
	Campy Cefex Agar	Inclusivity	150	149	0	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	9	0
LT/SH MALDI Smart System	CBA	Inclusivity	150	150	0	Not applicable	Not applicable
		Exclusivity	101 ^b	Not applicable	Not applicable	100	0
	mCCDA	Inclusivity	150 ^c	130	0	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	12	0
	CampyFood Agar	Inclusivity	150	146	0	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	11	0
	RAPID' <i>Campylobacter</i>	Inclusivity	150	146	101	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	4	0
	Campy Cefex Agar	Inclusivity	150	149	0	Not applicable	Not applicable
		Exclusivity	101	Not applicable	Not applicable	9	0

^a No organism identification possible (NOIP) for 14 out of 103

^b No organism identification possible (NOIP) for 1 out of 103

^c No organism identification possible (NOIP) for 18 out of 103

*Studies with EA< N are due to no growth and the indicated NOIP

For the exclusivity panel, many strains did not grow on the selective plates. In these cases, the final confirmation results were considered as negative as the confirmation procedure starts from the streaking on selective agar plates.

For the inclusivity panel, some strains did not grow on selective agar plates. Those strains included:

- *Campylobacter upsaliensis* Ad1139;
- *Campylobacter upsaliensis* ATCC 49815 (CampyFood Agar; RAPID' *Campylobacter*);
- *Campylobacter upsaliensis* ATCC 49816 (mCCDA, CampyFood Agar; RAPID' *Campylobacter*);
- *Campylobacter upsaliensis* CIP 103681 (mCCDA, CampyFood Agar; RAPID' *Campylobacter*).

Some non-target strains did not grow on all the plates. Only the isolates that grew were considered in the final confirmation results displayed in Table 8-EA. No discrepant results were observed on any of the tested culture medium. Only agreements were observed on all the tested culture medium.

2.3.4 Evaluation

The results of the method comparison study are summarized in the Table 9.

Table 9. Evaluation of the method comparison study results.

MALDI System	Media tested	Study	AL	ID	ID ≤ AL	ED	ED ≤ AL	Evaluation
LT/SH-MALDI-MS System	TSA	Inclusivity	1	0	$0 \leq 1$	Not applicable	Not applicable	Accepted
		Exclusivity	2	Not applicable	Not applicable	0	$0 \leq 2$	Accepted
	CCI	Inclusivity	1	0	$0 \leq 1$	Not applicable	Not applicable	Accepted
		Exclusivity	2	Not applicable	Not applicable	0	$0 \leq 2$	Accepted
	ESIA	Inclusivity	1	0	$0 \leq 1$	Not applicable	Not applicable	Accepted
		Exclusivity	2	Not applicable	Not applicable	0	$0 \leq 2$	Accepted
	TSA	Inclusivity	1	0	$0 \leq 1$	Not applicable	Not applicable	Accepted
		Exclusivity	2	Not applicable	Not applicable	0	$0 \leq 2$	Accepted
	CCI	Inclusivity	1	0	$0 \leq 1$	Not applicable	Not applicable	Accepted
		Exclusivity	2	Not applicable	Not applicable	0	$0 \leq 2$	Accepted
	ESIA	Inclusivity	1	0	$0 \leq 1$	Not applicable	Not applicable	Accepted
		Exclusivity	2	Not applicable	Not applicable	0	$0 \leq 2$	Accepted

4. Conclusions Method Comparison Study

For the inclusivity, the AL is met for both MBT Systems and for all the tested media, as the value for inclusivity deviation (ID) was lower than the fixed AL at 1 for 150 target strains tested.

For the exclusivity, the AL is met for both MBT Systems and for all the tested media, as the value for the exclusivity deviation (ED) was lower than the fixed AL at 2 for ≤103 (considering NOIP) non-target strains.

The data and interpretation confirm the reliability and robustness of the MALDI Biotyper as a confirmation method for *Campylobacter* spp and non-*Campylobacter* spp isolates from isolated colonies.

5 Interlaboratory Study

5.1 Protocol and Materials

A total of 17 participants from 7 different laboratories throughout United States and 1 laboratory in Canada. Five (5) of the laboratories had 2 separate analysts participate, two (2) laboratories had three (3) analysts participate and one (1) laboratory had 1 participant. Each collaborator analyzed 24 blind-coded organisms, 16 *Campylobacter* organisms and 8 non-*Campylobacter* organisms.

A detailed collaborative study packet outlining all necessary information related to the study including isolate preparation and documentation of results were sent to each collaborating laboratory prior to the initiation of the study. A detailed list of participants is presented in **Annex J**.

The list of the target strains (*Campylobacter* species) and the non-target strains (non-*Campylobacter*) evaluated by each participant is shown in **Table 10**. All organisms evaluated were well characterized prior to analysis using conventional procedures, additional molecular tests, or up to 500 base-pairs 16S rDNA sequencing (Accugenix®).

Table 10 - Organisms used during Inter-Laboratory Study

	Number	Genus	Species	Reference	Origin	Culture Collection
Campylobacter species organisms : inclusivity	1	<i>Campylobacter</i>	<i>coli</i>	QL ¹ 032817.1	Chicken Carcass	Q Laboratories
	2	<i>Campylobacter</i>	<i>coli</i>	ATCC ² 43481	Turkey Feces	Q Laboratories
	3	<i>Campylobacter</i>	<i>coli</i>	ATCC BAA-1061	Chicken Carcass	Q Laboratories
	4	<i>Campylobacter</i>	<i>coli</i>	CCUG ³ 10772	Pig Placenta	Q Laboratories
	5	<i>Campylobacter</i>	<i>coli</i>	CCUG 14537	Human Feces	Q Laboratories
	6	<i>Campylobacter</i>	<i>jejuni</i>	ATCC BAA-1062	Chicken Carcass	Q Laboratories
	7	<i>Campylobacter</i>	<i>jejuni</i>	QL 022117.1	Chicken Carcass	Q Laboratories
	8	<i>Campylobacter</i>	<i>jejuni</i>	QL 022117.2	Chicken Carcass	Q Laboratories
	9	<i>Campylobacter</i>	<i>jejuni</i>	QL 012516.11	Poultry	Q Laboratories
	10	<i>Campylobacter</i>	<i>jejuni</i>	QL 012516.12	Poultry	Q Laboratories
	11	<i>Campylobacter</i>	<i>jejuni</i>	QL 012516.13	Poultry	Q Laboratories
	12	<i>Campylobacter</i>	<i>jejuni</i>	QL 101998	Poultry House Environmental	Q Laboratories
	13	<i>Campylobacter</i>	<i>jejuni</i>	QL 040698	Turkey Thigh	Q Laboratories
	14	<i>Campylobacter</i>	<i>lari</i>	CCUG 15031	Human	Q Laboratories
	15	<i>Campylobacter</i>	<i>lari</i>	CCUG 15035	Sea Gull	Q Laboratories
	16	<i>Campylobacter</i>	<i>lari</i>	QL 040117.8	Chicken Carcass	Q Laboratories
Non-Campylobacter species Organisms:	1	<i>Citrobacter</i>	<i>farmeri</i>	ATCC 51633	Human Feces	Q Laboratories
	2	<i>Shimwellia</i>	<i>blattae</i>	ATCC 29907	Hindgut of Cockroach	Q Laboratories
	3	<i>Edwardsiella</i>	<i>tarda</i>	QL 11007.11	Clinical Isolate	Q Laboratories
	4	<i>Enterobacter</i>	<i>aerogenes</i>	ATCC 35029	Not Available	Q Laboratories
	5	<i>Escherichia</i>	<i>hermannii</i>	ATCC 33651	Human Arm Wound	Q Laboratories
	6	<i>Klebsiella</i>	<i>oxytoca</i>	ATCC 43165	Clinical Isolate	Q Laboratories
	7	<i>Proteus</i>	<i>mirabilis</i>	QL 11007.6	Clinical Isolate	Q Laboratories
	8	<i>Serratia</i>	<i>marcescens</i>	QL 11007.1	Bottled Water	Q Laboratories

¹ QL – Q Laboratories, Inc. Culture Collection

² ATCC – American Type Culture Collection

³ CCUG – University of Gothenburg Culture Collection

In order to demonstrate the equivalency between the reusable target and the disposable target, both were used by different participants during the evaluation. The reusable polished stainless steel target was able to be cleaned and reused and the disposable biotarget was for single use only. Refer to **Table 11** below for a list of participants who used each of the target plates.

Table 1 - Participation of each Collaborating Laboratory for Campylobacter

Collaborator ID #	Participating Laboratory	Analyst	Target	MBT Library
1	1	1	Disposable ^a	Version MSP 6903
2	1	2	Disposable	Version MSP 6903
3	2	1	Reusable ^b	Version MSP 6903
4	3	1	Disposable	Version MSP 6903
5	3	2	Disposable	Version MSP 6903
6	4	1	Disposable	Version MSP 6903
7	4	2	Disposable	Version MSP 6903
8	5	1	Reusable	Version MSP 6903
9	5	2	Reusable	Version MSP 6903
10	6	1	Reusable	Version MSP 6903
11	6	2	Reusable	Version MSP 6903
12	6	3	Reusable	Version MSP 6903
13	7	1	Reusable	Version MSP 6903
14	7	2	Reusable	Version MSP 6903
15	7	3	Reusable	Version MSP 6903
16	8	1	Reusable	Version MSP 6903
17	8	2	Reusable	Version MSP 6903

All isolates were labeled with a randomized and blind-coded three-digit number (both target and non-target organisms). Every isolate used in this evaluation was propagated onto Columbia Blood Agar (CBA, Hardy Diagnostics: Part#A16BX) from a Q Laboratories (Cincinnati, OH, USA) frozen stock culture collection stored at -70°C. Each organism was incubated for 24-48 hours at 41.5 ± 1°C under microaerobic conditions. Isolated colonies were streaked to CBA plates, and incubated for 24 - 48 hours at 41.5 ± 1°C under microaerobic conditions prior to being shipped to each site. Isolates were packaged into a BD GasPak™ EZ Campy Gas Generating Pouch System (Fisher: Part#B260685) in groups of 8 plates.

The differences between the MALDI Biotyper smart and the classical MALDI Biotyper are the MALDI Biotyper smart has a lifetime laser and it operates at a faster speed than the classical MALDI Biotyper.

5.2Transport

Isolates were shipped on a Saturday to arrive on Monday according to the Category B Dangerous Goods shipment regulations according to the International Air Transport Association (IATA). Upon receipt, each isolate was subcultured onto CBA for 24-48 hours at 41.5 ± 1°C under microaerobic conditions to ensure viable growth during the shipment and to re-isolate culturable growth.

5.3Analysis

From the CBA, each isolate (target and non-target) was subcultured onto four agars: CBA (Hardy Diagnostics or equivalent), mCCDA (Neogen or equivalent), RAPID' *Campylobacter* (Bio-Rad or equivalent) and Campy Cefex Agar (Neogen or equivalent). The selection of agars for the Interlaboratory Study is to align the method with the cultural reference methods and a chromogenic agar. The plates were incubated for 48 hours at 41.5 ± 1°C under micro-aerobic conditions. Following incubation, growth present on each plate was analyzed by the Bruker MALDI Biotyper using the direct transfer technique (DT). When needed, the extended direct

transfer (eDT) procedure was used. If necessary, the tube extraction (EXT) was used to obtain a reliable result. In order to demonstrate the equivalency between the reusable target and the disposable target, both were used by different participants during the evaluation.

Growth from the CBA plate were also evaluated following the ISO reference method: corkscrew morphology, darting motility, a spot oxidase test (Thermo Fisher Scientific or equivalent) and was struck to a fresh CBA plate and incubated under aerobic conditions at $25 \pm 1^\circ\text{C}$.

All the isolates recovered on CBA were tested by both methods, producing a complete data set.

5.4 Statistics and Results

The data obtained during the Interlaboratory Study were received from a total of 17 participants, excluding the expert laboratory. The raw data from each participant are provided in **Annex K**.

Some issues were observed for multiple participants during the Interlaboratory Study. A participant (#3) was not able to complete the full workflow, due to the fact they had an unexpected emergency schedule change. Participant 3, therefore, did not conduct testing on mCCDA, RAPID[®] *Campylobacter* Agar, and Campy Cefex Agar. Two participants (#8 and 15) were not able to obtain viable growth for the three *Campylobacter* isolates from mCCDA. One participant (#11) was unable to obtain growth for 2 *Campylobacter lari* isolates on RAPID[®] *Campylobacter* Agar and one participant (#15) was not able to obtain viable growth of a *Campylobacter jejuni* isolate from Campy Cefex Agar. Two of the participants (#6 & 7) did not test from the mCCDA as they obtained limited to no growth for each isolate. During the testing, participants 6 and 7 required a software patch update to allow for use of disposable target, this slightly delayed testing for these 2 participants. This delay may have resulted in the recovery issues with mCCDA and, therefore, that data for mCCDA was removed. The data generated for the additional agars is included in the overall results for participants 6 and 7.

A summary of responses from each participant are described below in **Table 12**. A summary of results is provided in **Tables 13-16**.

Table 12. Presentation of comments from Participants

Collaborator	Comments		Collaborator retained for interpretation
	Reference Method	Alternative Method	
Collaborator 1	Weak Agglutination Reaction	No comment	Yes
Collaborator 2	Weak Agglutination Reaction	No comment	Yes
Collaborator 3	Weak Agglutination Reaction	Unable to complete workflow for mCCDA, RAPID' <i>Campylobacter</i> agar, and Campy Cefex Agar	Yes, for CBA only
Collaborator 4	Weak Agglutination Reaction	No comment	Yes
Collaborator 5	Weak Agglutination Reaction	No comment	Yes
Collaborator 6	Weak Agglutination Reaction	Software patch required for disposable target, non-viable growth obtained on mCCDA only	Yes, for CBA, RAPID' <i>Campylobacter</i> Agar and Campy Cefex Agar only
Collaborator 7	Weak Agglutination Reaction	Software patch required for disposable target, non-viable growth obtained on mCCDA only	Yes, for CBA, RAPID' <i>Campylobacter</i> Agar and Campy Cefex Agar only
Collaborator 8	Weak Agglutination Reaction	2 <i>Campylobacter</i> strains with no growth on mCCDA	Yes, for CBA, RAPID' <i>Campylobacter</i> Agar and Campy Cefex Agar only
Collaborator 9	Weak Agglutination Reaction	No comment	Yes
Collaborator 10	Weak Agglutination Reaction	No comment	Yes
Collaborator 11	Weak Agglutination Reaction	2 <i>Campylobacter</i> strains with no growth on RAPID' <i>Campylobacter</i> agar	Yes, for CBA, mCCDA and Campy Cefex Agar
Collaborator 12	Weak Agglutination Reaction	No comment	Yes
Collaborator 13	Weak Agglutination Reaction	No comment	Yes
Collaborator 14	Weak Agglutination Reaction	No comment	Yes
Collaborator 15	Weak Agglutination Reaction	1 <i>Campylobacter</i> strain with no growth on mCCDA and 1 <i>Campylobacter</i> strain with no growth on Campy Cefex Agar.	Yes, for CBA and RAPID' <i>Campylobacter</i> Agar only
Collaborator 16	Weak Agglutination Reaction	No comment	Yes
Collaborator 17	Weak Agglutination Reaction	No comment	Yes

Table 2. Presentation of the results on CBA

Collaborators	Number of correctly confirmed strains in the inclusivity part (PA)		Number of correctly non-confirmed strains in the exclusivity part (NA)	
	Reference confirmation procedure	Alternative confirmation procedure	Reference confirmation procedure	Alternative confirmation procedure
Collaborator 1	16/16	16/16	8/8	8/8
Collaborator 2	16/16	16/16	8/8	8/8
Collaborator 3	16/16	16/16	8/8	8/8
Collaborator 4	16/16	16/16	8/8	8/8
Collaborator 5	16/16	16/16	8/8	8/8
Collaborator 6	16/16	16/16	8/8	8/8
Collaborator 7	16/16	16/16	8/8	8/8
Collaborator 8	16/16	16/16	8/8	8/8
Collaborator 9	16/16	16/16	8/8	8/8
Collaborator 10	16/16	16/16	8/8	8/8
Collaborator 11	16/16	16/16	8/8	8/8
Collaborator 12	16/16	16/16	8/8	8/8
Collaborator 13	16/16	16/16	8/8	8/8
Collaborator 14	16/16	16/16	8/8	8/8
Collaborator 15	16/16	16/16	8/8	8/8
Collaborator 16	16/16	16/16	8/8	8/8
Collaborator 17	16/16	16/16	8/8	8/8
Total	272/272	272/272	136/136	136/136
17 Collaborators	272/272	272/272	136/136	136/136

Table 3. Presentation of the results on mCCDA

Collaborators ⁷	Number of correctly confirmed strains in the inclusivity part (PA)	Number of correctly non-confirmed strains in the exclusivity part (NA)
	Alternative confirmation procedure	Alternative confirmation procedure
Collaborator 1	16/16	8/8 (4 with no growth)
Collaborator 2	16/16	8/8 (2 with no growth)
Collaborator 4	16/16	8/8 (1 with no growth)
Collaborator 5	16/16	8/8 (1 with no growth)
Collaborator 9	16/16	8/8 (1 with no growth)
Collaborator 10	16/16	8/8 (4 with no growth)
Collaborator 11	16/16	8/8 (2 with no growth)
Collaborator 12	16/16	8/8 (4 with no growth)
Collaborator 13	16/16	8/8 (1 with no growth)
Collaborator 14	16/16	8/8 (2 with no growth)
Collaborator 16	16/16	8/8 (2 with no growth)
Collaborator 17	16/16	8/8 (3 with no growth)
Total	192/192	96/96
12 Collaborators	192/192	96/96

⁷ Data from 5 collaborators were excluded from analysis due to technical deviations from the test protocol or non-viable growth was obtained.

Table 15. Presentation of the results on RAPID[®] Campylobacter Agar

Collaborators ⁸	Number of correctly confirmed strains in the inclusivity part (PA)	Number of correctly non-confirmed strains in the exclusivity part (NA)
	Alternative confirmation procedure	Alternative confirmation procedure
Collaborator 1	16/16	8/8 (7 with no growth)
Collaborator 2	16/16	8/8 (6 with no growth)
Collaborator 4	16/16	8/8 (7 with no growth)
Collaborator 5	16/16	8/8 (7 with no growth)
Collaborator 6	16/16	8/8 (6 with no growth)
Collaborator 7	16/16	8/8 (6 with no growth)
Collaborator 8	16/16	8/8 (7 with no growth)
Collaborator 9	16/16	8/8 (5 with no growth)
Collaborator 10	16/16	8/8 (8 with no growth)
Collaborator 12	16/16	8/8 (7 with no growth)
Collaborator 13	16/16	8/8 (8 with no growth)
Collaborator 14	16/16	8/8 (8 with no growth)
Collaborator 15	16/16	8/8 (8 with no growth)
Collaborator 16	16/16	8/8 (6 with no growth)
Collaborator 17	16/16	8/8 (8 with no growth)
Total	240/240	120/120
15 Collaborators	240/240	120/120

Table 4. Presentation of the results on Campy Cefex Agar

Collaborators ⁸	Number of correctly confirmed strains in the inclusivity part (PA)	Number of correctly non-confirmed strains in the exclusivity part (NA)
	Alternative Confirmation Procedure	Alternative confirmation procedure
Collaborator 1	16/16	8/8 (7 with no growth)
Collaborator 2	16/16	8/8 (4 with no growth)
Collaborator 4	16/16	8/8 (1 with no growth)
Collaborator 5	16/16	8/8 (1 with no growth)
Collaborator 6	16/16	8/8 (1 with no growth)
Collaborator 7	16/16	8/8 (2 with no growth)
Collaborator 8	16/16	8/8 (5 with no growth)
Collaborator 9	16/16	8/8 (4 with no growth)
Collaborator 10	16/16	8/8 (3 with no growth)
Collaborator 11	16/16	8/8 (8 with no growth)
Collaborator 12	16/16	8/8 (4 with no growth)
Collaborator 13	16/16	8/8 (8 with no growth)
Collaborator 14	16/16	8/8 (6 with no growth)
Collaborator 16	16/16	8/8 (1 with no growth)
Collaborator 17	16/16	8/8 (2 with no growth)
Total	240/240	120/120
15 Collaborators	240/240	120/120

⁸ Data from 2 collaborators were excluded from analysis due to technical deviations from the test protocol or non-viable growth was obtained.

The reference method produced correct confirmation rate for all the inclusivity and exclusivity panels from CBA. All the Campylobacter and the non-Campylobacter strains were correctly identified at the species level with the MALDI Biotyper from all the tested cultures plates, producing a confirmation rate of 100%. The summary of results and the statistical analysis from the Interlaboratory Study are summarized below in **Tables 17 and 18**.

Table 5. Summary of results for Interlaboratory Study

Media tested	Number of labs	Study	N	IA	ID	EA*	ED
CBA	17	Inclusivity	272	272	0	Not Applicable	Not Applicable
	17	Exclusivity	136	Not Applicable	Not Applicable	136	0
mCCDA	12	Inclusivity	192	192	0	Not Applicable	Not Applicable
	12	Exclusivity	96	Not Applicable	Not Applicable	69	0
RAPID' Campylobacter Agar	15	Inclusivity	240	240	0	Not Applicable	Not Applicable
	15	Exclusivity	120	Not Applicable	Not Applicable	16	0
Campy Cefex Agar	15	Inclusivity	240	240	0	Not Applicable	Not Applicable
	15	Exclusivity	120	Not Applicable	Not Applicable	63	0

*Studies with EA<N for each medium are due to no growth

Table 6. Statistical Analysis for the Interlaboratory Study

Media tested	Number of labs	Study	N	AL	ID	ID≤AL	ED	ED≤AL	Evaluation
CBA	17	Inclusivity	272	3	0	0≤3	Not Applicable	Not Applicable	Accepted
	17	Exclusivity	136	3	Not Applicable	Not Applicable	0	0≤3	Accepted
mCCDA	12	Inclusivity	192	2	0	0≤2	Not Applicable	Not Applicable	Accepted
	12	Exclusivity	96	2	Not Applicable	Not Applicable	0	0≤2	Accepted
RAPID' Campylobacter Agar	15	Inclusivity	240	3	0	0≤3	Not Applicable	Not Applicable	Accepted
	15	Exclusivity	120	3	Not Applicable	Not Applicable	0	0≤3	Accepted
Campy Cefex Agar	15	Inclusivity	240	3	0	0≤3	Not Applicable	Not Applicable	Accepted
	15	Exclusivity	120	3	Not Applicable	Not Applicable	0	0≤3	Accepted

According to ISO 16140-6, the AL was met for all four media types evaluated during the Interlaboratory Study indicating that the alternative method is fit for purpose.

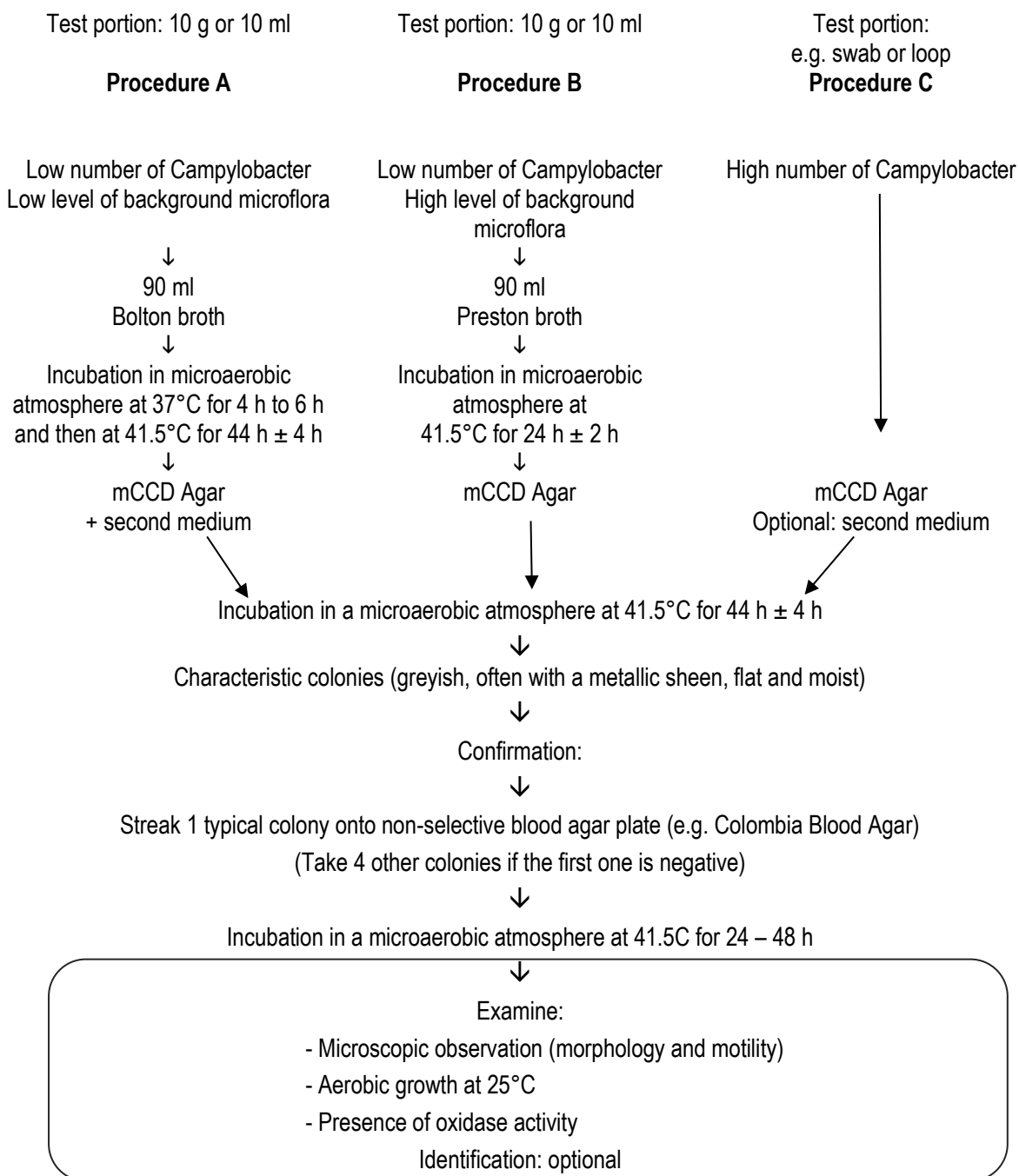
6 Conclusions of Interlaboratory Study

The AL is met for the four tested media during the interlaboratory study (Columbia Blood Agar, mCCDA, Campy Cefex and RAPID'*Campylobacter*), as the value for inclusivity deviation (ID) was lower than the AL, as well as the value for exclusivity deviation (ED) for the exclusivity study.

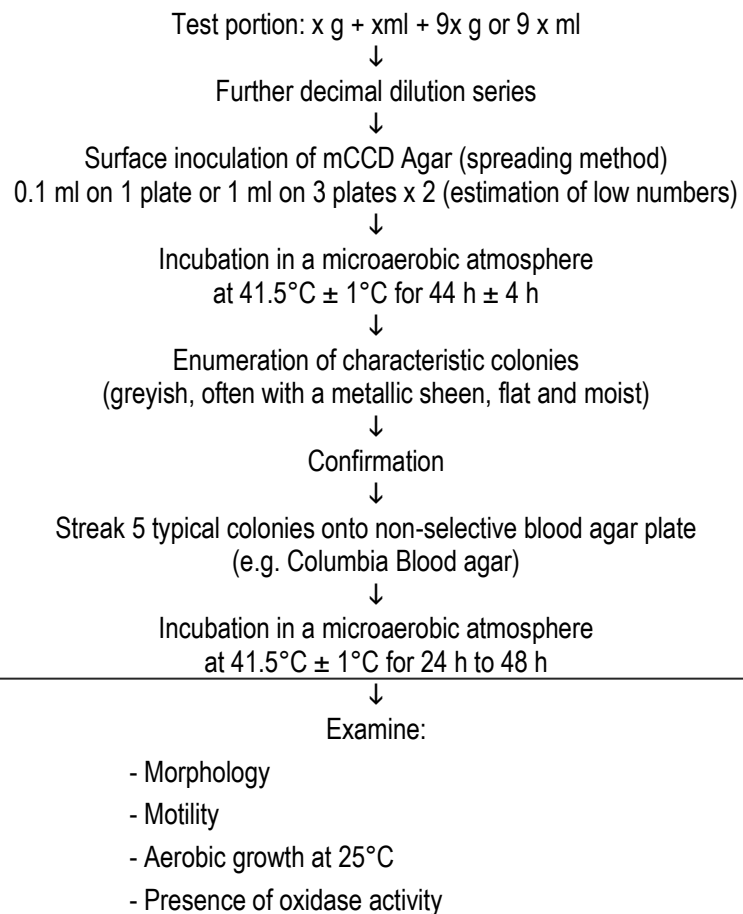
The data and interpretation confirm the reliability and robustness of the MALDI Biotyper as a confirmation method for *Campylobacter* spp.

**Annex A - Flow diagram of the reference method: ISO 10272 (June 2017) -
Microbiology of the food chain - Horizontal method for detection and
enumeration of *Campylobacter* spp.**

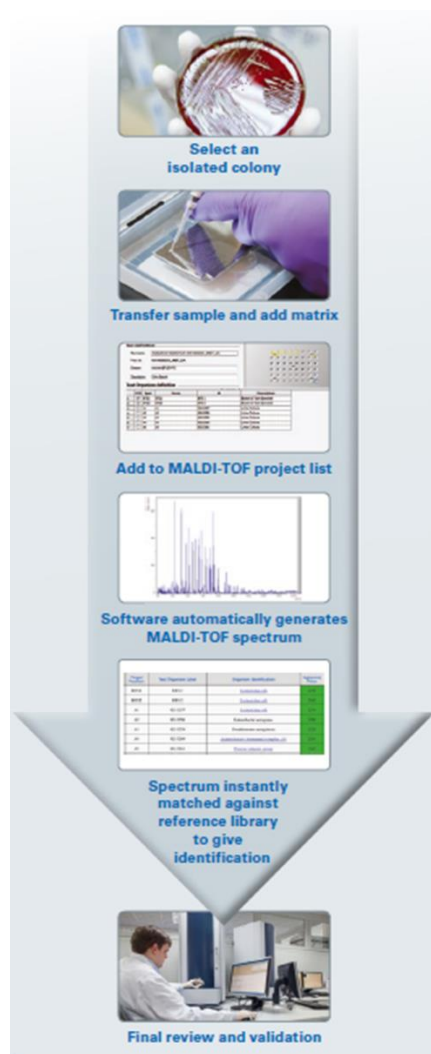
Part 1: detection method

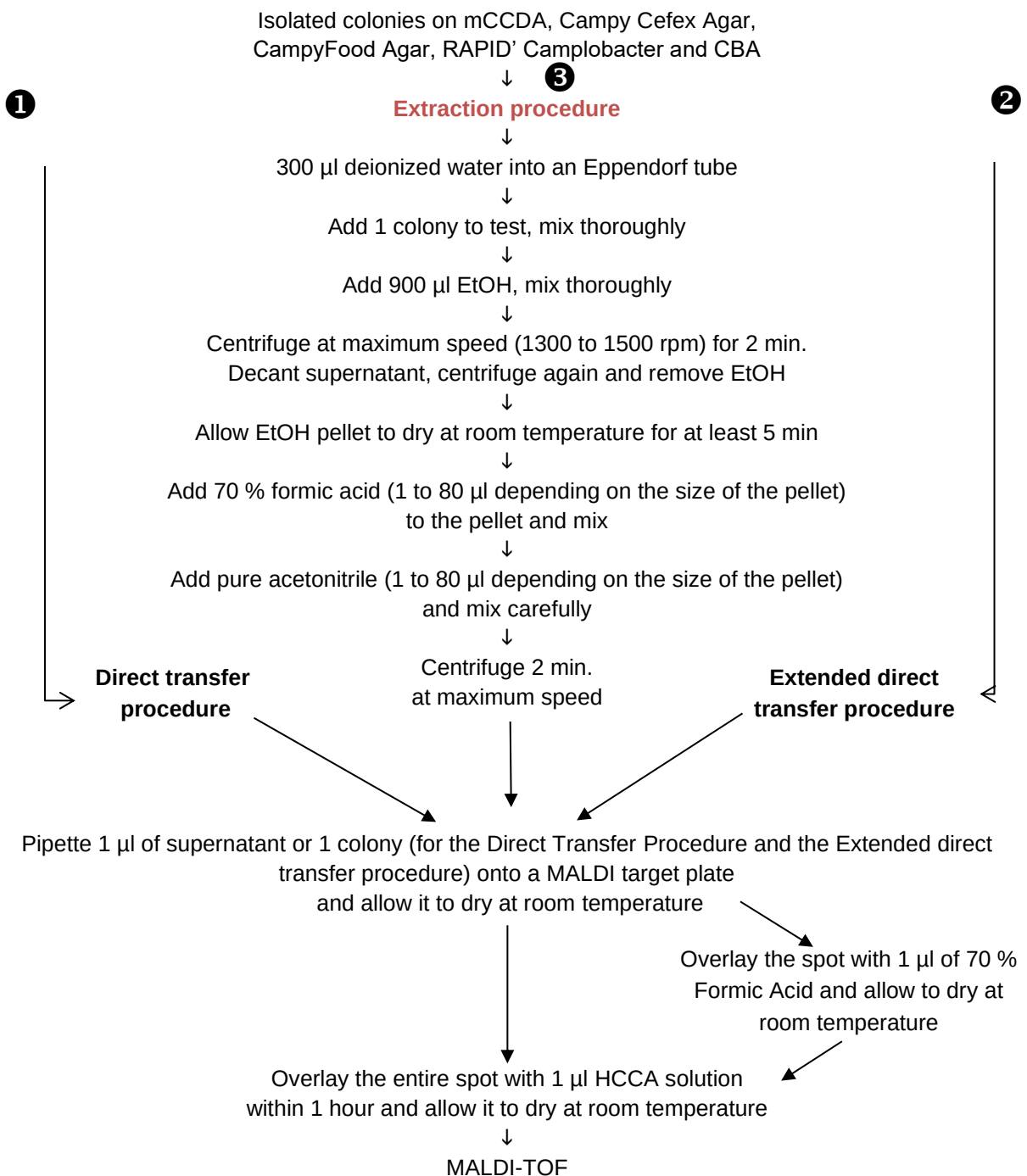


Part 2: Colony-count technique



Annex B - Flow diagram of the alternative method





The Direct Transfer (DT) Sample preparation can identify 90%-95% of the bacterial isolates. When no identification is obtained, this procedure should be repeated. If the no result is repeated, then the users should run the eDT. When needed the EXT sample preparation should be carried out. In some rare cases, a sub-culture on a nonselective agar may be required.

Annex C - Kit inserts

MALDI Biotyper



Standard Operating Procedure

Direct Transfer (DT) Procedure

1. Required Chemicals

- Deionized water
- Acetonitrile (ACN)
- Trifluoroacetic acid (TFA)
- Matrix HCCA; portioned # 8255344 (Referred to as HCCA in this SOP)
- Standard solvent (acetonitrile 50%, water 47.5% and trifluoroacetic acid 2.5%) from Sigma-Aldrich (# 19182), which has been tested by Bruker Daltonik GmbH and is recommended for solubilization of HCCA)¹
- Wooden application sticks, pipet tips or plastic inoculation loop to apply material
- 50–1000 µL pipet tips and a suitable pipet
- 2–200 µL pipet tips and a suitable pipet
- 0.5–10 µL pipet tips and a suitable pipet
- MALDI target plate

Note: Use only Eppendorf tubes (e.g. article no. 0030 120.086) and Eppendorf tips. Plasticizers may be released from other materials.

Note: Only use chemicals of highest purity (suitable for MALDI or HPLC).

Note: Please ensure that the BDAL Standard Library is used.

2. Required Laboratory Instrument

- Vortexer (Vortex Genie 2; # 9730060; Fisher Scientific or similar)

3. General Cultivation Conditions

Cultivation conditions have little effect on the identification. Media such as Columbia-Blood-Agar, Chocolate-Agar, or others can be used regardless of different growth phases or temperatures.

These different cultivation conditions produce only very small variations in observed peaks. Nearly all peaks are reproducible. Nevertheless the same medium and growth conditions should always be used for the cultivation of microorganisms.

If possible freshly grown material (grown overnight) should be used, or in the case of slow-growing bacteria, grown for several days. If cultivation plates are stored at 4°C the quality of spectra deteriorates relatively quickly (within a couple of days). Storing the plates at room temperature for several days is acceptable.

¹ Use commercially available standard solvent or mix 475 µL HPLC-grade water, 25 µL trifluoroacetic acid, and 500 µL acetonitrile in a 1.5 mL Eppendorf tube to produce 1 mL standard solvent. Store aliquots in screw cap tubes with screw cap tightly closed.

MALDI Biotyper



4. To prepare the HCCA matrix solution

1. Add 250 μ L standard solvent to a tube of HCCA.
2. Dissolve the HCCA by vortexing at room temperature until the solution is clear.

5. Direct Transfer Procedure:

1. Smear bacteria or yeast (single colony) as a thin film directly onto a spot on a MALDI target plate.

Note: Use only a small amount of material for direct transfer. Even if only a little biological material can be observed on the target, this is sufficient for measurement. For examples showing the correct amount of biological material please refer to the “**Demonstration Section**” at the end of this document.

2. Overlay the spot with 1 μ L of HCCA solution within 1 hour and allow to dry at room temperature.

6. Demonstration Section – “how much material is enough”

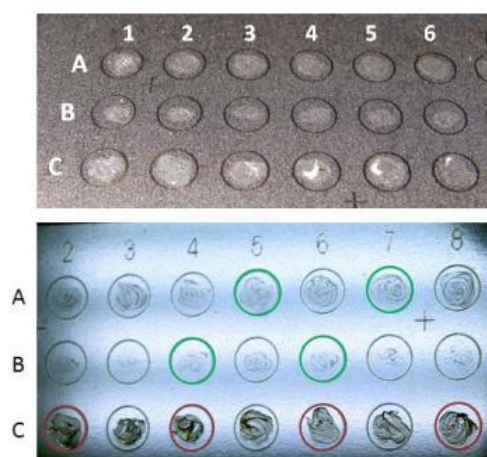


Fig 1: Different amounts of biological material: both pictures show the same MALDI target after sample application and before matrix addition; (entire row A) “appropriate amount” of biological material, (entire row B) small but still sufficient amount of biological material, (entire row C) too much biological material. Circles mark the target positions which were visualised as shown in Fig. 3 after matrix addition (as seen in flexControl).

MALDI Biotyper

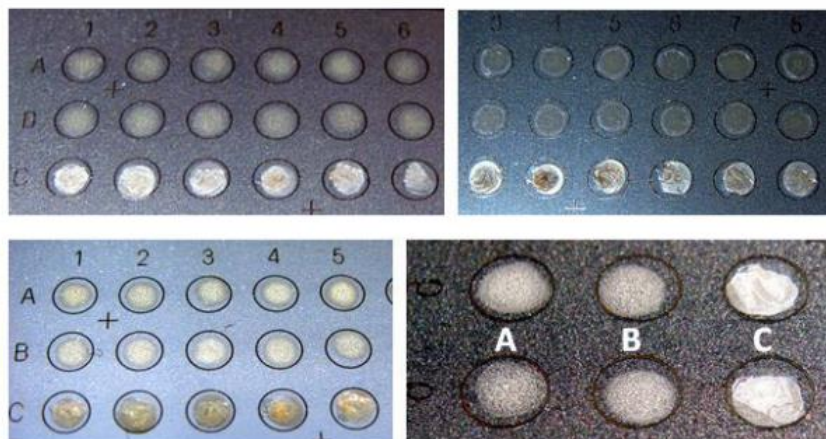


Fig 2: Same MALDI target as shown in Fig 1 after matrix application and drying.

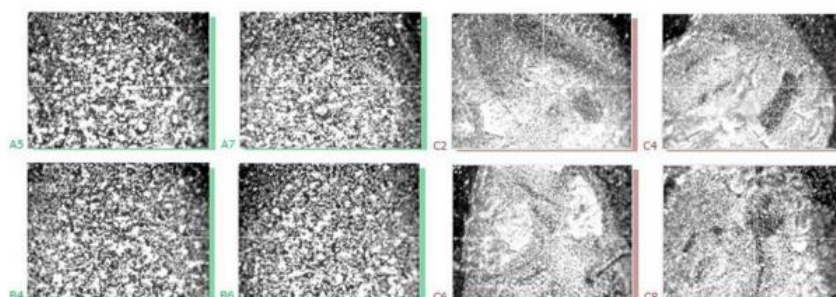


Fig 3: Illustration of crystal formation. "Camera image view" in flexControl:
A5, A7, B4, B6 demonstrate optimal crystal formation on spots where the correct or small (but still sufficient) amounts of biological material were smeared.
C2, C4, C6, C8 are examples for suboptimal crystal formation, due to excessive biological material applied in the direct transfer step, before application of the matrix.

All statements and applications pertaining to the MALDI Biotyper are for research use only.

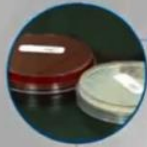
Please send comments and questions to biotyper.support@bdal.de

Descriptions and specifications supersede all previous information and are subject to change without notice.

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MALDI Biotyper

Direct Transfer Technique



- Select colony from fresh culture plate



- Smear onto target plate



- Cover with HCCA matrix
> dry
> analyze



- Pick isolated colony with any sterile transfer device

Standard Operating Procedure

Extended Direct Transfer (eDT) Procedure

1. Required Chemicals

- 70% formic acid (FA)
- Deionized water
- Acetonitrile (ACN)
- Trifluoroacetic acid (TFA)
- Matrix HCCA, portioned; # 8255344 (referred to as HCCA in this SOP)
- Standard solvent (acetonitrile 50%, water 47.5% and trifluoroacetic acid 2.5%) from Sigma-Aldrich (# 19182), which has been tested by Bruker Daltonik GmbH and is recommended for solubilization of HCCA)¹
- Wooden application sticks, pipet tips or plastic inoculation loop to apply material
- 50–1000 µL pipet tips and a suitable pipet
- 2–200 µL pipet tips and a suitable pipet
- 0.5–10 µL pipet tips and a suitable pipet
- MALDI target plate

Note: Use only Eppendorf tubes (e.g. article no. 0030 120.086) and Eppendorf tips. Plasticizers may be released from other materials.

Note: Only use chemicals of highest purity (suitable for MALDI or HPLC).

Note: Please ensure that the BDAL Standard Library is used.

2. Required Laboratory Instrument

- Vortexer (Vortex Genie 2; # 9730060; Fisher Scientific or similar)

3. General Cultivation Conditions

Cultivation conditions have little effect on the identification. Media such as Columbia-Blood-Agar, Chocolate-Agar, or others can be used regardless of different growth phases or temperatures.

These different cultivation conditions produce only very small variations in observed peaks. Nearly all peaks are reproducible. Nevertheless the same medium and growth conditions should always be used for the cultivation of microorganisms.

If possible freshly grown material (grown overnight) should be used, or in the case of slow-growing bacteria, grown for several days. If cultivation plates are stored at 4°C the quality of spectra deteriorates relatively quickly (within a couple of days). Storing the plates at room temperature for several days is acceptable.

¹ Use commercially available standard solvent or mix 475 µL HPLC-grade water, 25 µL trifluoroacetic acid, and 500 µL acetonitrile in a 1.5 mL Eppendorf tube to produce 1 mL standard solvent. Store aliquots in screw cap tubes with screw cap tightly closed.

MALDI Biotyper



4. To prepare the HCCA matrix solution

1. Add 250 μ L standard solvent to a tube of HCCA.
2. Dissolve the HCCA by vortexing at room temperature until the solution is clear.

5. Extended Direct Transfer Procedure:

1. Smear bacteria or yeast (single colony) as a thin film directly onto a spot on a MALDI target plate.

Note: Use only a small amount of material for direct transfer. Even if only a little biological material can be observed on the MALDI target plate, this is sufficient for measurement.

2. Overlay the spot with 1 μ L of 70% FA and allow to dry at room temperature.
3. Immediately overlay the spot with 1 μ L of HCCA solution and allow to dry at room temperature.

All statements and applications pertaining to the MALDI Biotyper are for research use only.

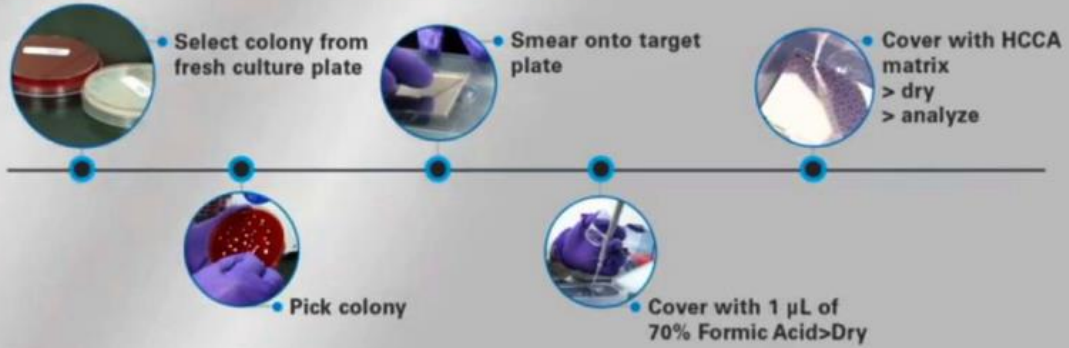
Please send comments and questions to biotyper.support@bdal.de

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MALDI Biotyper

Extended Direct Transfer Technique



Standard Operating Procedure

Extraction (Ext) Procedure

1. Required Chemicals and Accessories

- Deionized water
- Absolute ethanol (EtOH)
- Acetonitrile (ACN)
- Formic acid (70%)
- Trifluoroacetic acid (TFA)
- Matrix HCCA, portioned; # 8255344 (referred to as HCCA in this SOP)
- Standard solvent (acetonitrile 50%, water 47.5% and trifluoroacetic acid 2.5%) from Sigma-Aldrich (# 19182), which has been tested by Bruker Daltonik GmbH and is recommended for solubilization of HCCA)¹
- Wooden application sticks, pipet tips or plastic inoculation loop to apply material
- 50–1000 µL pipet tips and a suitable pipet
- 2–200 µL pipet tips and a suitable pipet
- 0.5–10 µL pipet tips and a suitable pipet
- Microtube (Eppendorf Safe-Lock), PCR clean 1.5 mL
- MALDI Target Plate

Note: Use only Eppendorf tubes (e.g. article no. 0030 120.086) and Eppendorf tips. Plasticizers may be released from other materials.

Note: Only use chemicals of highest purity (suitable for MALDI or HPLC).

Note: Please ensure that the BDAL Standard Library is used.

2. Required Laboratory Instruments

- Benchtop centrifuge (Biofuge fresco; # 75005510; Thermo Scientific or similar)
- Vortexer (Vortex Genie 2; # 9730060; Fisher Scientific or similar)

¹ Use commercially available standard solvent or mix 475 µL HPLC-grade water, 25 µL trifluoroacetic acid, and 500 µL acetonitrile in a 1.5 mL Eppendorf tube to produce 1 mL standard solvent. Store aliquots in screw cap tubes with screw cap tightly closed.

3. General Cultivation Conditions

Cultivation conditions have little effect on the identification. Media such as Columbia-Blood-Agar, Chocolate-Agar, or others can be used regardless of different growth phases or temperatures.

These different cultivation conditions produce only very small variations in observed peaks. Nearly all peaks are reproducible. Nevertheless the same medium and growth conditions should always be used for the cultivation of microorganisms.

If possible freshly grown material (grown overnight) should be used, or in the case of slow-growing bacteria, grown for several days. If cultivation plates are stored at 4°C the quality of spectra deteriorates relatively quickly (within a couple of days). Storing the plates at room temperature for several days is acceptable.

4. To prepare the HCCA matrix solution

1. Add 250 µL standard solvent to a tube of HCCA.
2. Dissolve the HCCA by vortexing at room temperature until the solution is clear.

5. Extraction Procedure:

1. Pipette 300 µL deionized water into an Eppendorf tube.
2. Place a quantity of biological material (between one single colony and 5-10 mg) into the tube. Mix thoroughly. Pipetting or vortexing should be appropriate.
3. Add 900 µL EtOH, mix thoroughly.
4. Centrifuge at max speed (13,000 to 15,000 rpm) for 2 minutes, decant supernatant, centrifuge again and remove all the residual EtOH by carefully pipetting it off to waste without disturbing the pellet.
5. Allow the EtOH-pellet to dry at room temperature for at least 5 minutes.
6. Add 70% formic acid (1 to 80 µL) to the pellet and mix very well by pipetting and/or by vortexing.

Note: If only a little material (one single colony) was used, the volume of formic acid must be adjusted accordingly.

	small single colony	large single colony	1 µL inoculating loop	10 µL inoculating loop
Formic Acid 70%	1–5 µL	10–20 µL	20–40 µL	>40–80 µL
Acetonitrile	1–5 µL	10–20 µL	20–40 µL	>40–80 µL

7. Add pure ACN (1 to 80 µL) and mix carefully. Use the same volume as formic acid!
8. Centrifuge for 2 minutes at maximum speed, such that all material is collected neatly in a pellet.
9. Pipette 1 µL of supernatant onto a MALDI target plate and allow it to dry at room temperature.
10. Overlay the entire spot with 1 µL of HCCA solution within 1 hour and allow it to dry at room temperature.

All statements and applications pertaining to the MALDI Biotyper are for research use only.

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Annex D – Certificate of equivalence



STATEMENT

Equivalency of MBT 4.0 and MBT Compass

Bruker Daltonik GmbH manufactures devices and reagents based on the MALDI-TOF-MS. This method has proven to be a reliable method for the classification and identification of microorganisms.

The microflex LT/SH and microflex LT/SH smart can be used qualitatively in conjunction with software, reference libraries, reagents and workflows. The MBT Compass software is equivalent to MBT 4.0 software in the following points:

- Instrument control software: flexControl
- Recorded mass range: 2,000 – 20,000 m/z
- Matching Algorithm (calculates matches by comparing a new spectrum against each single reference entry of a reference database)
- Current database
- Workflow
- Calibration and Quality Control
- The same rationale for data interpretation

Bruker Daltonik GmbH is certified according to ISO 9001:2008 and to ISO 13485:2012. Our devices are approved worldwide (including the European Union (IVD-CE Mark), the USA (FDA) and in China (CFDA) and are successfully established worldwide in the medical device market and for research and development and quality assurance purposes. The products of the Bruker Daltonik GmbH are manufactured to high quality.

A handwritten signature in blue ink, appearing to read "ppa. Dr. Wolfgang Pusch", written over a horizontal line.

ppa. Dr. Wolfgang Pusch
Executive Vice President
Business Area Clinical MALDI

A handwritten signature in blue ink, appearing to read "ppa. Dr. Markus Kostrzewa", written over a horizontal line.

ppa. Dr. Markus Kostrzewa
Vice President
Clinical Mass Spectrometry, R&D

Bremen, 2016-12-06

Innovation with Integrity

Method Comparison Study	
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Annex F - Raw data inclusivity study

See Excel File: 2017LR74 - Microval MCS Campylobacter (Version 1).xlsx; Tab
« Raw data Incl »

Annex G - Raw data exclusivity study

See Excel File: 2017LR74 - Microval MCS Campylobacter (Version 1).xlsx; Tab
« Raw data Excl »

Annex H - Results inclusivity study:

+: the strain was confirmed to be the target

-: the strain was not confirmed to be the target

NOIP: NO Organism Identification Possible

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-MS System														
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
1	Campylobacter	coli	Ad1004		Turkey skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
2	Campylobacter	coli	Ad1005		Turkey skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	
3	Campylobacter	coli	Ad1007		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	
4	Campylobacter	coli	Ad1008		Turkey skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	
5	Campylobacter	coli	Ad1009		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	
6	Campylobacter	coli	Ad1010		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID <i>Campylobacter</i>			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
7	<i>Campylobacter</i>	<i>coli</i>	Ad1011		Turkey skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
8	<i>Campylobacter</i>	<i>coli</i>	Ad1012		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
9	<i>Campylobacter</i>	<i>coli</i>	Ad1018		Chicken leg	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
10	<i>Campylobacter</i>	<i>coli</i>	Ad1024		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
11	<i>Campylobacter</i>	<i>coli</i>	Ad1025		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
12	<i>Campylobacter</i>	<i>coli</i>	Ad1072		Turkey neck skin	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
13	<i>Campylobacter</i>	<i>coli</i>	Ad1073		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-MS System														
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		CBA			mCCDA			CampyFood Agar			RAPID' <i>Campylobacter</i>			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
14	<i>Campylobacter</i>	<i>coli</i>	Ad1074		Turkey neck skin	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
15	<i>Campylobacter</i>	<i>coli</i>	Ad1075		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
16	<i>Campylobacter</i>	<i>coli</i>	Ad1077		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
17	<i>Campylobacter</i>	<i>coli</i>	Ad1087		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
18	<i>Campylobacter</i>	<i>coli</i>	Ad1123		Pork meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
19	<i>Campylobacter</i>	<i>coli</i>	Ad1477		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
20	<i>Campylobacter</i>	<i>coli</i>	Ad1478		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID' <i>Campylobacter</i>			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
21	<i>Campylobacter</i>	<i>coli</i>	Ad1479		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
22	<i>Campylobacter</i>	<i>coli</i>	Ad1480		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
23	<i>Campylobacter</i>	<i>coli</i>	Ad1481		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
24	<i>Campylobacter</i>	<i>coli</i>	Ad1485		Faecum	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
25	<i>Campylobacter</i>	<i>coli</i>	Ad1889		Pork environmental sample	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
26	<i>Campylobacter</i>	<i>coli</i>	Ad1894		Pork faecum	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
27	<i>Campylobacter</i>	<i>coli</i>	Ad1895		Pork faecum	Adria	+	+	PA	IA	NO IP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID' <i>Campylobacter</i>			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
28	<i>Campylobacter</i>	<i>coli</i>	Ad1897		Pork faecum	Adria	+	+	PA	IA	NO IP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
29	<i>Campylobacter</i>	<i>coli</i>	Ad1899		Pork faecum	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
30	<i>Campylobacter</i>	<i>coli</i>	Ad1900		Pork faecum	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
31	<i>Campylobacter</i>	<i>coli</i>	Ad1901		Chicken meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
32	<i>Campylobacter</i>	<i>coli</i>	Ad1902	<i>Campylobacter coli / jejuni</i>	Chicken meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
33	<i>Campylobacter</i>	<i>coli</i>	Ad1905		Leg duck	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
34	<i>Campylobacter</i>	<i>coli</i>	Ad1908		Duck meat	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID <i>Campylobacter</i>			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
35	<i>Campylobacter</i>	<i>coli</i>	Ad1959		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
36	<i>Campylobacter</i>	<i>coli</i>	Ad1960		Pork	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
37	<i>Campylobacter</i>	<i>coli</i>	Ad1961		Pork	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
38	<i>Campylobacter</i>	<i>coli</i>	Ad1962		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
39	<i>Campylobacter</i>	<i>coli</i>	Ad1963		Pork	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
40	<i>Campylobacter</i>	<i>coli</i>	Ad1964		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
41	<i>Campylobacter</i>	<i>coli</i>	Ad1965		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID' <i>Campylobacter</i>			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
42	<i>Campylobacter</i>	<i>coli</i>	Ad1967		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
43	<i>Campylobacter</i>	<i>coli</i>	Ad1968		Pork	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
44	<i>Campylobacter</i>	<i>coli</i>	Ad1969		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
45	<i>Campylobacter</i>	<i>jejuni</i>	Ad1002		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
46	<i>Campylobacter</i>	<i>jejuni</i>	Ad1003		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
47	<i>Campylobacter</i>	<i>jejuni</i>	Ad1013		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
48	<i>Campylobacter</i>	<i>jejuni</i>	Ad1015		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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49	<i>Campylobacter</i>	<i>jejuni</i>	Ad1016		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
50	<i>Campylobacter</i>	<i>jejuni</i>	Ad1021		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
51	<i>Campylobacter</i>	<i>jejuni</i>	Ad1023		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
52	<i>Campylobacter</i>	<i>jejuni</i>	Ad1082		Turkey neck skin	Adria	+	+	PA	IA	NO IP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
53	<i>Campylobacter</i>	<i>jejuni</i>	Ad1083		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
54	<i>Campylobacter</i>	<i>jejuni</i>	Ad1086		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
55	<i>Campylobacter</i>	<i>jejuni</i>	Ad1088		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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56	<i>Campylobacter</i>	<i>jejuni</i>	Ad1089		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
57	<i>Campylobacter</i>	<i>jejuni</i>	Ad1910	<i>Campylobacter jejuni / coli</i>	Chicken meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
58	<i>Campylobacter</i>	<i>jejuni</i>	Ad1922		Chicken	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
59	<i>Campylobacter</i>	<i>jejuni</i>	Ad1923		Chicken	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
60	<i>Campylobacter</i>	<i>jejuni</i>	Ad1947		Turkey	Adria	+	+	PA	IA	NO IP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
61	<i>Campylobacter</i>	<i>jejuni</i>	Ad1948		Turkey	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
62	<i>Campylobacter</i>	<i>jejuni</i>	Ad1949		Turkey	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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63	<i>Campylobacter</i>	<i>jejuni</i>	Ad1992		Wild crow	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
64	<i>Campylobacter</i>	<i>jejuni</i>	Ad1998		Poultry meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
65	<i>Campylobacter</i>	<i>lari</i>	Ad1067		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
66	<i>Campylobacter</i>	<i>lari</i>	Ad1130		Unknown	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
67	<i>Campylobacter</i>	<i>lari</i>	ATCC35222		Dog crap	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
68	<i>Campylobacter</i>	<i>upsaliensis</i>	Ad1139		Faeces	Adria	+	+	PA	IA	+	PA	IA	- (no growth)	NA (no growth)	IA	- (no growth)	NA (no growth)	NA (no growth)	+	PA	IA

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69	<i>Campylobacter</i>	<i>upsaliensis</i>	ATCC49815		Human feces	Adria	+	+	PA	IA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	IA	- (no growth)	NA (no growth)	NA (no growth)	+	PA	IA
70	<i>Campylobacter</i>	<i>upsaliensis</i>	ATCC49816		Human feces	Adria	+	+	PA	IA	+	PA	IA	- (no growth)	NA (no growth)	IA	- (no growth)	NA (no growth)	NA (no growth)	+	PA	IA
71	<i>Campylobacter</i>	<i>upsaliensis</i>	CIP103681		Dog crap	Adria	+	+	PA	IA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	IA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
72	<i>Campylobacter</i>	<i>subantarcticus</i>	Ad1891		Unknown	Adria	+	+	PA	IA	NO IP	No result	No result	+	PA	IA	NO IP	No result	No result	+	PA	IA
73	<i>Campylobacter</i>	<i>lari subsp. concheus</i>	Ad1911		Unknown	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
74	<i>Campylobacter</i>	<i>hyointestinalis</i>	Ad1898		Pork faeces	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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75	<i>Campylobacter</i>	<i>hyointestinalis</i>	Ad1896		Pork faeces	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
76	<i>Campylobacter</i>	<i>coli</i>	QL 032817.1		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
77	<i>Campylobacter</i>	<i>coli</i>	ATCC 43481		Turkey feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
78	<i>Campylobacter</i>	<i>coli</i>	ATCC BAA-1061		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
79	<i>Campylobacter</i>	<i>coli</i>	QL S810S		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
80	<i>Campylobacter</i>	<i>coli</i>	QL A1-10111999		Turkey	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
81	<i>Campylobacter</i>	<i>coli</i>	QL CCS08-03		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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82	<i>Campylobacter</i>	<i>coli</i>	QL CC01A		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
83	<i>Campylobacter</i>	<i>coli</i>	QL CC01B		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
84	<i>Campylobacter</i>	<i>coli</i>	QL CC02A		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
85	<i>Campylobacter</i>	<i>coli</i>	QL CC02B		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
86	<i>Campylobacter</i>	<i>coli</i>	CCUG 8320		Intestinal tract of swine	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
87	<i>Campylobacter</i>	<i>coli</i>	QL 011616.3		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
129	<i>Campylobacter</i>	<i>coli</i>	QL 031016.8		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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130	<i>Campylobacter</i>	<i>coli</i>	CCUG 10772		Pig placenta	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
131	<i>Campylobacter</i>	<i>coli</i>	CCUG 14537		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
132	<i>Campylobacter</i>	<i>coli</i>	CCUG 15360		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
133	<i>Campylobacter</i>	<i>coli</i>	CCUG 15362		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
134	<i>Campylobacter</i>	<i>coli</i>	CCUG 10955		Porcine feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
135	<i>Campylobacter</i>	<i>coli</i>	CCUG 10960		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
136	<i>Campylobacter</i>	<i>coli</i>	QL 013017.7		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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88	<i>Campylobacter</i>	<i>jejuni</i>	ATCC BAA-1062		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
89	<i>Campylobacter</i>	<i>jejuni</i>	QL 022117.1		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
90	<i>Campylobacter</i>	<i>jejuni</i>	QL 022117.2		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
91	<i>Campylobacter</i>	<i>jejuni</i>	QL AL03-112498		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
92	<i>Campylobacter</i>	<i>jejuni</i>	QL AL61-101998		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
93	<i>Campylobacter</i>	<i>jejuni</i>	QL AH03-040698		Turkey Thigh	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
94	<i>Campylobacter</i>	<i>jejuni</i>	QL GH12-062298		Turkey Neck	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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95	<i>Campylobacter</i>	<i>jejuni</i>	QL GH22-062298		Turkey	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
96	<i>Campylobacter</i>	<i>jejuni</i>	QL GH50-062298		Turkey	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
97	<i>Campylobacter</i>	<i>jejuni</i>	QL GHP20		Turkey	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
98	<i>Campylobacter</i>	<i>jejuni</i>	QL AHP14-040798		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
99	<i>Campylobacter</i>	<i>jejuni</i>	QL CH70-012599		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
100	<i>Campylobacter</i>	<i>jejuni</i>	QL BL-021599		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
101	<i>Campylobacter</i>	<i>jejuni</i>	QL BLP03-030599		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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102	<i>Campylobacter</i>	<i>jejuni</i>	QL BHP21-061498		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
103	<i>Campylobacter</i>	<i>jejuni</i>	QL BH65-081998		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
104	<i>Campylobacter</i>	<i>jejuni</i>	QL F-E4-23-112010		Turkey Neck	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
105	<i>Campylobacter</i>	<i>jejuni</i>	QL W-E2-25(2)-062011		Turkey Skin	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
106	<i>Campylobacter</i>	<i>jejuni</i>	QL F2D4-14F-112010		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
107	<i>Campylobacter</i>	<i>jejuni</i>	QL F2A2-7W		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
108	<i>Campylobacter</i>	<i>jejuni</i>	QL F2D4-4F		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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109	<i>Campylobacter</i>	<i>jejuni</i>	QL F2C4-24R		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
110	<i>Campylobacter</i>	<i>jejuni</i>	QL F3D4-19F		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
111	<i>Campylobacter</i>	<i>jejuni</i>	QL A1-08091999		Turkey Thigh	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
112	<i>Campylobacter</i>	<i>jejuni</i>	QL C01		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
113	<i>Campylobacter</i>	<i>jejuni</i>	QL I02-1		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
114	<i>Campylobacter</i>	<i>jejuni</i>	QL M03		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
115	<i>Campylobacter</i>	<i>jejuni</i>	QL CC5-1		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
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N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		CBA			mCCDA			CampyFood Agar			RAPID' <i>Campylobacter</i>			Campy Cefex Agar		
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116	<i>Campylobacter</i>	<i>jejuni</i>	QL I4568-060601-SH06		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
117	<i>Campylobacter</i>	<i>jejuni</i>	QL I13262-090703-018-03-23-4-02		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
118	<i>Campylobacter</i>	<i>jejuni</i>	QL I12572-02-07-02-TU01		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
119	<i>Campylobacter</i>	<i>jejuni</i>	QL D1-39-FT1-42		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
120	<i>Campylobacter</i>	<i>jejuni</i>	QL 012516.12		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
121	<i>Campylobacter</i>	<i>jejuni</i>	QL C1-13F-T2-42		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
122	<i>Campylobacter</i>	<i>jejuni</i>	QL 012516.11		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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								CBA			mCCDA			CampyFood Agar			RAPID' <i>Campylobacter</i>			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
1 2 3	<i>Campylobacter</i>	<i>lari</i>	CCUG 19512		Veterinary	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
1 2 4	<i>Campylobacter</i>	<i>lari</i>	QL 021216.3		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
1 2 5	<i>Campylobacter</i>	<i>lari</i>	ATCC BAA-1060		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
1 2 6	<i>Campylobacter</i>	<i>lari</i>	CCUG 20707		Veterinary	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
1 2 7	<i>Campylobacter</i>	<i>lari</i>	CCUG 22395		Herring gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
1 2 8	<i>Campylobacter</i>	<i>lari</i>	CCUG 22396		Herring gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
1 3 7	<i>Campylobacter</i>	<i>lari</i>	CCUG 24266		Veterinary	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
138	<i>Campylobacter</i>	<i>lari</i>	QL 022216.5		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
139	<i>Campylobacter</i>	<i>lari</i>	CCUG 15031		Human	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
140	<i>Campylobacter</i>	<i>lari</i>	CCUG 15035		Sea gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
141	<i>Campylobacter</i>	<i>lari</i>	CCUG 12773		Unknown	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
142	<i>Campylobacter</i>	<i>lari</i>	CCUG 12774		Human child feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
143	<i>Campylobacter</i>	<i>lari</i>	CCUG 19528		Human	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
144	<i>Campylobacter</i>	<i>lari</i>	QL 040117.8		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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145	<i>Campylobacter</i>	<i>lari</i>	CCUG 20581		Human	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
146	<i>Campylobacter</i>	<i>lari</i>	CCUG 29405		Herring gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
147	<i>Campylobacter</i>	<i>lari</i>	CCUG 29406		Veterinary	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
148	<i>Campylobacter</i>	<i>lari</i>	CCUG 55788		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
149	<i>Campylobacter</i>	<i>lari</i>	CCUG 55789		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
150	<i>Campylobacter</i>	<i>lari</i>	CCUG 19528		Herring gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
Total	N								150				150				150				150		
	PA								150				134				146				145		
	ND								0				0				0				0		

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	NA							0				0				0				0				
	NA (no growth)							0				2				4				4				1
	PD							0				0				0				0				0
	No result							0				14				0				1				0

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1	<i>Campylobacter</i>	<i>coli</i>	Ad1004		Turkey skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
2	<i>Campylobacter</i>	<i>coli</i>	Ad1005		Turkey skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
3	<i>Campylobacter</i>	<i>coli</i>	Ad1007		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
4	<i>Campylobacter</i>	<i>coli</i>	Ad1008		Turkey skin	Adria	+	+	PA	IA	NOIP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
5	<i>Campylobacter</i>	<i>coli</i>	Ad1009		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
6	<i>Campylobacter</i>	<i>coli</i>	Ad1010		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
7	<i>Campylobacter</i>	<i>coli</i>	Ad1011		Turkey skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
8	<i>Campylobacter</i>	<i>coli</i>	Ad1012		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
9	<i>Campylobacter</i>	<i>coli</i>	Ad1018		Chicken leg	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
10	<i>Campylobacter</i>	<i>coli</i>	Ad1024		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
11	<i>Campylobacter</i>	<i>coli</i>	Ad1025		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
12	<i>Campylobacter</i>	<i>coli</i>	Ad1072		Turkey neck skin	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
13	<i>Campylobacter</i>	<i>coli</i>	Ad1073		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
14	<i>Campylobacter</i>	<i>coli</i>	Ad1074		Turkey neck skin	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
15	<i>Campylobacter</i>	<i>coli</i>	Ad1075		Turkey neck skin	Adria	+	+	PA	IA	NOIP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
16	<i>Campylobacter</i>	<i>coli</i>	Ad1077		Turkey neck skin	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA

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17	Campylobacter	coli	Ad1087		Chicken neck skin	Adria	+	+	PA	IA	NoIP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
18	Campylobacter	coli	Ad1123		Pork meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
19	Campylobacter	coli	Ad1477		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
20	Campylobacter	coli	Ad1478		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
21	Campylobacter	coli	Ad1479		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
22	Campylobacter	coli	Ad1480		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
23	Campylobacter	coli	Ad1481		Carcass	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
24	Campylobacter	coli	Ad1485		Faecum	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
25	Campylobacter	coli	Ad1889		Pork environmental sample	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
26	Campylobacter	coli	Ad1894		Pork faecum	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
27	Campylobacter	coli	Ad1895		Pork faecum	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
28	Campylobacter	coli	Ad1897		Pork faecum	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
29	Campylobacter	coli	Ad1899		Pork faecum	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
30	Campylobacter	coli	Ad1900		Pork faecum	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
31	Campylobacter	coli	Ad1901		Chicken meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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32	Campylobacter	coli	Ad1902	Campylobacter coli / jejuni	Chicken meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
33	Campylobacter	coli	Ad1905		Leg duck	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
34	Campylobacter	coli	Ad1908		Duck meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
35	Campylobacter	coli	Ad1959		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
36	Campylobacter	coli	Ad1960		Pork	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
37	Campylobacter	coli	Ad1961		Pork	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
38	Campylobacter	coli	Ad1962		Pork	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
39	Campylobacter	coli	Ad1963		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
40	Campylobacter	coli	Ad1964		Pork	Adria	+	+	PA	IA	NOIP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
41	Campylobacter	coli	Ad1965		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
42	Campylobacter	coli	Ad1967		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
43	Campylobacter	coli	Ad1968		Pork	Adria	+	+	PA	IA	No peak	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
44	Campylobacter	coli	Ad1969		Pork	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
45	Campylobacter	jejuni	Ad1002		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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46	Campylobacter	jejuni	Ad1003		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
47	Campylobacter	jejuni	Ad1013		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
48	Campylobacter	jejuni	Ad1015		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
49	Campylobacter	jejuni	Ad1016		Chicken skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
50	Campylobacter	jejuni	Ad1021		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
51	Campylobacter	jejuni	Ad1023		Turkey neck skin	Adria	+	+	PA	IA	NOI P	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
52	Campylobacter	jejuni	Ad1082		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
53	Campylobacter	jejuni	Ad1083		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
54	Campylobacter	jejuni	Ad1086		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
55	Campylobacter	jejuni	Ad1088		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
56	Campylobacter	jejuni	Ad1089		Chicken neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
57	Campylobacter	jejuni	Ad1910	Campylobacter jejuni / coli	Chicken meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
58	Campylobacter	jejuni	Ad1922		Chicken	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
59	Campylobacter	jejuni	Ad1923		Chicken	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

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60	<i>Campylobacter</i>	<i>jejuni</i>	Ad1947		Turkey	Adria	+	+	PA	IA	NOIP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
61	<i>Campylobacter</i>	<i>jejuni</i>	Ad1948		Turkey	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
62	<i>Campylobacter</i>	<i>jejuni</i>	Ad1949		Turkey	Adria	+	+	PA	IA	NOIP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
63	<i>Campylobacter</i>	<i>jejuni</i>	Ad1992		Wild crow	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
64	<i>Campylobacter</i>	<i>jejuni</i>	Ad1998		Poultry meat	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
65	<i>Campylobacter</i>	<i>lari</i>	Ad1067		Turkey neck skin	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
66	<i>Campylobacter</i>	<i>lari</i>	Ad1130		Unknown	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
67	<i>Campylobacter</i>	<i>lari</i>	ATCC35222		Dog crap	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
68	<i>Campylobacter</i>	<i>upsaliensis</i>	Ad1139		Feces	Adria	+	+	PA	IA	+	PA	IA	- (no growth)	NA (no growth)	IA	- (no growth)	NA (no growth)	NA (no growth)	+	PA	IA
69	<i>Campylobacter</i>	<i>upsaliensis</i>	ATCC49815		Human feces	Adria	+	+	PA	IA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	IA	- (no growth)	NA (no growth)	NA (no growth)	+	PA	IA
70	<i>Campylobacter</i>	<i>upsaliensis</i>	ATCC49816		Human feces	Adria	+	+	PA	IA	+	PA	IA	- (no growth)	NA (no growth)	IA	- (no growth)	NA (no growth)	NA (no growth)	+	PA	IA
71	<i>Campylobacter</i>	<i>upsaliensis</i>	CIP103681		Dog crap	Adria	+	+	PA	IA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	IA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-smart System														
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		CBA			mCCDA			CampyFood Agar			RAPID/Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
72	Campylobacter	subantarcticus	Ad1891		Unknown	Adria	+	+	PA	IA	NOIP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
73	Campylobacter	lari subsp. concheus	Ad1911		Unknown	Adria	+	+	PA	IA	NOIP	No result	No result	+	PA	IA	+	PA	IA	+	PA	IA
74	Campylobacter	hyointestinalis	Ad1898		Pork feces	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
75	Campylobacter	hyointestinalis	Ad1896		Pork feces	Adria	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
76	Campylobacter	coli	QL 032817.1		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
77	Campylobacter	coli	ATCC 43481		Turkey feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
78	Campylobacter	coli	ATCC BAA-1061		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
79	Campylobacter	coli	QL S810S		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
80	Campylobacter	coli	QL A1-10111999		Turkey	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
81	Campylobacter	coli	QL CCS08-03		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
82	Campylobacter	coli	QL CC01A		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
83	Campylobacter	coli	QL CC01B		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
84	Campylobacter	coli	QL CC02A		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
85	Campylobacter	coli	QL CC02B		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-smart System														
								CBA			mCCDA			CampyFood Agar			RAPID Campylobacter			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
86	<i>Campylobacter</i>	<i>coli</i>	CCUG 8320		Intestinal tract of swine	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
87	<i>Campylobacter</i>	<i>coli</i>	QL 011616.3		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
129	<i>Campylobacter</i>	<i>coli</i>	QL 031016.8		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
130	<i>Campylobacter</i>	<i>coli</i>	CCUG 10772		Pig placenta	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
131	<i>Campylobacter</i>	<i>coli</i>	CCUG 14537		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
132	<i>Campylobacter</i>	<i>coli</i>	CCUG 15360		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
133	<i>Campylobacter</i>	<i>coli</i>	CCUG 15362		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
134	<i>Campylobacter</i>	<i>coli</i>	CCUG 10955		Porcine feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
135	<i>Campylobacter</i>	<i>coli</i>	CCUG 10960		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
136	<i>Campylobacter</i>	<i>coli</i>	QL 013017.7		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-smart System														
								CBA			mCCDA			CampyFood Agar			RAPID/Campylobacter			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
88	Campylobacter	jejuni	ATCC BAA-1062		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
89	Campylobacter	jejuni	QL 022117.1		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
90	Campylobacter	jejuni	QL 022117.2		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
91	Campylobacter	jejuni	QL AL03-112498		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
92	Campylobacter	jejuni	QL AL61-101998		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
93	Campylobacter	jejuni	QL AH03-040698		Turkey Thigh	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
94	Campylobacter	jejuni	QL GH12-062298		Turkey Neck	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
95	Campylobacter	jejuni	QL GH22-062298		Turkey	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
96	Campylobacter	jejuni	QL GH50-062298		Turkey	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
97	Campylobacter	jejuni	QL GHP20		Turkey	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
98	Campylobacter	jejuni	QL AHP14-040798		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
99	Campylobacter	jejuni	QL CH70-012599		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
100	Campylobacter	jejuni	QL BL-021599		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
101	Campylobacter	jejuni	QL BLP03-030599		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-smart System														
								CBA			mCCDA			CampyFood Agar			RAPID/Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
102	Campylobacter	jejuni	QL BHP21-061498		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
103	Campylobacter	jejuni	QL BH65-081998		Poultry House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
104	Campylobacter	jejuni	QL F-E4-23-112010		Turkey Neck	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
105	Campylobacter	jejuni	QL W-E2-25(2)-062011		Turkey Skin	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
106	Campylobacter	jejuni	QL F2D4-14F-112010		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
107	Campylobacter	jejuni	QL F2A2-7W		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
108	Campylobacter	jejuni	QL F2D4-4F		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
109	Campylobacter	jejuni	QL F2C4-24R		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
110	Campylobacter	jejuni	QL F3D4-19F		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
111	Campylobacter	jejuni	QL A1-08091999		Turkey Thigh	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
112	Campylobacter	jejuni	QL C01		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
113	Campylobacter	jejuni	QL I02-1		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
114	Campylobacter	jejuni	QL M03		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
115	Campylobacter	jejuni	QL CC5-1		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
116	Campylobacter	jejuni	QL I4568-060601-SH06		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
117	Campylobacter	jejuni	QL I13262-090703-018-03-23-4-02		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																						
Target Strain							Reference method	Alternative method														
								LT/SH MALDI-smart System														
								CBA			mCCDA			CampyFood Agar			RAPID/Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
118	Campylobacter	jejuni	QL I12572-02-07-02-TU01		Chicken Carcass Rinse	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
119	Campylobacter	jejuni	QL D1-39-FT1-42		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
120	Campylobacter	jejuni	QL 012516.12		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
121	Campylobacter	jejuni	QL C1-13F-T2-42		Slaughter House Environmental	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
122	Campylobacter	jejuni	QL 012516.11		Poultry	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
123	Campylobacter	lari	CCUG 19512		Veterinary	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
124	Campylobacter	lari	QL 021216.3		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
125	Campylobacter	lari	ATCC BAA-1060		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
126	Campylobacter	lari	CCUG 20707		Veterinary	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
127	Campylobacter	lari	CCUG 22395		Herring gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
128	Campylobacter	lari	CCUG 22396		Herring gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
127	Campylobacter	lari	CCUG 24266		Veterinary	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
138	Campylobacter	lari	QL 022216.5		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA
139	Campylobacter	lari	CCUG 15031		Human	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA

INCLUSIVITY																							
Target Strain							Reference method	Alternative method															
								LT/SH MALDI-smart System															
								CBA			mCCDA			CampyFood Agar			RAPID/Campylobacter			Campy Cefex Agar			
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	
140	Campylobacter	lari	CCUG 15035		Sea gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
141	Campylobacter	lari	CCUG 12773		Unknown	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
142	Campylobacter	lari	CCUG 12774		Human child feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
143	Campylobacter	lari	CCUG 19528		Human	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
144	Campylobacter	lari	QL 040117.8		Chicken carcass	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
145	Campylobacter	lari	CCUG 20581		Human	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
146	Campylobacter	lari	CCUG 29405		Herring gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
147	Campylobacter	lari	CCUG 29406		Veterinary	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
148	Campylobacter	lari	CCUG 55788		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
149	Campylobacter	lari	CCUG 55789		Human feces	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
150	Campylobacter	lari	CCUG 19528		Herring gull	Q Labs	+	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	+	PA	IA	
Total	N								150				150				150				150		
	PA								150				130				146				146		

INCLUSIVITY																						
Target Strain								Alternative method														
								LT/SH MALDI-smart System														
								CBA			mCCDA			CampyFood Agar			RAPID®Campylobacter			Campy Cefex Agar		
N	Species	Sub species	Reference	rDNA Sequencing	Origin	Lab	Reference method	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation			
	ND								0		0		0		0		0					
	NA							0	0		0		0		0							
	NA (no growth)							0	2		4		4		1							
	PD							0	0		0		0		0							
	No result							0	18		0		0		0							

Annex I - Results exclusivity study

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation			
1	Acinetobacter	baumanii	Ad1090		Hemoglobin	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
2	Acinetobacter	calcoaceticus	Ad1092		Poultry meat	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
3	Acinetobacter	calcoaceticus var. anitratus	1		Poultry	Adria	-	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

Optimal growth																							
EXCLUSIVITY																							
Non-Target Strain								Alternative method															
LT/SH MALDI-MS System																							
CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar											
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	
4	Acinetobacter	johnsonii	Ad1317		Whole egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
5	Aeromonas	allosaccharophile	Ad1318		Whole egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
6	Aeromonas	hydrophila	Ad1570		Water	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
7	Aeromonas	punctata	Ad1517		Whole liquid egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation			
8	<i>Aeromonas</i>	<i>salmonicida</i>	Ad1319		Whole liquid egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
9	<i>Alcaligenes</i>	<i>faecalis</i>	ATCC 8750		Unknown	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
10	<i>Arcobacter</i>	<i>butzleri</i>	Ad1126		Chicken	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
11	<i>Arcobacter</i>	<i>butzleri</i>	Ad1881		Environmental sample	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	-	NA	EA	-	NA	EA	-	NA	EA

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12	<i>Arcobacter</i>	<i>cryaerophilus</i>	Ad1124		Chicken	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
13	<i>Burkholderia</i>	<i>vietnamiensis</i>	Ad1538		Floor	Adria	-	-	NA	EA	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
14	<i>Burkholderia</i>	<i>cepacia</i>	Ad1541		Floor	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
15	<i>Candida</i>	<i>zeylanoides</i>	Ad2241		Process water	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

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								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
16	<i>Carnobacterium</i>	<i>maltaromaticum</i>	Ad1321		Liquid egg product	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
17	<i>Comamonas</i>	<i>aquatica</i>	Ad1543		Water	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
18	<i>Chryseobacterium</i>	<i>indologenes</i>	26		Whole egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
19	<i>Chryseobacterium</i>	<i>ureilyticum</i>	Ad1340		Whole egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

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20	Enterobacter	intermedius	88a	Raoultella planticola / ornithinolytica	Gizzard	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)		
21	Escherichia	fergusonii	2876		Environmental sample	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)		
22	Enterococcus	durans	Ad148	Enterococcus durans	Ham	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)		
23	Enterococcus	faecalis	25		Chicken meat	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)		

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24	<i>Enterococcus</i>	<i>faecium</i>	Ad1883		Turkey skin	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
25	<i>Enterococcus</i>	<i>gallinarum</i>	Ad1885		Poultry	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
26	<i>Escherichia</i>	<i>coli</i>	Ad1915		Chicken legs	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	-	NA	EA
27	<i>Escherichia</i> **	<i>fergusonii</i>	Ad1381		Water	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

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28	<i>Escherichia</i>	<i>coli</i>	CIP103982		Unknown	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	-(no growth)	NA (no growth)	NA (no growth)	-	NA	EA	
29	<i>Hafnia</i>	<i>alvei</i>	168		Duck meat	Adria	-	-	NA	EA	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	
30	<i>Klebsiella</i>	<i>oxytoca</i>	Ad1509		Milk powder	Adria	-	-	NA	EA	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	
31	<i>Klebsiella</i>	<i>pneumoniae</i>	47		Chicken skin	Adria	-	-	NA	EA	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	-(no growth)	NA (no growth)	NA (no growth)	

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32	<i>Lactobacillus</i>	<i>sakei</i>	85L905		Meat	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
33	<i>Myroides</i>	<i>odoratimimus</i>	Ad1341		Whole egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
34	<i>Ochrobactrum</i>	<i>pseudintermedium</i>	Ad1057		Turkey skin	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	-	NA	EA
35	<i>Photobacterium</i>	<i>phosphoreum</i>	Ad1506		Salmon	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

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36	<i>Plesiomonas</i>	<i>shigelloïdes</i>	Ad673		Fish	Adria	-	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
37	<i>Providencia</i>	<i>stuartii</i>	46		Poultry meat	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
38	<i>Pseudomonas</i>	<i>fragi</i>	Ad1327		Whole egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
39	<i>Pseudomonas</i>	<i>otitidis</i>	Ad1880		Skin meat	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)

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40	<i>Pseudomonas</i>	<i>putida</i>	Ad1331		Whole liquid egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
41	<i>Psychrobacter</i>	<i>psychrophilus</i>	Ad1343		Whole liquid egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
42	<i>Ralstonia</i>	<i>mannitolilytica</i>	Ad1059		Turkey skin	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	-	NA	EA	-	NA	EA
43	<i>Ralstonia</i>	<i>mannitolilytica</i>	Ad2504		Buffer solution	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

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44	<i>Rhizobium</i>	<i>radiobacter</i>	Ad1550		Water	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	
45	<i>Serratia</i>	<i>liquefaciens</i>	87a		Gizzard	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	
46	<i>Shewanella</i>	<i>putrefasciens</i>	EN15/34	<i>Shigella flexneri</i> / <i>sonnei</i> ; <i>Escherichia</i> ** <i>fergusonii</i>	Trout	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	

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47	Staphylococcus	aureus	Ad906		Sausage	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
48	Trichosporon	asahii	Ad2732		fromage frais	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
49	Vibrio	alginolyticus	Ad1888		Olster	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
50	Vibrio	cholerae	Ad1887		Fish filet	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

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51	Yersinia	enterocolitica	A00C066		Cockerel	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	-	NA	EA	-	NA	EA	
52	Aeromonas	hydrophila	ATCC 49140		Clinical Isolate	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	
53	Aerococcus	viridans	QL 17041.8		Raw Milk	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	
54	Citrobacter	amalonaticus	ATCC 25407		Feces	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	

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55	<i>Citrobacter</i>	<i>braakii</i>	ATCC 43162		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
56	<i>Citrobacter</i>	<i>farmeri</i>	ATCC 51633		Human Feces	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
57	<i>Citrobacter</i>	<i>freundii</i>	QL 11007.10		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
58	<i>Citrobacter</i>	<i>youngae</i>	ATCC 11102		Not Available	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	

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								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
59	<i>Cronobacter</i>	<i>malonaticus</i>	QL 17031.3		Infant formula	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
60	<i>Cronobacter</i>	<i>sakazakii</i>	QL 17031.4		Infant formula	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
61	<i>Cronobacter</i>	<i>sakazakii</i>	ATCC 51329		Rice flour	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
62	<i>Edwardsiella</i>	<i>tarda</i>	QL 11007.11		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
63	Enterobacter	aerogenes	ATCC 35029		Not Available	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
64	Enterobacter	amnigenus	ATCC 51816		Milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
65	Enterobacter	cancerogenus	QL110 10-1	Shigella, Escherichia, Enterobacter	Bottled Water	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
66	Enterobacter	cloacae	ATCC 23355		Spinal Fluid	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
LT/SH MALDI-MS System																						
CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar										
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		Reference method	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain
67	Enterococcus	faecalis	ATCC 51299		Peritoneal fluid	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
68	Enterococcus	faecium	ATCC 8459		Dairy Product	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
69	Escherichia	fergusonii	ATCC 35470	Escherichia fergussoni	Not Available	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
70	Escherichia	hermannii	ATCC 33651		Human Arm Wound	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
71	<i>Escherichia coli</i>	<i>coli</i> O157:H7	ATCC 43895		Raw Hamburger	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
72	<i>Escherichia coli</i>	<i>coli</i>	QL 11010.2		Bottled Water	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
73	<i>Escherichia coli</i>	<i>vulneris</i>	ATCC 29943	<i>Enterobacter</i>	Human Wound	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
74	<i>Havnia</i>	<i>alvei</i>	ATCC 51815		Milk	Q Labs	-	-	NA	EA	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation			
75	<i>Klebsiella</i>	<i>pneumoniae</i>	ATCC 10031		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
76	<i>Klebsiella</i>	<i>oxytoca</i>	ATCC 43165		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
77	<i>Listeria</i>	<i>innocua</i>	QL 32811.1		Seasoning Powder	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
78	<i>Listeria</i>	<i>ivanovii</i>	QL 03091 1-11		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
79	Listeria	monocytogenes	QL 03091 1.10		Shellfish	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
80	Morganella	morganii	ATCC 25829		Human	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
81	Proteus	mirabilis	QL 11007.6		Veterinary	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
82	Proteus	vulgaris	ATCC 6380		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation			
83	<i>Providencia</i>	<i>stuartii</i>	QL 11007.5		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
84	<i>Pseudomonas</i>	<i>extremorientalis</i>	QL 17041.1		Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
85	<i>Pseudomonas</i>	<i>gessardii</i>	QL 17041.12		Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
86	<i>Pseudomonas</i>	<i>proteolytica</i>	QL 17041.4		Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation			
87	<i>Pseudomonas</i>	<i>rhodesiae</i>	QL 17041.6		Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
88	<i>Pseudomonas</i>	<i>aeruginosa</i>	ATCC 35032		Not Available	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
89	<i>Pseudomonas</i>	<i>fluorescens</i>	QL 17041.3		Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
90	<i>Serratia</i>	<i>marcescens</i>	QL 11007.1		Bottled Water	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
N	Species	Sub species	Refere nce	rDNA sequencing	Origin	La b		LT/SH MALDI-MS System														
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Fi na l re su lt (+/-)	Altern ative meth od confir matio n compared to identit y of the strain	Final Interpre tatio n	Fin al res ult (+/-)	Alternati ve method confirm ation compar ed to identity of the strain	Final Interpre tation	Fin al res ult (+/-)	Alternati ve method confirm ation compar ed to identity of the strain	Final Interpre tation	Fin al res ult (+/-)	Alternati ve method confirm ation compar ed to identity of the strain	Final Interpre tation			
91	<i>Siccibacter</i>	<i>turicensis</i>	QL 17031. 7		Infant formula	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
92	<i>Salmonella</i>	Agona	ATCC 51957		Not Available	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
93	<i>Salmonella</i>	Typhi	QL 16078- 2A		Chicken	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
94	<i>Salmonella</i>	California	NCTC 6018		Not Available	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)

Optimal growth																						
EXCLUSIVITY																						
Non-Target Strain								Alternative method														
LT/SH MALDI-MS System																						
CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar										
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		Reference method	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain
95	Staphylococcus	aureus	QL 03091 1.4		Seasoning Powder	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
96	Staphylococcus	epidermidis	QL 03091 1.1		Bottled Water	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
97	Staphylococcus	haemolyticus	QL 03091 1.2		Veterinary	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
98	Staphylococcus	chromogenes	QL 17041.10		Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

Optimal growth																							
EXCLUSIVITY																							
Non-Target Strain								Alternative method															
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System															
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar			
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	
99	<i>Staphylococcus</i>	<i>sciuri</i>	QL 17041.9		Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
100	<i>Streptococcus</i>	<i>pneumoniae</i>	ATCC 19615		Pharynx of Child	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
101	<i>Vibrio</i>	<i>vulnificus</i>	QL021 111A		Seafood Product	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	
Total	N								101				101				101				101		
	PA								0				0				0				0		
	ND								0				0				0				0		

Optimal growth																									
EXCLUSIVITY																									
Non-Target Strain								Alternative method																	
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		LT/SH MALDI-MS System																	
								CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar					
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation						
	NA							101				12				11				4			9		
	NA (no growth)							0				89				90				97			92		
	PD							0				0				0				0			0		
	No result							0				0				0				0			0		

EXCLUSIVITY																						
Non-target strain							Reference method	Alternative method														
								LT/SH MALDI-smart System														
N	Species	Sub species	Reference	rDNA sequencing	Origin	Lab		CBA			mCCDA			CampyFood Agar			RAPID'Campylobacter			Campy Cefex Agar		
								Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation	Final result (+/-)	Alternative method confirmation compared to identity of the strain	Final Interpretation
1	<i>Acinetobacter</i>	<i>baumanii</i>	Ad1090		Hemoglobin	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
2	<i>Acinetobacter</i>	<i>calcoaceticus</i>	Ad1092		Poultry meat	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
3	<i>Acinetobacter</i>	<i>calcoaceticus</i> var. <i>anitrat</i>	1		Poultry	Adria	-	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

4	<i>Acinetobacter</i>	<i>johnsonii</i>	Ad1317		Whole egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
5	<i>Aeromonas</i>	<i>allosaccharophila</i>	Ad1318		Whole egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
6	<i>Aeromonas</i>	<i>hydrophila</i>	Ad1570		Water	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
7	<i>Aeromonas</i>	<i>punctata</i>	Ad1517		Whole liquid egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
8	<i>Aeromonas</i>	<i>salmonicida</i>	Ad1319		Whole liquid egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
9	<i>Alcaligenes</i>	<i>faecalis</i>	ATCC 8750		Unknown	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
10	<i>Arcobacter</i>	<i>butzleri</i>	Ad1126		Chicken	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

11	<i>Arcobacter</i>	<i>butzleri</i>	Ad1881		Environmental sample	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA	-	NA	EA	-	NA	EA
12	<i>Arcobacter</i>	<i>cryaerophilus</i>	Ad1124		Chicken	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
13	<i>Burkholderia</i>	<i>vietnamiensis</i>	Ad1538		Floor	Adria	-	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
14	<i>Burkholderia</i>	<i>cepacia</i>	Ad1541		Floor	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
15	<i>Candida</i>	<i>zeylanoides</i>	Ad2241		Process water	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
16	<i>Carnobacterium</i>	<i>maltaromaticum</i>	Ad1321		Liquid egg product	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
17	<i>Comamonas</i>	<i>aquatica</i>	Ad1543		Water	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

18	<i>Chryseobacterium</i>	<i>indologenes</i>	26		Whole egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
19	<i>Chryseobacterium</i>	<i>ureilyticum</i>	Ad1340		Whole egg	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
20	<i>Enterobacter</i>	<i>intermedius</i>	88a	<i>Raoultella planticola / ornithinolytica</i>	Gizzard	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
21	<i>Escherichia</i>	<i>fergusonii</i>	2876		Environmental sample	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
22	<i>Enterococcus</i>	<i>durans</i>	Ad148	<i>Enterococcus durans</i>	Ham	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
23	<i>Enterococcus</i>	<i>faecalis</i>	25		Chicken meat	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
24	<i>Enterococcus</i>	<i>faecium</i>	Ad1883		Turkey skin	Adria	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

25	<i>Enterococcus</i>	<i>gallinarum</i>	Ad1885		Poultry	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
26	<i>Escherichia</i>	<i>coli</i>	Ad1915		Chicken legs	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
27	<i>Escherichia</i> **	<i>fergusonii</i>	Ad1381		Water	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
28	<i>Escherichia</i>	<i>coli</i>	CIP103982		Unknown	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
29	<i>Hafnia</i>	<i>alvei</i>	168		Duck meat	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
30	<i>Klebsiella</i>	<i>oxytoca</i>	Ad1509		Milk powder	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
31	<i>Klebsiella</i>	<i>pneumoniae</i>	47		Chicken skin	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

32	<i>Lactobacillus</i>	<i>sakei</i>	85L905		Meat	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
33	<i>Myroides</i>	<i>odoratimimus</i>	Ad1341		Whole egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
34	<i>Ochrobactrum</i>	<i>pseudintermedium</i>	Ad1057		Turkey skin	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
35	<i>Photobacterium</i>	<i>phosphoreum</i>	Ad1506		Salmon	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
36	<i>Plesiomonas</i>	<i>shigelloides</i>	Ad673		Fish	Adria	-	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA
37	<i>Providencia</i>	<i>stuartii</i>	46		Poultry meat	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
38	<i>Pseudomonas</i>	<i>fragi</i>	Ad1327		Whole egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

39	<i>Pseudomonas</i>	<i>otitidis</i>	Ad1880		Skin meat	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)
40	<i>Pseudomonas</i>	<i>putida</i>	Ad1331		Whole liquid egg	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
41	<i>Psychrobacter</i>	<i>psychrophilus</i>	Ad1343		Whole liquid egg	Adria	-	NOIP	No result	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
42	<i>Ralstonia</i>	<i>mannitolilytica</i>	Ad1059		Turkey skin	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	-	NA	EA	-	NA	EA
43	<i>Ralstonia</i>	<i>mannitolilytica</i>	Ad2504		Buffer solution	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
44	<i>Rhizobium</i>	<i>radiobacter</i>	Ad1550		Water	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
45	<i>Serratia</i>	<i>liquefaciens</i>	87a		Gizzard	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

46	<i>Shewanella</i>	<i>putrefasciens</i>	EN15/34	<i>Shigella flexneri / sonnei; Escherichia** fergusonii</i>	Trout	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
47	<i>Staphylococcus</i>	<i>aureus</i>	Ad906		Sausage	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
48	<i>Trichosporon</i>	<i>asahii</i>	Ad2732		fromage frais	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
49	<i>Vibrio</i>	<i>alginolyticus</i>	Ad1888		Olster	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
50	<i>Vibrio</i>	<i>cholerae</i>	Ad1887		Fish filet	Adria	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
51	<i>Yersinia</i>	<i>enterocolitica</i>	A00C066		Cockerel	Adria	-	-	NA	EA	-	NA	EA	-	NA	EA	-	NA	EA	-	NA	EA
52	<i>Aeromonas</i>	<i>hydrophila</i>	ATCC 49140		Clinical Isolate	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

53	<i>Aerococcus</i>	<i>viridans</i>	QL 17041.8		Raw Milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
54	<i>Citrobacter</i>	<i>amalonaticus</i>	ATCC 25407		Feces	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
55	<i>Citrobacter</i>	<i>braakii</i>	ATCC 43162		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
56	<i>Citrobacter</i>	<i>farmeri</i>	ATCC 51633		Human Feces	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
57	<i>Citrobacter</i>	<i>freundii</i>	QL 11007.10		Clinical Isolate	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
58	<i>Citrobacter</i>	<i>youngae</i>	ATCC 11102		Not Available	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
59	<i>Cronobacter</i>	<i>malonaticus</i>	QL 17031.3		Infant formula	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

60	<i>Cronobacter</i>	<i>sakazakii</i>	QL 17031.4		Infant formula	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
61	<i>Cronobacter</i>	<i>sakazakii</i>	ATCC 51329		Rice flour	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
62	<i>Edwardsiella</i>	<i>tarda</i>	QL 11007.11		Clinical Isolate	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
63	<i>Enterobacter</i>	<i>aerogenes</i>	ATCC 35029		Not Available	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
64	<i>Enterobacter</i>	<i>amnigenus</i>	ATCC 51816		Milk	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
65	<i>Enterobacter</i>	<i>cancerogenus</i>	QL110 10-1	<i>Shigella, Escherichia, Enterobacter</i>	Bottled Water	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
66	<i>Enterobacter</i>	<i>cloacae</i>	ATCC 23355		Spinal Fluid	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

67	<i>Enterococcus</i>	<i>faecalis</i>	ATCC 51299		Peritoneal fluid	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
68	<i>Enterococcus</i>	<i>faecium</i>	ATCC 8459		Dairy Product	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
69	<i>Escherichia</i>	<i>fergusonii</i>	ATCC 35470	<i>Escherichia fergussonii</i>	Not Available	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
70	<i>Escherichia</i>	<i>hermannii</i>	ATCC 33651		Human Arm Wound	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
71	<i>Escherichia coli</i>	<i>coli</i> O157:H7	ATCC 43895		Raw Hamburger	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
72	<i>Escherichia</i>	<i>coli</i>	QL 11010.2		Bottled Water	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
73	<i>Escherichia</i>	<i>vulneris</i>	ATCC 29943	<i>Enterobacter</i>	Human Wound	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

74	<i>Havnia</i>	<i>alvei</i>	ATCC 51815		Milk	Q La bs	-	-	NA	EA	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
75	<i>Klebsiella</i>	<i>pneumoniae</i>	ATCC 10031		Clinical Isolate	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
76	<i>Klebsiella</i>	<i>oxytoca</i>	ATCC 43165		Clinical Isolate	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
77	<i>Listeria</i>	<i>innocua</i>	QL 32811.1		Seasoning Powder	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
78	<i>Listeria</i>	<i>ivanovii</i>	QL 03091 1-11		Clinical Isolate	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
79	<i>Listeria</i>	<i>monocytogenes</i>	QL 03091 1.10		Shellfish	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)
80	<i>Morganella</i>	<i>morganii</i>	ATCC 25829		Human	Q La bs	-	-	NA	EA	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)	- (no gro wth)	NA (no growth)	NA (no growth)

81	<i>Proteus</i>	<i>mirabilis</i>	QL 11007.6		Veterinary	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
82	<i>Proteus</i>	<i>vulgaris</i>	ATCC 6380		Clinical Isolate	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
83	<i>Providencia</i>	<i>stuartii</i>	QL 11007.5		Clinical Isolate	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
84	<i>Pseudomonas</i>	<i>extremorientalis</i>	QL 17041.1		Raw milk	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
85	<i>Pseudomonas</i>	<i>gessardii</i>	QL 17041.12		Raw milk	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
86	<i>Pseudomonas</i>	<i>proteolytica</i>	QL 17041.4		Raw milk	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
87	<i>Pseudomonas</i>	<i>rhodesiae</i>	QL 17041.6		Raw milk	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

88	<i>Pseudomonas</i>	<i>aeruginosa</i>	ATCC 35032		Not Available	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
89	<i>Pseudomonas</i>	<i>fluorescens</i>	QL 17041.3		Raw milk	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
90	<i>Serratia</i>	<i>marcescens</i>	QL 11007.1		Bottled Water	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
91	<i>Siccibacter</i>	<i>turicensis</i>	QL 17031.7		Infant formula	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
92	<i>Salmonella</i>	Agona	ATCC 51957		Not Available	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
93	<i>Salmonella</i>	Typhi	QL 16078-2A		Chicken	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)
94	<i>Salmonella</i>	California	NCTC 6018		Not Available	Q Labs	-	-	NA	EA	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)	- (no growth)	NA (no growth)	NA (no growth)

95	<i>Staphylococcus aureus</i>	QL 03091 1.4	Seasoning Powder	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
96	<i>Staphylococcus epidermidis</i>	QL 03091 1.1	Bottled Water	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
97	<i>Staphylococcus haemolyticus</i>	QL 03091 1.2	Veterinary	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
98	<i>Staphylococcus chromogenes</i>	QL 17041.10	Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
99	<i>Staphylococcus sciuri</i>	QL 17041.9	Raw milk	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
100	<i>Streptococcus pneumoniae</i>	ATCC 19615	Pharynx of Child	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)
101	<i>Vibrio vulnificus</i>	QL021 111A	Seafood Product	Q Labs	-	-	NA	EA	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)	(no growth)	NA (no growth)	NA (no growth)

T o t a l	N			101		101		101		101		101
	PA			0		0		0		0		0
	ND			0		0		0		0		0
	NA			100		12		11		4		9
	NA (no growth)			0		89		90		97		92
	PD			0		0		0		0		0
	No result			1		0		0		0		0

Annex J - Detailed list of participants

- ☐ William Glover, Raymond Gee, Michael Tran – Washington DOH - Washington
- ☐ Paul Park – California Department of Public Health – Richmond, CA
- ☐ Amanda Harrington, Kathleen McKinley, Samuel Collier – Loyola (GS) – Maywood, IL
- ☐ Ryan Jepson, Jennifer Elwood, and Valerie Reeb – Iowa SLH (JS) – Coralville, IA
- ☐ Paula Snippes and Melissa Hargreaves – Minnesota Department of Public Health (JS) – St. Paul, MN
- ☐ David Simon, Alexander Nemethy, Melba Marshall, and Shipra Mohan – Florida Department of Agriculture and Consumer Services – Kissimmee, Florida
- ☐ Durda Slavic – Animal Health Laboratory: University of Guelph – Ontario, Canada
- ☐ Dustin Loy – University of Nebraska-Lincoln: Vet Diagnostic Center – Lincoln, Nebraska

**Annex B - Raw data generated during the Bruker MADLI Biotyper
Inter-Laboratory Study for Campylobacter**

*See attached Excel spread sheet: ILS Campylobacter Results
(Bruker Campylobacter Raw Data 2017LR74 (QLAB))*